

Front cover

In vitro image of astrocytes (green) and microglia (red).
Cell nuclei are shown in blue.

Credit: Khalil Rawji (Franklin lab)



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Wellcome and the Medical Research Council.



UNIVERSITY OF
CAMBRIDGE



Medical
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About Us

The Wellcome - MRC Cambridge Stem Cell Institute is a world-leading research centre with a mission to transform human health through a deep understanding of stem cell biology.

The Institute is core funded by Wellcome and the Medical Research Council to study the fundamentals of stem cell biology in health and disease, and to translate our research findings from bench to bedside to deliver tangible patient benefit.

We currently have 28 research groups with over 300 biological, clinical and physical scientists operating across a wide range of tissues and at multiple scales. This arrangement allows commonalities and differences in stem cell biology to be explored in a cohesive and interdisciplinary manner.

As the hub of a wider stem cell community across the University of Cambridge and neighbouring Institutes, our scientists have developed close collaborations with a wide range of researchers across disciplines.

The Institute currently has 35 affiliated researchers across 15 University departments, as well as the Babraham Institute, Microsoft Research, Wellcome Sanger Institute, MRC Laboratory of Molecular Biology and EMBL- European Bioinformatics Institute.

Institute research aims to have direct and tangible impacts on society. Our dedicated public engagement team engage groups on a local, national and international level through dedicated events and resources, co-created with charity and research partners. This ensures our scientists are involved in wider conversations about stem cells in Cambridge and beyond.

As well as delivering outstanding research, the Institute is committed to providing a diverse and inclusive working environment for all staff and students. In line with central University activities, the Institute promotes and delivers a range of social and wellbeing initiatives.

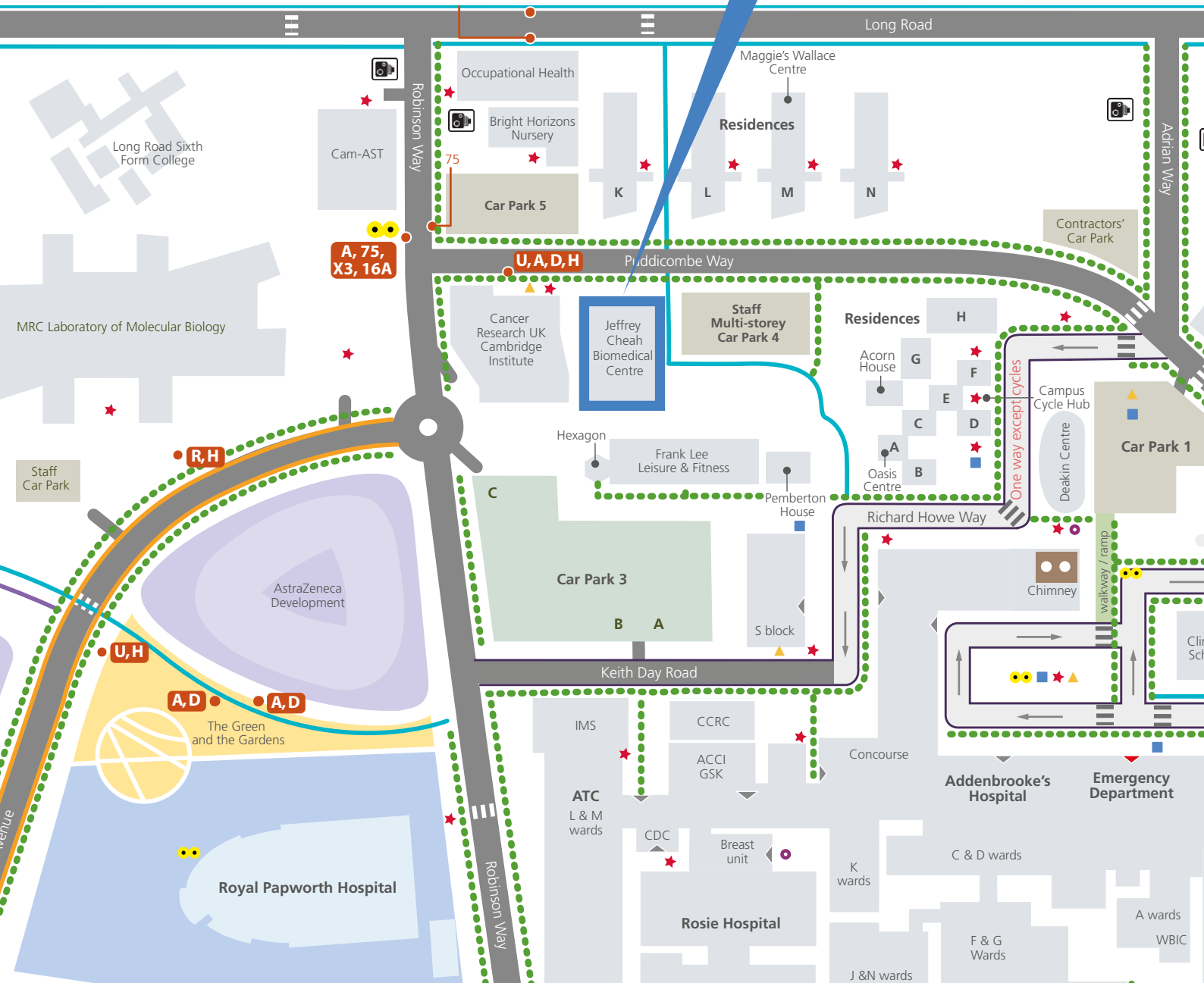
In September 2019, the Institute moved to its new home on the Cambridge Biomedical Campus, following extensive work on all levels to prepare for the relocation.

The move represents a major milestone for the Institute and groups from seven different buildings are now co-located in one building.





We are located in the
Jeffrey Cheah Biomedical Centre



Institute Research

Research at the Institute falls under three key themes: Stem Cell States, Stem Cells in Disease, and Stem Cells and Therapeutics. Many of our scientists contribute to more than one theme, and within these themes we have particular strengths in pluripotency, haematopoiesis, neural and epithelial stem cells.

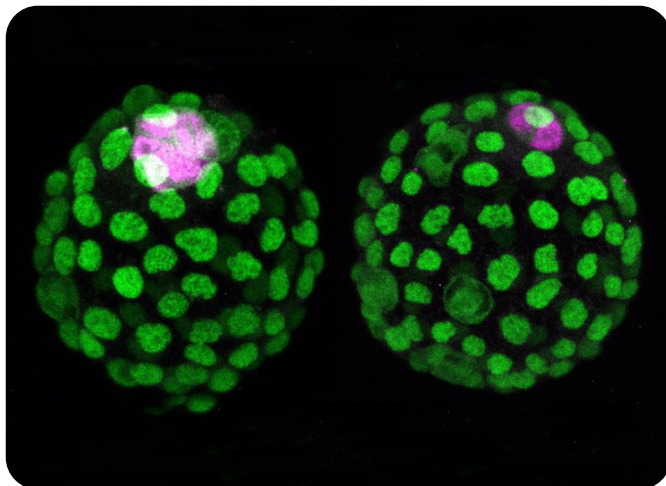
Stem Cell States

Stem cells have the extraordinary ability to develop into any type of cell in the body. We study the fundamentals of stem cell biology to understand the mechanisms by which they self-renew, maintain their states and commit to differentiate into all the cell types of the body.

Establishing new understanding of stem cell biology and behaviour complements and informs our studies of stem cell dysfunction in disease and provides the foundations of our translational aspirations for stem cells and therapeutics.

Group Leaders working on this theme:

M. Alcolea, S. Basu, K. Chalut, A. Cvejic, R. Franklin, B. Göttgens, T. Green, B. Hendrich, T. Káradóttir, E. Laurenti, J.-H. Lee, S. Méndez-Ferrer, J. Nichols, A. Philpott, D. Rowitch, J. Silva, B. Simons, S. Sinha, A. Smith & L. Vallier.



Hendrich Lab

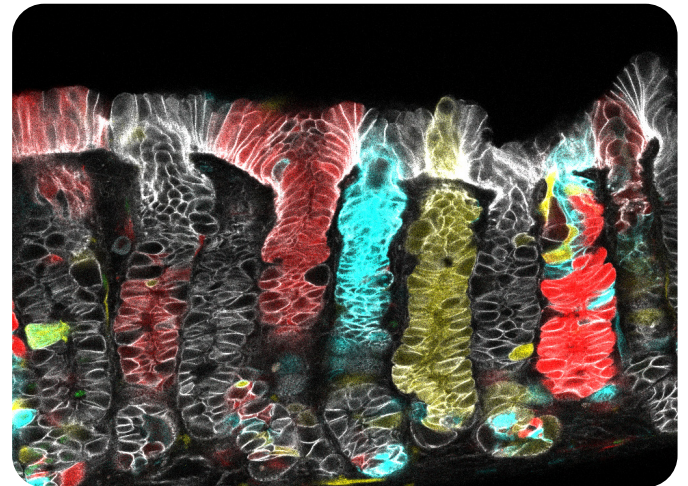
Stem Cells in Disease

Stem cell dysfunction underlies a range of diseases and health challenges that face the global population today. From neurodegenerative and cardiovascular diseases to cancer and ageing, stem cell dysregulation is implicated across the disease spectrum.

Underpinned by our exploration of normal stem cell states, we are investigating the mechanisms responsible for pathological behaviours of stem and progenitor cells. Our researchers focus particularly on different cancer pathophysiology and regenerative failure.

Group Leaders working on this theme:

M. Alcolea, S. Buczacki, K. Chalut, R. Franklin, B. Göttgens, T. Green, D. Hodson, B. Huntly, T. Káradóttir, E. Laurenti, J.-H. Lee, S. Méndez-Ferrer, A. Philpott, I. Ringshausen, D. Rowitch, B. Simons & G. Vassiliou.



Simons Lab

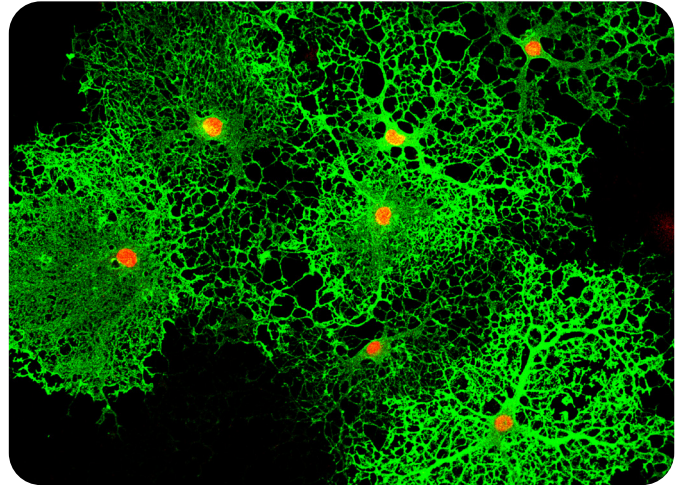
Stem Cells & Therapeutics

Building on the research undertaken into stem cell states and stem cells in disease, Institute researchers are using stem cells to model diseases *in vitro* and to generate new diagnostic and therapeutic approaches to deliver patient benefit.

Several investigators are developing first-in-human clinical trials of cellular therapies using stem cell derivatives, while others work on new diagnostic and prognostic approaches to improve patient outcomes.

Group Leaders working on this theme:

R. Barker, R. Franklin, C. Ghevaert, T. Green, B. Huntly, A. McCaskie, S. Méndez-Ferrer, D. Rowitch, S. Sinha & L. Vallier, G. Vassiliou.



Franklin Lab

Core Facilities

Alongside state-of-the-art laboratories, Institute researchers also benefit from a range of core facilities located within the Jeffrey Cheah Biomedical Centre. These facilities include Bioinformatics, Flow Cytometry, Next Generation Sequencing, Imaging, Histology, Biofacilities and Tissue Culture.

Highly-skilled facility staff provide key services and training to researchers throughout the Cambridge Stem Cell Institute, as well as to affiliated researchers and the wider University.

Institute members also have access to the NIHR Cambridge BRC Phenotyping Hub which is equipped with state-of-the-art equipment including high speed cell sorters, bench top analysers, microscopes and high content/high throughput equipment.



Credit: Chris Loades

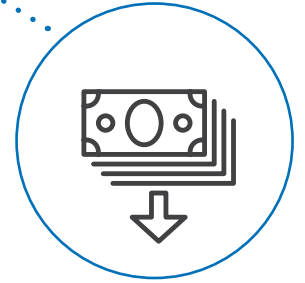
>2300

Items of equipment
moved to JCBC



£117.7M

Total grant income for
Institute group leaders



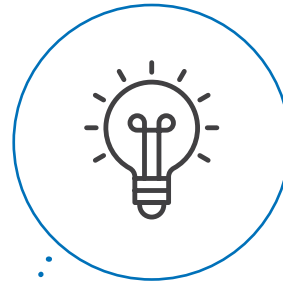
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Research groups
under one roof

Cambridge Stem Cell Institute

>1200

New Twitter followers



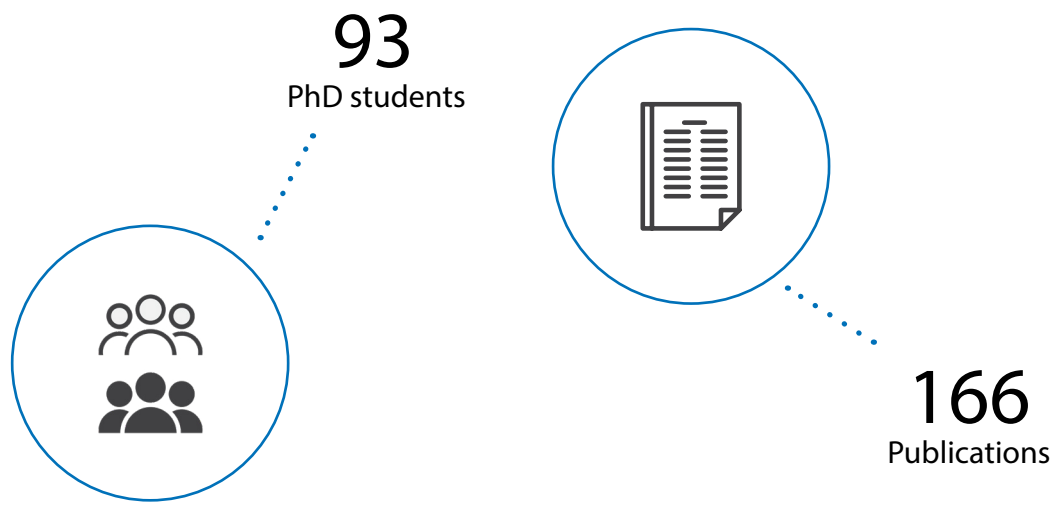
7

Patents and invention
disclosures

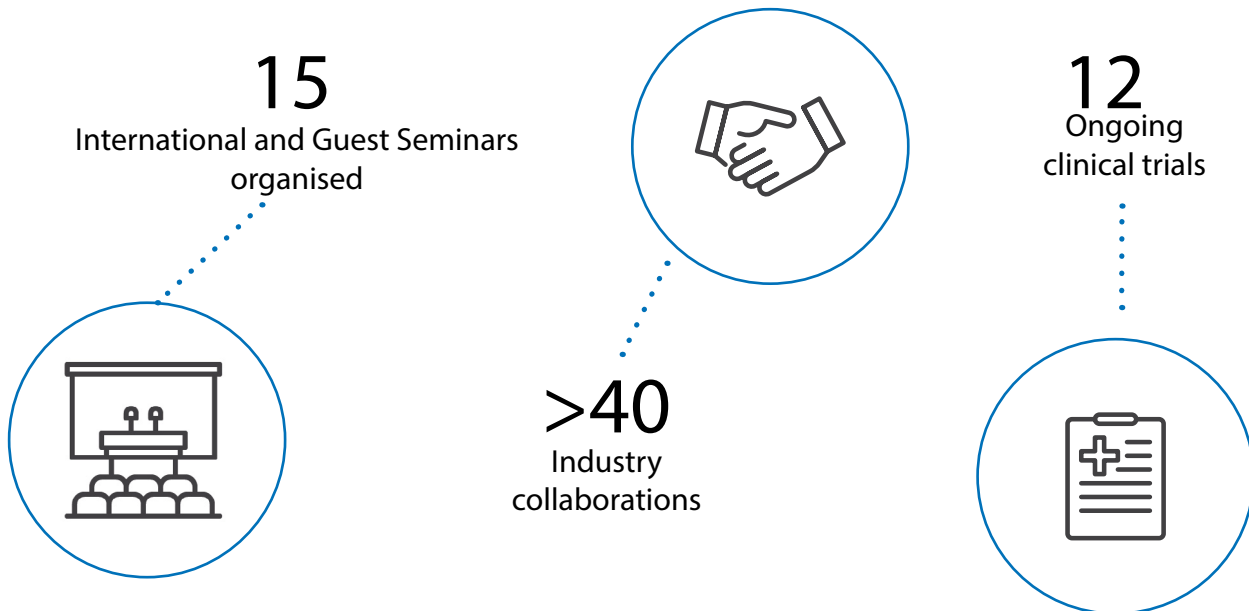
20

Public Engagement
projects and activities
organised





2019 in numbers



Relocation to the Cambridge Biomedical Campus

Following an extension of the building completion, the Institute moved to its new home on the Cambridge Biomedical Campus in September 2019.

Over the last three years, work was carried out at all levels of the Institute to prepare for the relocation. Working groups were established to facilitate the smooth transition and a full HR consultation process with staff took place, resulting in a new administrative infrastructure to support the Institute.

Over 3 months, a complex moving plan was implemented, with 29 research groups, 7 core facilities, and over 2300 items of equipment, from 7 different buildings spread across Cambridge moving to the Jeffrey Cheah Biomedical Centre.

The relocation has already facilitated cross institute interactions and a programme of weekly internal seminars has been launched for PhD students and postdocs to present their work to the Institute community.

There are also several activities, social and research-related, jointly organised with our neighbours at CRUK Cambridge Institute. Further collaborative events and initiatives are also planned for the future.

In addition to the Cambridge Stem Cell Institute, the Jeffrey Cheah Biomedical Centre is also home to the Cambridge Institute of Therapeutic Immunology & Infectious Disease and the Milner Therapeutics Institute.

Stem Cell Institute Research groups are located on three floors of the building, with core facilities supporting the building's researchers located on the ground floor and in the basement.



Jeffrey Cheah
Biomedical Centre

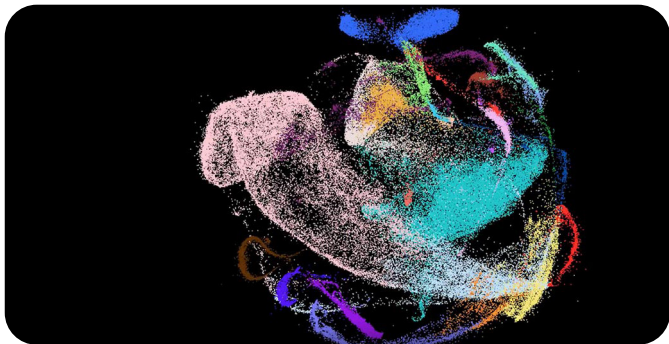
Jeffrey Cheah
Biomedical Centre



Publication of Transformative Discoveries

Elisa Laurenti co-authored a study where scientists created the first human developmental liver cell atlas, providing crucial insights into how the blood and immune systems develop in the foetus. It is a comprehensive, high-resolution resource that improves our understanding of normal development and assists medical efforts to tackle diseases, such as leukaemia and immune disorders. *Image credit: Newcastle University*

Popescu D-M, Botting RA, Stephenson E, ..., **Laurenti E**, Teichmann SA, Haniffa M. Decoding human fetal liver haematopoiesis. **Nature**, 574, 365–371(2019).



David Rowitch co-led two new studies reporting progress in using stem cells to develop new therapies for Pelizaeus-Merzbacher disease (PMD), a rare genetic condition affecting boys that can be fatal before 10 years of age.

Nobuta H, Yang N, Ng YH, ..., **Franklin RJM**, **Rowitch DH**, Wernig M. Oligodendrocyte Death in Pelizaeus-Merzbacher Disease Is Rescued by Iron Chelation. **Cell Stem Cell**, 25(4):531-541.e6. & Gupta N, Henry RG, Kang SM, ..., Huhn SL, Barkovich AJ, **Rowitch DH**. Long-Term Safety, Immunologic Response, and Imaging Outcomes following Neural Stem Cell Transplantation for Pelizaeus-Merzbacher Disease. **Stem Cell Reports**. 13;13(2):254-261.

Robin Franklin led new research suggesting that metformin (a common diabetes drug used worldwide to treat diabetes) could hold the key to stopping multiple sclerosis, through its ability to restore cells to a younger, healthier state.

Metformin Restores CNS Remyelination Capacity by Rejuvenating Aged Stem Cells. Neumann B, Baror, Zhao C, ..., **Chalut K**, van Wijngaarden P, **Franklin RJM**. **Cell Stem Cell**, 25, (4), 473-485.e8.



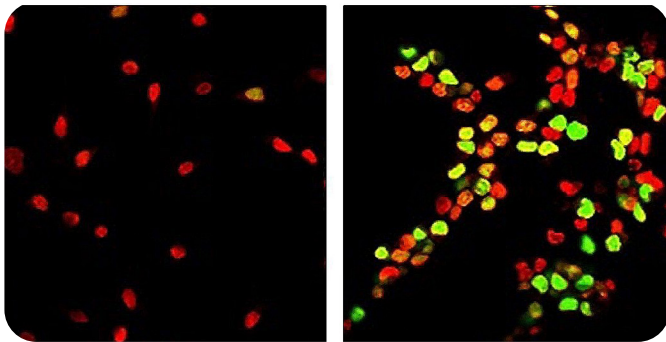
Jose Silva revealed for the first time that cells can actually follow a range of distinct molecular routes in order to reach the same final cell type, demonstrating a surprising flexibility in cell decision making during cell fate change.

Stuart HT, Stirparo GG, Lohoff T, ..., **Nichols J**, **Göttgens B**, **Silva JCR**. Distinct Molecular Trajectories Converge to Induce Naive Pluripotency. **Cell Stem Cell**, 25(3):388-406.e8.

Institute researchers produced 166 publications in 2019, including 112 peer-reviewed primary research reports. Particular highlights are listed below, with a full list of 2019 Institute publications included on page 78.

Kevin Chalut and **Robin Franklin** led a multi-disciplinary team that showed how increasing brain stiffness as we age causes brain stem cell dysfunction, and demonstrates new ways to reverse older stem cells to a younger, healthier state. The results could help develop much-needed treatments for age-related brain diseases.

Segel M, Neumann B, Hill MFE, ..., Franze K, **Franklin RJM**, **Chalut KJ**. Niche stiffness underlies the ageing of central nervous system progenitor cells. **Nature**, 573(7772):130-134.



Bertie Göttgens co-authored multi-disciplinary research studying the genetic activity of over 100,000 embryonic cells to establish the molecular blueprint of early embryo development. This new insight provides fundamentally important information on how mammalian embryos develop during gastrulation, a key stage of development, and paves the way for new understanding of the earliest stages of life.

Pijuan-Sala B, Griffiths JA, Guibentif C, ..., **Nichols J**, Marioni JC, **Göttgens B**. A single-cell molecular map of mouse gastrulation and early organogenesis. **Nature**, 566(7745):490-495.

Sanjay Sinha and his team found that, by transplanting an area of damaged tissue with a combination of both heart muscle cells and supportive cells taken from the outer layer of the heart wall, they may be able to help the organs recover from the damage caused by a heart attack.

Bargehr J, Ong LP, Colzani M, ..., Bennett MR, Murry CE, **Sinha S**. Epicardial cells derived from human embryonic stem cells augment cardiomyocyte-driven heart regeneration. **Nature Biotechnology**, 37(8):895-906.



David Rowitch co-led a new study pinpointing the cell types affected in brains of multiple sclerosis patients. The research showed that projection neurons are damaged by the body's own immune cells, and that this damage could underpin the brain shrinkage and cognitive changes associated with MS. These new findings provide a platform to develop new specific MS therapies that target damaged brain cells.

Schirmer L, Velmeshev D, Holmqvist S, ..., Shioh LR, Kriegstein AR, **Rowitch DH**. Neuronal vulnerability and multilineage diversity in multiple sclerosis. **Nature**, 573(7772):75-82.

Delivery of early phase clinical trials

Institute researchers are involved in a number of active clinical trials which show promise for the translation of stem cell research from bench to bedside.

Roger Barker is involved in several clinical trials investigating new treatments for patients with Parkinson's disease and Huntington's disease, including the TRANSEURO study, an open label transplant study in Parkinson's Disease trial which treated its last patient in May 2018, with results on the primary end point expected in April 2021 (NCT01898390).

Work is ongoing on to develop a trial using human embryonic stem cell-derived midbrain dopaminergic (STEM-PD) progenitor cells. This study is pending regulatory approval, with a planned start April 2021.

Brian Huntly's dose escalation study to investigate the clinical activity of BET inhibitors in subjects with relapsed, refractory hematologic malignancies is ongoing, with primary completion date scheduled for 2020 (NCT01943851).

Simón Méndez-Ferrer's trial investigating the redeployment of tamoxifen to modulate the bone marrow niche and treat myeloproliferative neoplasms was extended in 2018 to follow up on response and durability (EudraCT 2015-005497-38).

Simón was also involved in the trial studying the Effects of Sympathomimetic Agonists on the Disease Course and Mutant Allele Burden in Patients with JAK2-mutated Myeloproliferative Neoplasms. The results for this trial were published in 2019 (NCT02311569).

David Rowitch concluded a five-year long term follow-up study of a neural stem cell therapy trial for Connatal Pelizaeus-Merzbacher Disease, with results published in 2019, in Stem Cell Reports, 13;13(2):254-261 (NCT01005004 / NCT01391637).

Andrew McCaskie is leading a trial to evaluate the efficacy of adipose derived mesenchymal stromal cells in patients with knee osteoarthritis (NCT02838069).

Robin Franklin continued the trial of Retinoid X receptor gamma agonists in multiple sclerosis patients to regulate the differentiation of central nervous system progenitors into remyelinating oligodendrocytes. All 50 people with MS on the trial have completed the treatment phase and many have come to the end of the trial. Results are expected in 2020 (EudraCT 2014-003145-99).

Cédric Ghevaert designed the recovery and survival of stem cell originated red cells (RESTORE) trial, which is due to commence patient recruitment in 2020.

Dan Hodson designed the DLBCL Interim Response Evaluation for Customised Therapy (DIRECT) trial which is currently pending regulatory approval and is expected to open in 2020.

Increased engagement with industry

Institute members are actively engaged with industry and enterprise partners as reflected by licensing agreements, collaborative projects and start-up companies. The Institute currently has over 40 active commercial and industrial collaborations. In 2019, 4 patents and 3 invention disclosures were filed.

Ludovic Vallier started a new project facilitated by the Milner Institute focusing on the identification of new therapeutics for liver disease. Working with GlaxoSmithKline and Ferring Pharmaceuticals, this project will bring together a novel in vitro human model for non-alcohol fatty liver disease/non-alcoholic steatohepatitis developed in the Vallier group and NASH/NAFLD expertise from the two companies.

Ludovic also established a spinout company, Hepatotarget Therapeutics, that will use stem cell differentiation technology to manufacture mature hepatocytes as a cell therapy for liver disease.

Ingo Ringshausen submitted a patent for Therapeutic Treatment using Protein Kinase C (PKC) Inhibitors and Cytotoxic Agents (pending).

Cedric Ghevaert has established two spinout companies based on his research. Tropofour Therapeutics is developing a compound to control platelet formation and CellAdvice Ltd is a consultancy for companies involved in transfusion products or cell therapies.



Production of skilled stem cell scientists

The Institute is committed to training the next generation of skilled and sought-after clinical and non-clinical stem cell scientists to strengthen the global academic and industrial community.

PhD Programmes

The 4-year Wellcome PhD Programme in 'Stem Cell Biology & Medicine' competitively recruited 4 new students from 174 applications in 2019. We also recruited 1 student to join the 4-year MRC PhD Programme in 'The Physical Biology of Stem Cells'. These two programmes represent the heart of a vibrant PhD student community at the Institute, with a total of 77 students as of October 2019 and a further 16 students completing their studies last year. In 2019, the PhD students co-authored 43 publications, 24 as first author.

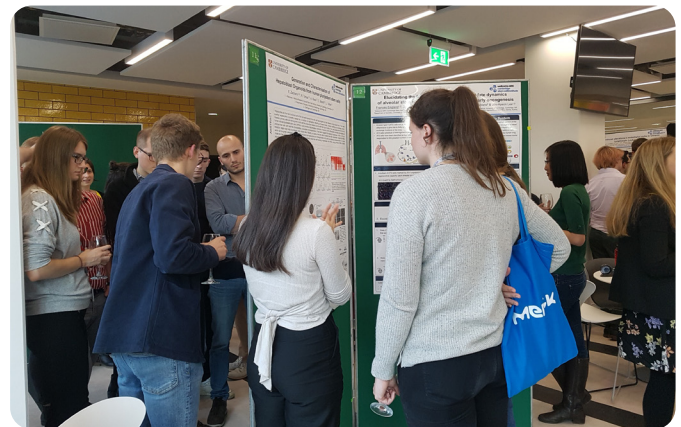
Each year the Institute runs a productive and enjoyable PhD Symposium and Away Day. In 2019, 13 PhD students presented their research projects and Sir John Gurdon gave the keynote lecture 'Half a century at the bench, and still there'. Over 40 posters were on display from students based in Institute and affiliated research groups.

The Institute has been awarded funding to extend the 4-year Wellcome PhD Programme in Stem Cell Biology and Medicine for a further 5 years.

Clinician Scientists

Approximately 40% of group leader positions at the Institute are clinically based and facilitate collaborations between basic scientists and those driven by disease-focused questions.

The Institute currently hosts 12 clinically trained PhD students and 5 clinical lecturers and clinically trained postdocs/fellows.



Accessible and authoritative voice

As a world-leading Institute, we are committed to providing an accessible and authoritative voice on a range of stem cell topics and issues.

Communications & Engagement

Researchers and the Institute communications team work together with Wellcome, the MRC and University press offices to ensure the best research from the Institute is successfully showcased via the press and on social media. The Institute Twitter account (@SCICambridge) saw an increase of over 1200 followers in 2019, with over 650K tweet impressions.

The Public Engagement team continue to deliver a diverse range of events and activities to ensure our scientists engage with a broad spectrum of public voices. These encounters serve to enhance the quality of the scientific questions we ask and help the public to build trust in stem cell research. In 2019, face-to-face public events allowed researchers to actively engage with thousands of individuals, highlights of which can be found on page 18.



Representation & Impact

Institute members hold senior positions on a number of research society boards, research funding committees (including Blood Cancer UK, Biotechnology and Biological Sciences Research Council & National Institute of Health Research) and journal editorial boards (including Blood, Development & Journal of Experimental Medicine).

In 2019, the Institute collaborated with EuroStemCell, a public-facing organisation, through contribution to the initiative #openupstemcells, a digital engagement campaign where four of our researchers produced quick and easily accessible films explaining their research in stem cells. The films are now freely available on eurostemcell.org, a hub for people looking for reliable information on stem cell therapies and current research.

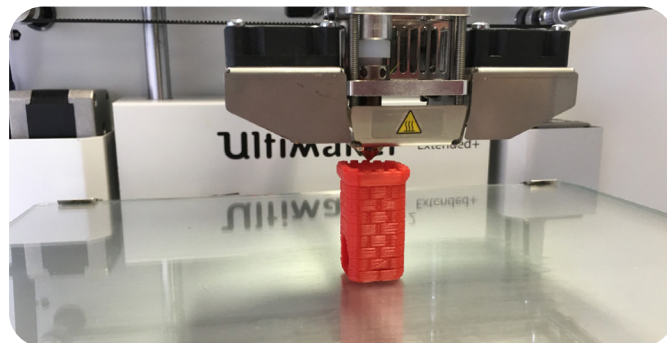


Seminars and Events

The Institute runs a range of interdisciplinary research events throughout the year to bring together the vibrant and wide-reaching stem cell community across Cambridge and beyond.

3D Printing Workshop for Stem Cell Scientists

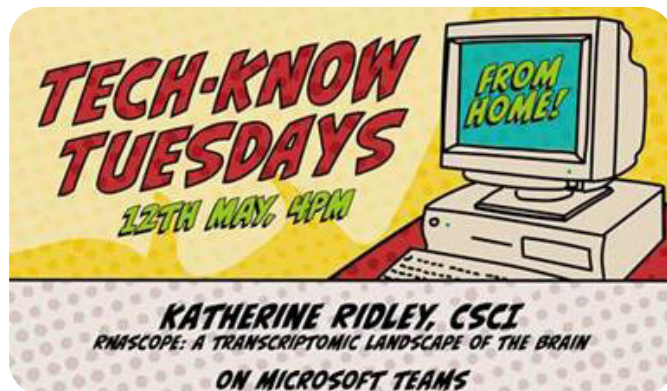
With the support of the University's interdisciplinary small grants schemes, a group of biologists and engineers from the Cambridge Stem Cell Institute ran an interactive three-day workshop to teach the stem cell community about the practical uses of 3D printing technology. The workshop was led by three members of the Kent lab and used worked examples to explore how 3D printing can be used to build practical tools to enhance research in a fast and inexpensive way. The attendees joined from 11 different University Departments in a truly cross-disciplinary event.



Techknow Tuesdays

This event, set up by institute post-docs Dan Prins (Green lab) and Craig McDonald (Kent lab), brings together researchers from across Cambridge to discuss the latest technologies and their application to stem cell research.

Talks are held monthly and are followed by a chance to network with researchers from across the Institute and other departments from the Biomedical Campus and central Cambridge.



Stem Cells & Society Seminar Series

The Institute also partnered with the Department of Sociology at the University of Cambridge to run a series of five consecutive seminars on the subject "Stem Cells and Society". The series was led by sociologists Prof Sarah Franklin, Dr Noémie Merleau-Ponty and Dr Karen Jent and was designed to offer a comprehensive introduction to the social issues raised by stem cell research.



Topics up for discussion included how the legislation covering embryo research emerged in the UK, the importance of open dialogue in the development of UK science policy and the role of media coverage and public engagement in shaping perceptions of stem cell research.

StemCells UK

Working together with The Francis Crick Institute and the Centre for Regenerative Medicine, Edinburgh, we were very pleased to organise the first Stem Cells UK meeting, on 1st October 2019. The meeting brought together stem cell researchers from across the UK sharing research ideas, forging collaborations and meeting friends old and new.

The one day event had over 100 attendees from across the UK stem cell community, with a programme of talks from leading researchers, and networking sessions.

International Seminar Series

The Institute's International Seminar Series features world-leading scientists who are invited to present their work to all Institute researchers and affiliates. Since the move to our new building, these Seminars have had a large increase in attendance. They are open to everyone and advertised widely across the Cambridge stem cell community. Seminar topic highlights in 2019 include: cortical interneuron diversity (Oscar Marin, King's College London), reproducibility in research (Glenn Begley, Biocurate, Australia), developmental lineages (Alexander Schier, Basel's Biozentrum/Harvard University), angiogenesis and vascular metabolism (Peter Carmeliet, KU Leuven).

Institute Annual Retreat

This event focusses on fostering an integrated, collaborative culture between all levels of the Cambridge Stem Cell Institute and affiliated researchers. The Retreat features talks from Institute and affiliate group leaders as well as flash talks and poster presentations from postdocs and PhD students.

The evening social event allows all attendees to relax and interact over a glass of wine or 'Regenerator', the Institute beer developed as part of a public engagement project with Moonshine Brewery.

A promotional poster for the Stem Cells UK meeting. It features a blue background with a pattern of white circles in the top left corner. A hand in a blue glove holds a petri dish containing a red liquid with a white, branching, tree-like structure. The text is white and blue.

Stem Cells UK
Tuesday 1st October - The Francis Crick Institute

Speakers:
Bertie Göttgens
Shukry Habib
Majlinda Lako
Madeline Lancaster
Joo-Hyeon Lee
Cristina Lo Celso
Donal O'Carroll
Katrin Ottersbach
Simona Parrinello
Wolf Reik
Paul Riley
Abdenour Soufi

Alexander Schier

Director of the University of Basel's Biozentrum

**"Reconstructing developmental lineages
and trajectories"**

Hosted by Ludovic Vallier

Weds 9 October, 12:30-13:30

@JCBC Lecture Theatre

Cambridge Biomedical Campus



Public Engagement

Our public engagement strategy mandates our commitment to engagement at an Institute level in support of the Centre's strategic mission.

Aim 1: Reach beyond Cambridge to raise awareness about stem cells on a national and international level



4L Rally, February

Two PhD students took to the road to engage local communities and deliver educational resources as part of a two-week adventure across the Moroccan desert.



#openupstemcells, October

Researchers answered public questions about stem cells as part of a digital engagement campaign led by EuroStemCell for Stem Cell Awareness Day.



@Seuninscience

Oluwaseun Ogundele's YouTube vlog and Instagram made a big impact in 2019, sharing personal stories to inspire young BAME students into lab science.

Aim 2: Connect to local under-served communities and empower them to access stem cell research



LifeLab, September

Institute researchers took stem cells to Ely as part of European Researchers Night, engaging 300 local families and residents on a busy Saturday at the Cathedral.



Flying in the Air, October

The Ghevaert group combined research and music for a charity concert for local churchgoers in Newmarket. Raising £1,348 for East Anglian Air Ambulance.



Stem Cell Beer

We continued our collaboration with local brewery Moonshine to create a second brew, supporting researchers to reach new faces at beer festivals and pubs.

In 2019 the Institute put its new strategy into action, delivering over 20 projects and activities for thousands of people online and in person.

Aim 3: Give patients a voice in our research and engagement activities which is valued and utilised



Art + Science at Jeffrey Cheah Biomedical Centre, December

We opened our ground floor public space with a big celebration in December in collaboration with Kettle's Yard gallery. A series of new collaborative artworks were unveiled by artists Victoria Morton, Harold Offeh and Dalziel and Scullion, including a

piece created with a local branch of the MS Society. A free-standing pinball machine will join the artworks in 2020, encouraging hands-on game play with stem cell differentiation. Looking ahead, our café and public space will be a place where public and

patient groups on the campus can learn about our science and mix with our researchers, supported by a tailored programme of public events and exhibitions.

Aim 4: Create and open and engaged research culture



Following the move to Jeffrey Cheah Biomedical Centre we set up a Lab Champion network to support 'grass-roots' interest and uptake of public engagement. The group meets informally once a month over coffee to discuss new ideas and share opportunities and training.

Dr Cedric Ghevaert has taken over as academic champion for public engagement, providing leadership through our Steering Committee.

Finally, the Institute awarded £4,750 in public engagement seed funding in 2019, to five researcher-led projects aligned to our strategy. Watch this space for what they deliver in 2020!

Principal Investigators



Maria Alcolea
Epithelial cell fate & plasticity



Roger Barker
Parkinson's & Huntington's disease



Srinjan Basu
Single-cell and single-molecule imaging



Simon Buczaccki
Colorectal cancer cell identity & tumour evolution



Kevin Chalut
Physical biology of pluripotency & differentiation



Brian Hendrich
Transcriptional control of stem cell fate



Daniel Hodson
Mutation timing in lymphomagenesis



Brian Huntly
Leukaemia stem cell biology & leukaemogenesis



Thóra Káradóttir
Neurotransmitter signalling to CNS progenitor cells



Elisa Laurenti
Human haematopoietic stem cells biology in health & disease



Ingo Ringshausen
Haematopoietic stem cells & malignancies



David Rowitch
Glial cells & response to injury



José Silva
Reprogramming and programming cell identity



Ben Simons
Stem cell fate in development, maintenance & disease



Sanjay Sinha
Vascular diseases



Ana Cvejic
Haematopoietic
stem cells



Robin Franklin
Adult neural
stem cells & CNS
regeneration



Cédric Ghevaert
In vitro production of
platelets for
transfusion



Bertie Göttgens
Decision making in
haematopoietic
stem cells



Tony Green
Haematopoiesis



Joo-Hyeon Lee
Stem cells
& niches



Andrew McCaskie
Regenerative
therapies for bone
& cartilage repair



Simón Méndez-Ferrer
Blood stem cell
niches



Jennifer Nichols
Embryonic
pluripotency



Anna Philpott
Proneural
transcription factors



Austin Smith
Stem cell potency



Ludovic Vallier
Differentiation of
pluripotent stem cells
into definitive endoderm



George Vassiliou
Leukaemic
haemopoietic
stem cells

MARIA
ALCOLEA

Epithelial cell fate and plasticity



LAB MEMBERS

Maria Bejar Serrano
Sara Bisetto
Adrien Hallou
Paula Jimenez-Gomez
Jamie McGinn
Jesus Eduardo Rojo
Greta Skrupskelyte

Our research focusses on studying the behaviour of progenitor cells in the mouse oesophagus as a model to unveil the basic rules underlying squamous epithelial cell fate. Our work in the field has revealed how this tissue is maintained under homeostatic conditions, and how these rules switch upon injury.

More recently we have been able to identify how progenitor cells alter and adapt their behaviour in response to preneoplastic mutations, reflecting their remarkable cellular plasticity. Investigating the cellular and molecular mechanisms governing this dynamic behaviour, and the potential implications for early cancer development, constitutes the basis of our ongoing research programme.

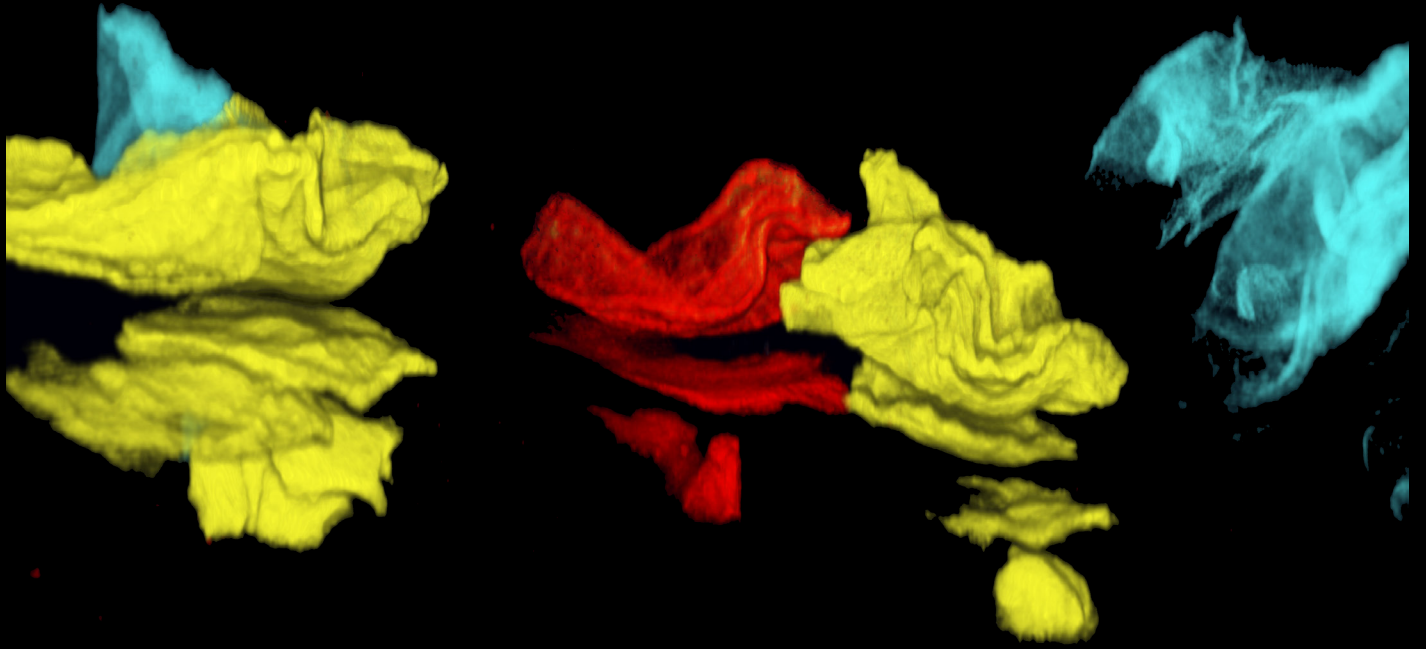
To answer these questions, we will make use of a combination of *in vivo* lineage tracing techniques, transcriptional network analysis, as well as 3D organoid and explant culture systems.

KEY PUBLICATIONS

Alcolea MP, Greulich P, Wabik A, Frede J, Simons BD, Jones PH. Differentiation imbalance in single Oesophageal progenitor cells causes clonal immortalization and field change. **Nature Cell Biology**. 2014 Jun;16(6):615-22.

Alcolea MP, Jones PH. Tracking cells in their native habitat: lineage tracing in epithelial neoplasia. **Nature Reviews Cancer**. 2013 Mar;13(3):161-71.

Doupe DP1, **Alcolea MP**, Roshan A, Zhang G, Klein AM, Simons BD, Jones PH. A single progenitor population switches behavior to maintain and repair esophageal epithelium. **Science**. 2012 Aug 31;337(6098):1091-3.



Multicolour genetic marking of oesophageal stem cells allows to track behavior at single cell resolution.

Credit: Jamie McGinn

ROGER BARKER



LAB MEMBERS

Katie Andresen
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 Danielle Daft
 Shaline Fazal
 Kate Harris
 Xiaoling He
 Sam Hewitt
 Wei-Li Kuan
 Sarah Mason
 Venkat Pisupati
 Shama Qarin
 Priya Rogers
 Sophie Skidmore
 Kelli Tornsey
 Pam Tyers
 Benjamin Vallin
 Zanna Voysey
 Alice White

Parkinson's and Huntington's disease

Our main interests revolve around two relatively common, chronic neurodegenerative disorders of the nervous system - Parkinson's disease (PD) and Huntington's disease (HD).

We are interested in better understanding how these diseases develop and then how they change over time with the idea of better classifying patients into different subtypes of disease. These subtypes can then be used to test new therapies as some types of these diseases may be better suited for one type of experimental treatment whilst others may not: e.g. dopamine cell therapies from stem cells treatment may be better suited to younger PD patients with a more benign clinical course.

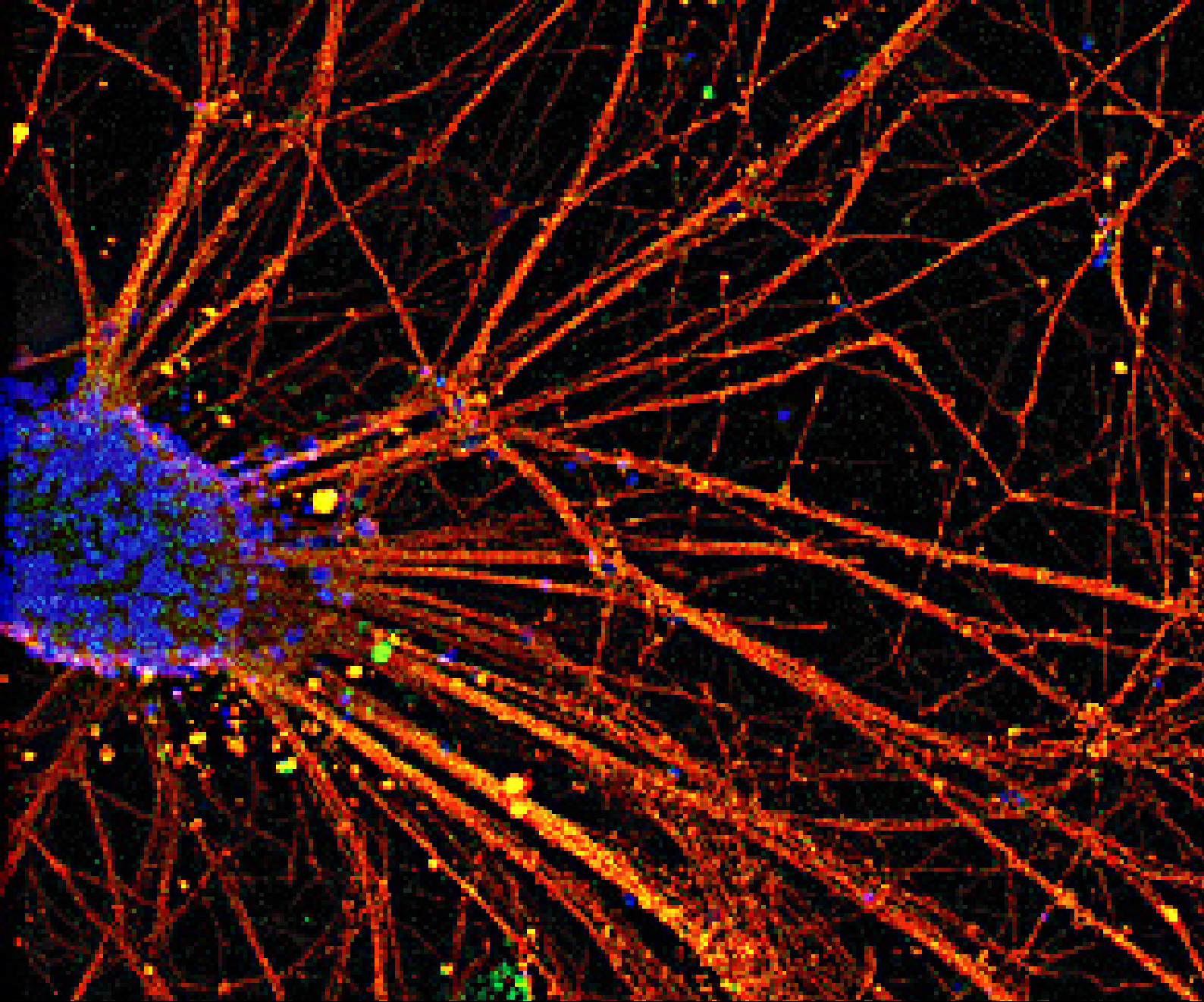
In addition, this ability to stratify patients also enables us to undertake studies looking at how these disease subtypes may arise using cells grown from the patients themselves. Typically we harvest these cells from the skin and then turn them into nerve cells in the lab, and by so doing we hope that we can recapitulate what goes wrong in the brain nerve cells in such patients.

KEY PUBLICATIONS

Barker RA; TRANSEURO consortium. Designing stem-cell-based dopamine cell replacement trials for Parkinson's disease. **Nature Medicine**, 25(7):1045-1053.

La Manno G, Gyllborg D, Codeluppi S, Nishimura K, Salto C, Zeisel A, Borm LE, Stott SR, Toledo EM, Villaescusa JC, Lönnerberg P, Ryge J, **Barker RA**, Arenas E, Linnarsson S. Molecular Diversity of Midbrain Development in Mouse, Human, and Stem Cells. **Cell**, 167(2):566-580.

Pircs K, Petri R, Madsen S, ..., Parmar M, **Barker RA**, Jakobsson J. Huntingtin Aggregation Impairs Autophagy, Leading to Argonaute-2 Accumulation and Global MicroRNA Dysregulation. **Cell Reports**, 24(6):1397-1406.



Dopaminergic neurons differentiated from human pluripotent stem cells using GMP compliant differentiation protocol—TH (green), btub (red)

Credit: Civia Chen

SRINJAN BASU

Single-cell and single-molecule imaging approaches in stem cell biology



LAB MEMBERS

Igor de Almeida
Stanley Strawbridge
Maike Steindel
Oliver Davis
Jorg Morf

The generation of diverse cell types from stem cells is a hallmark of metazoan biology, yet the mechanisms that initiate and subsequently stabilise cell type-specific gene expression programmes remain ill-defined.

Our research focuses on developing single-cell and single-molecule imaging approaches to improve understanding of the dynamics of stem cell renewal and differentiation. Specifically, we are interested in how the dynamics of signalling molecules and chromatin binding proteins regulate gene expression during stem cell fate transitions and why these pathways are often mis-regulated during early cancer progression.

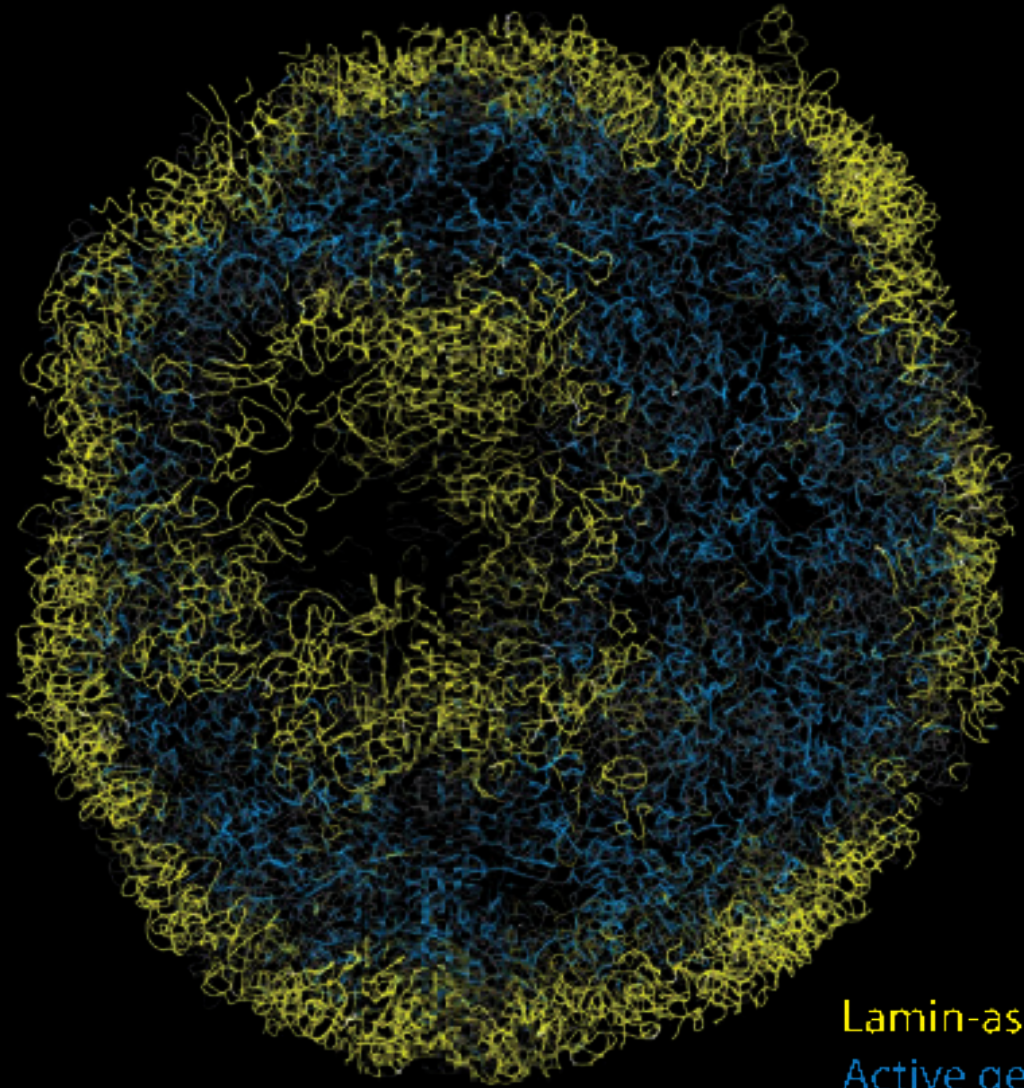
Single-cell and single-molecule approaches are key to understanding how these proteins work due to the considerable cellular and molecular heterogeneity that occurs during stem cell fate transitions. In recent years, we have developed several biophysical and computational approaches to answer these questions. For example, we have established a method combining imaging and single-cell Hi-C to study genome architecture inside individual mouse embryonic stem cells. To understand how proteins interact with each other and with chromatin, we have set up several in vitro and live-cell single-molecule imaging approaches capable of localising single proteins at <15 nm resolution and tracking them as they perform their function in live cells.

KEY PUBLICATIONS

Basu S, Needham L-M, Lando D, Taylor EJR... Eggeling C, Hendrich B, Klenerman D, Lee SF, Laue ED (2018). FRET enhanced photostability allows improved single-molecule tracking of proteins and protein complexes in live mammalian cells. **Nature Communications**. 2018 Jun 28;9(1):2520.

Stevens TJ, Lando D, **Basu S**, Atkinson LP... Wutz A, Hendrich B, Klenerman D, Laue ED (2017). 3D structure of individual mammalian genomes studied by single cell Hi-C. **Nature**. 2017 Apr 6;544(7648):59-64.

Lando D, **Basu S**, Stevens TJ, Riddell A... Lee SF, Hendrich B, Klenerman D, Laue ED (2017). Combining fluorescence imaging with Hi-C to study 3D genome architecture of the same single cell. **Nature Protocols**. 2018 May;13(5):1034-1061.



Lamin-associated
Active genes

Genome architecture of a single mouse embryonic stem cell with gene-poor lamin regions in yellow separating away from active genes in blue

Credit: Tim Stevens, MRC-LMB

SIMON BUCZACKI



LAB MEMBERS

Mahnaz Darvish Damavandi
Christopher Ward

Colorectal cancer cell identity and tumour evolution

We are interested primarily in the role sub-clonal interactions play in colorectal cancer cell identity and behaviour. We believe that the behaviour of normal intestinal cells is often analogous to that seen in oncogenically transformed cells. We therefore also study the behaviour of progenitor and differentiated cells from normal intestinal tissues to provide insights into cancer cell behaviour.

We are particularly interested in understanding the mechanisms and links behind cancer cell plasticity and identity switching. We use genetic manipulation of patient derived organoids to understand the role of competitive and co-operative interactions in tumour evolution and cellular identity.

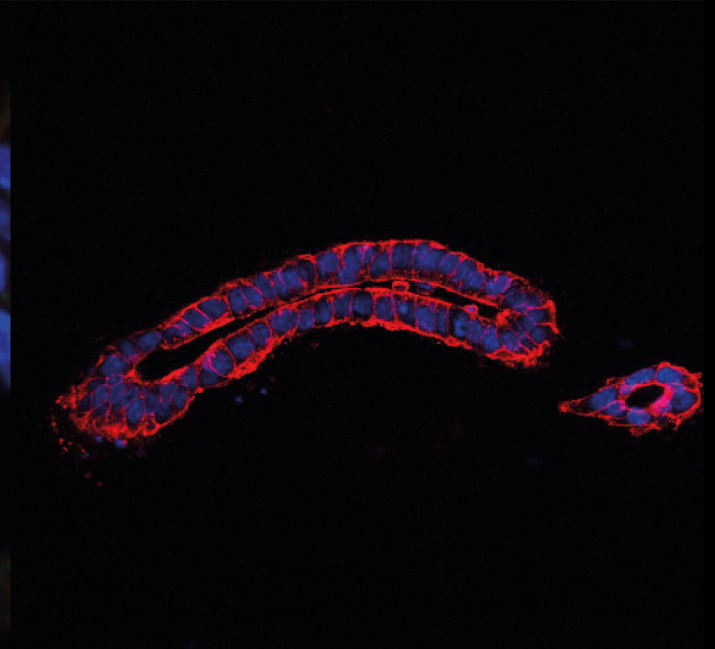
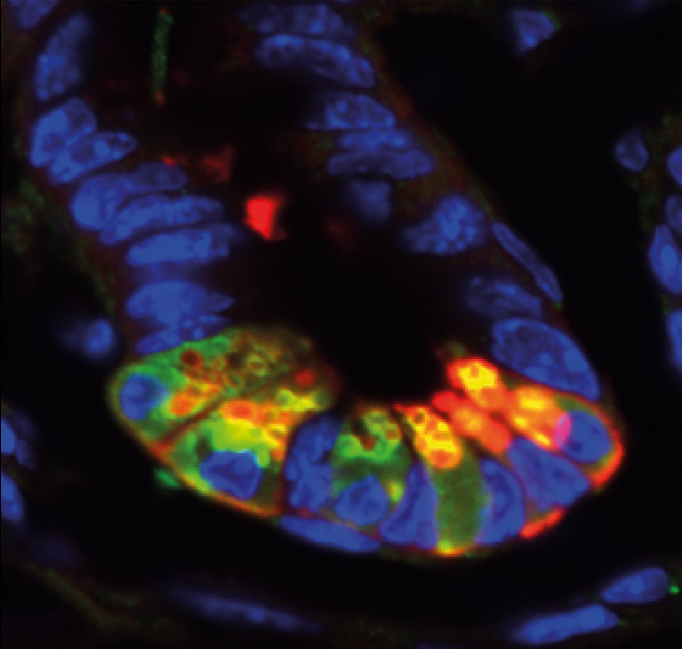
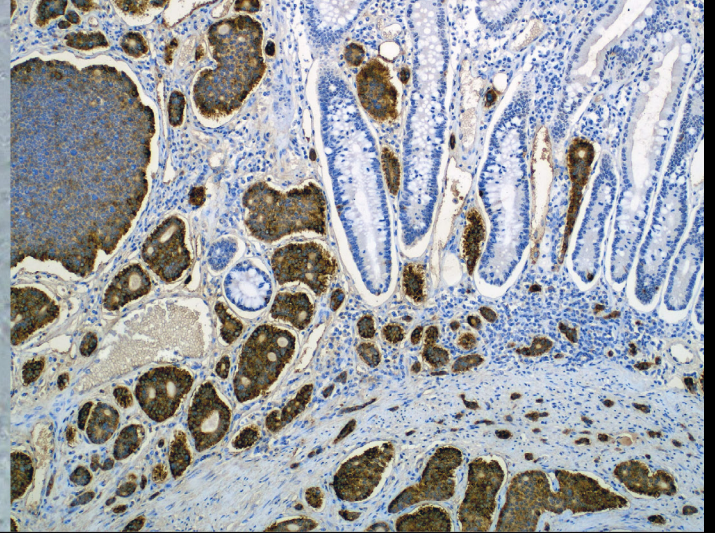
The lab is also committed to understanding the fundamental biology of small intestinal neuro-endocrine tumours through organoid biology and mouse modelling.

KEY PUBLICATIONS

Buczacki SJA, Popova S, Biggs E, Koukorava C, Buzzelli J, Vermeulen L, Hazelwood L, Francies H, Garnett MJ and Winton DJ. Itraconazole targets cell-cycle heterogeneity in colorectal cancer. **The Journal of Experimental Medicine**. 2018. Jul 2;215(7): 1891-1912.

Vermeulen L, Morrissey E, van der Heijden M, Nicholson AM, Sottoriva A, **Buczacki S**, Kemp R, Tavare S, Winton DJ. Defining stem cell dynamics in models of intestinal tumor initiation. **Science**. 2013; 342(6161): 995-998 .

Buczacki SJA, Ireland Zecchini H, Nicholson AM, Russell R, Vermeulen L, Kemp R, Winton DJ. Intestinal label-retaining cells are secretory precursors expressing Lgr5. **Nature**. 2013. Mar 7;495(7439):65-9.



A human small bowel organoid (upper left), a human intestinal neuroendocrine tumour (upper right), a mouse intestinal crypt highlighting Paneth cells (lower left) and a human colorectal cancer organoid (lower right).

Credit: Simon Buczacki

KEVIN CHALUT

The physical biology of pluripotency and differentiation



LAB MEMBERS

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Andrew Hodgson

Celine Labouesse

Carla Mulas

Sarah Pallett

Bao Xiu Tan

The transformation of a stem cell into a mature tissue cell consists of a progression of highly regulated steps, which has primarily been studied from a biochemical perspective, while mechanical aspects, despite their importance, have been largely overlooked.

We are focused on illuminating biophysical aspects of transitions between states during stem cell differentiation and embryonic development by utilising tools and concepts of physics and bioengineering alongside molecular biology.

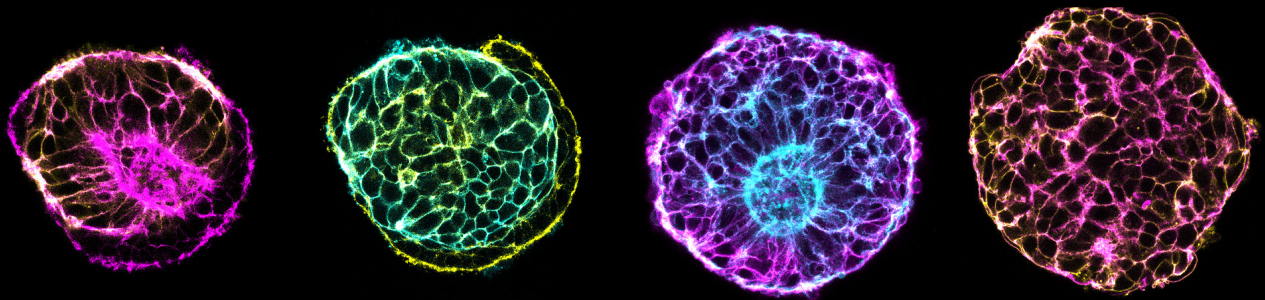
The biophysical aspects we focus on include cell mechanics and matrix signalling, as well as how nuclear mechanics influence gene expression and transport of signalling molecules through nuclear pore complexes. We are also developing single cell microfluidic techniques to study transitions between states in embryonic stem cell differentiation.

KEY PUBLICATIONS

Segel M, Neumann B, Hill MFE, Weber IP, Viscomi C, Zhao C, Young A, Agle CC, Thompson AJ, Gonzalez GA, Sharma A, Holmqvist S, Rowitch DH, Franze K, Franklin RJM, **Chalut KJ**. Niche stiffness underlies the ageing of central nervous system progenitor cells. **Nature** 573, 130–134.

Verstreken CM, Labouesse C, Agle CC, **Chalut KJ**. Embryonic stem cells become mechanoresponsive upon exit from ground state of pluripotency. **Open Biology**. 2019, 9. <http://doi.org/10.1098/rsob.180203>.

Pagliara S, Franze K, McClain C, Wylde G, Fisher C, Franklin R, Kabla A, Keyser U, **Chalut KJ**. Auxetic nuclei in embryonic stem cells exiting pluripotency. **Nature Materials**. 2014, 13, 638–644.



Two confocal slices of two human Embryonic Stem Cell colonies showing overlay of actin and EPCAM (a cell adhesion molecule) in changing colors. hES cells were grown on soft hydrogels and appear to self-organise in 3d structures. Images were taken with a 40x, oil objective on Leica SP5. Colonies are approximately 40 μ m in diameter.

Credit: Celine Labouesse/Patrick Rericha



LAB MEMBERS

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Elisa Panada
Anna Maria Ranzoni
Simone Riva
Andrea Tangherloni
Henry Xu

Blood stem cells need to both perpetuate (self-renew) themselves and differentiate into all mature blood cells to maintain blood formation throughout life. Clarifying how haematopoietic stem and progenitor cells (HSPCs) differentiate into diverse cell types is important to understand how this process is subverted in the generation of blood pathologies.

The aim of our group is to decipher how differentiation pathways of HSPCs are influenced by different microenvironments. To achieve that we use state-of-the-art single-cell RNA-seq data generation combined with computational analysis to establish principles of blood lineage differentiation. In particular, we are focusing on the dissection of the heterogeneity of cellular states in the blood system. Our research involves the use of both human samples and model organism (zebrafish, *Danio rerio*). Currently we focus our research on human foetal haematopoietic cells to reveal the dynamics and cellular programmes active during human blood development. We have also performed extensive analysis of lung cancer patient samples to investigate the influence of tumour microenvironment in the context of pathological differentiation of myeloid progenitors.

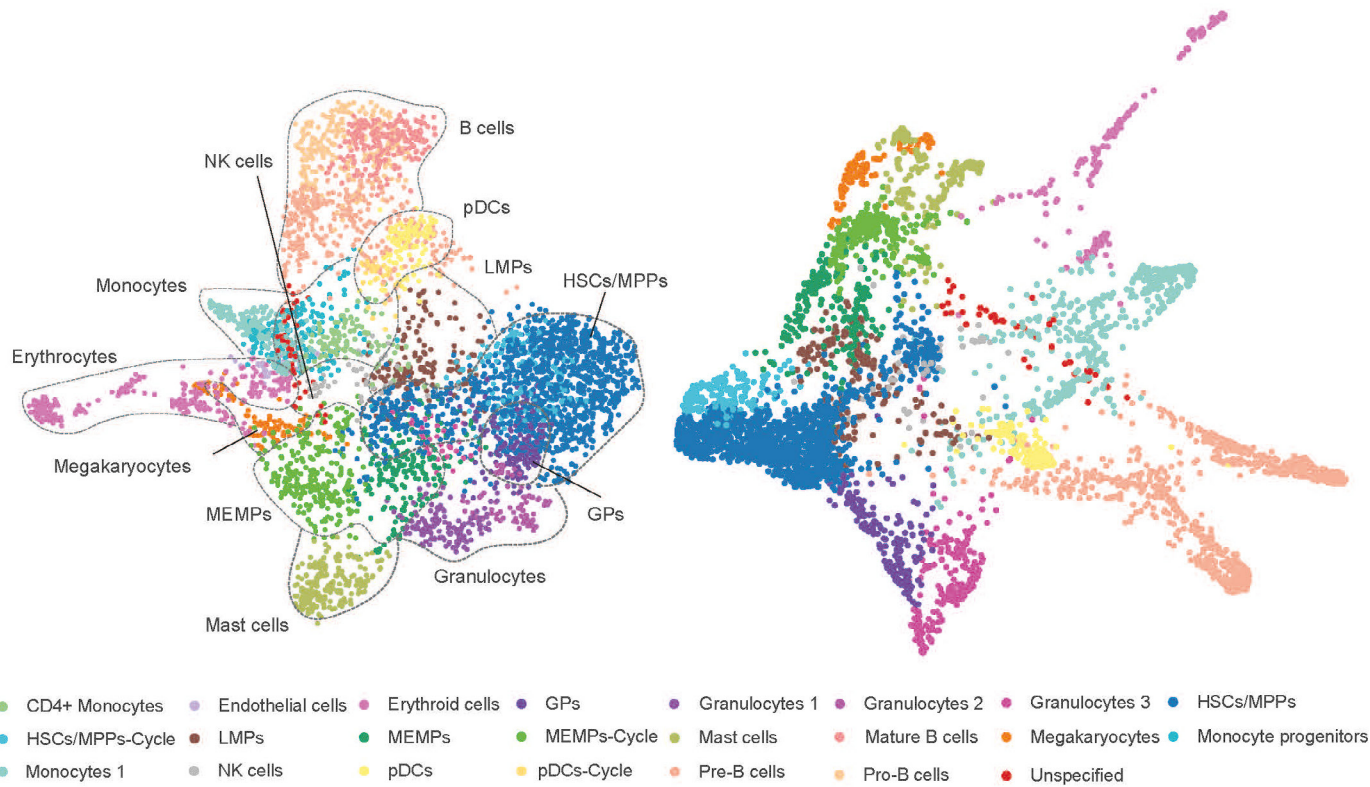
The results from our studies will advance our understanding of how normal fate decisions are instigated and provide clues for the design of novel therapies for blood pathologies.

KEY PUBLICATIONS

Hernández PP, Strzelecka PM, Athanasiadis EI, Hall D, Robalo AF, Collins CM, Boudinot P, Levraud JP, **Cvejic A**. Single-cell transcriptional analysis reveals ILC-like cells in zebrafish. **Science Immunology**. 2018 Nov 16;3(29).

Athanasiadis EI, Botthof JG, Andres H, Ferreira L, Lio P, **Cvejic A**. Single-cell RNA-sequencing uncovers transcriptional states and fate decisions in haematopoiesis. **Nature Communications**. 2017 Dec 11;8(1):2045.

Macaulay IC, Svensson V, Labalette C, Ferreira L, Hamey F, Voet T, Teichmann SA, **Cvejic A**. Single-Cell RNA-Sequencing Reveals a Continuous Spectrum of Differentiation in Hematopoietic Cells. **Cell Reports**. 2016 Feb 2;14(4):966-977.



Uniform Manifold Approximation and Projection (UMAP) visualisation (left) and Force-Directed Graph (FDG) representation (right) of human foetal haematopoietic cells from liver and bone marrow.

Credit: Andrea Tangherloni & Anna Maria Ranzoni

ROBIN FRANKLIN

Adult neural stem cells and CNS regeneration



Our research investigates the mechanisms of Central Nervous System (CNS) regeneration with a particular focus on remyelination, a regenerative process mediated by adult stem cells in which new myelin sheaths are restored to demyelinated axons.

Using a wide range of experimental approaches, we are examining extrinsic (environmental) and intrinsic (transcriptional/epigenetic) factors that govern the responses of adult neural stem cells to injury and their differentiation into oligodendrocytes and other glia following CNS injury.

The potential medical benefits of this research are to stop nerve cell degeneration and therefore provide a treatment for the currently untreatable secondary progressive phase of multiple sclerosis.

LAB MEMBERS

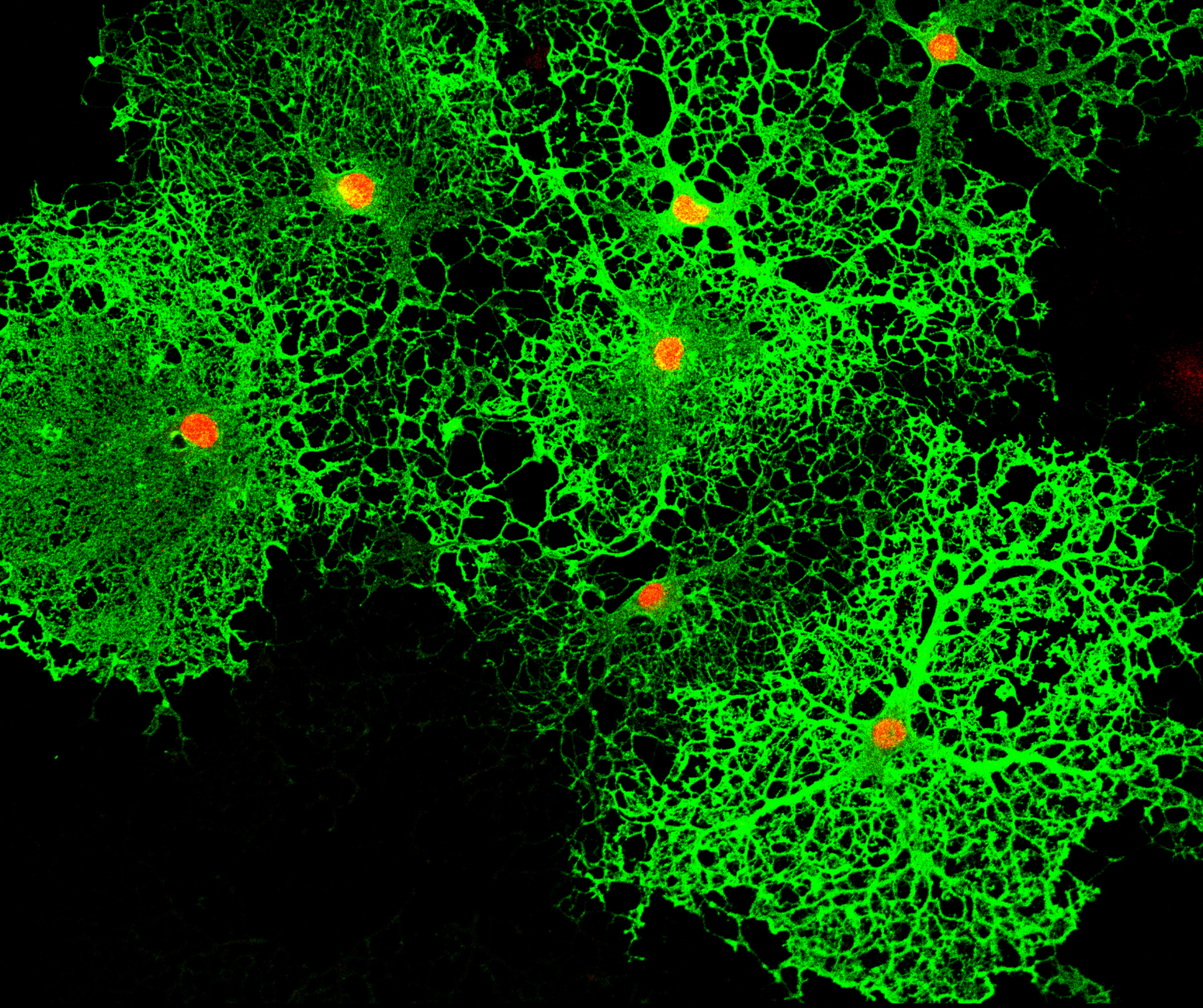
Nejma Belaadi
 Juan Cubillos
 Penelope Dimas
 Sarah Foerster
 Tanay Ghosh
 Ginez Gonzalez
 Myfanwy Hill
 Daniel Morrison
 Bjoern Neumann
 Feride Oeztuerk-Winder
 Khalil Rawji
 Amar Sharma
 Zhaozong Wu
 Adam Young
 Chao Zhao

KEY PUBLICATIONS

Neumann B, Baror R, Zhao C, Segel M, Dietmann S, Rawji KS, Foerster S, McClain C, Chalut KC, van Wijngaarden P, **Franklin RJM**. Metformin restores the potential of aged stem cells for remyelination of the central nervous system. **Cell Stem Cell** (2019) 25: 473-485.

Segel M, Neumann B, Hill MFE, Weber I, Viscomi C, Zhao C, Young A, Agle CC, Thompson AJ, Gonzalez G, Sharma A, Holmqvist S, Rowitch DH, Franze K, **Franklin RJM***, Chalut KC*. Niche stiffness underlies the aging of central nervous system progenitor cells. **Nature** (2019) 573: 130-134

McMurrin CE, Guzman de la Fuente A, Penalva R, Ben Menachem-Zidon O, Dombrowski Y, Falconer J, Gonzalez GA, Zhao C, Krause FN, Young AMH, Griffin JL, Jones CA, Hollins C, Heimesaat MM, Fitzgerald DC, **Franklin RJM**. The microbiota regulates murine inflammatory responses to toxin-induced CNS demyelination but has minimal impact on remyelination. **Proceedings of the National Academy of Sciences USA** (2019) 15: 201905787.



Oligodendrocytes differentiated from adult CNS progenitors

Credit: Bjoern Neumann

CÉDRIC GHEVAERT

In vitro production of platelets for transfusion



LAB MEMBERS

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Holly Foster
Nina Herbert
Daniel Howard
Winnie Lau
Moyra Lawrence
Amanda Niven
Adam Pullen
Amie Waller
James Warland
Souradip Mookerjee

The main focus of our research is the production of blood cells for human use, namely red cells and platelets. We have developed particular expertise in the production of these cell types from human pluripotent stem cells using methodologies that are compatible with the production of clinical grade products within the constraints of affordable manufacturing processes.

To this end, we are combining cellular programming through knowledge and manipulation of transcription factor networks and the creation of 3D biocompatible niches and bioreactors.

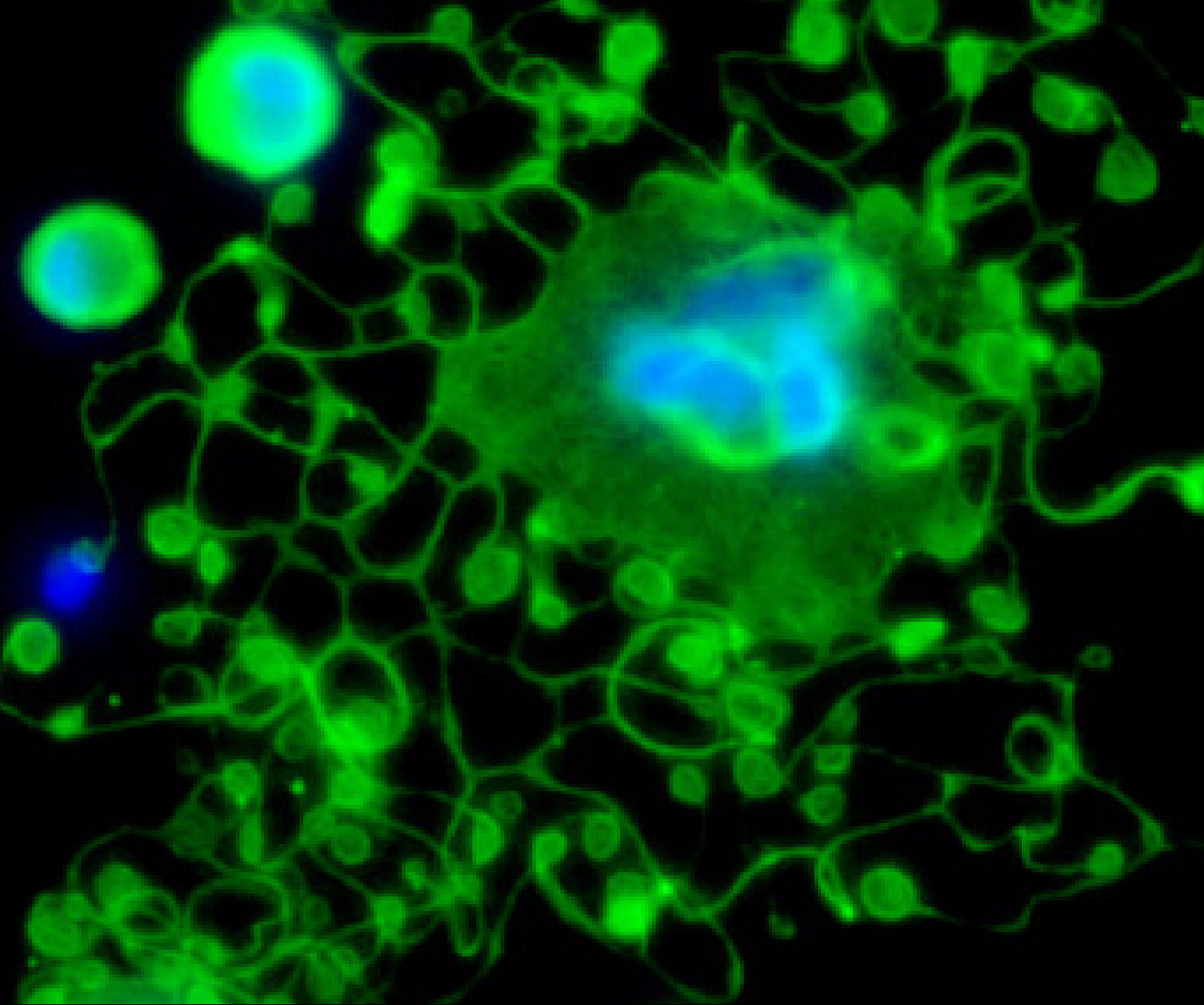
Our expertise is recognised world-wide in carrying out first-in-man studies of blood cell survival and recovery in human volunteers. The RESTORE trial, which is looking at recovery and survival of manufactured red cells in healthy volunteers, is due to start in 2019.

KEY PUBLICATIONS

Shepherd JH, Howard D, Waller AK, Foster HR, Mueller A, Moreau T, ... **Ghevaert C**, Best S, Cameron R. Structurally graduated collagen scaffolds applied to the ex vivo generation of platelets from human pluripotent stem cell-derived megakaryocytes: Enhancing production and purity. **Biomaterials**. 2018;182:135-44.

Dalby A, Ballester-Beltrán J, Lincetto C, Mueller A, Foad N, Evans A, Baye J, Turro E, Moreau T, Tijssen MR, **Ghevaert C**. Transcription Factor Levels after Forward Programming of Human Pluripotent Stem Cells with GATA1, FLI1, and TAL1 Determine Megakaryocyte versus Erythroid Cell Fate Decision. **Stem Cell Reports**. 2018; 11(6):1462-1478.

Moreau T, Evans AL, Vasquez L, ..., Ouwehand WH, Pedersen RA, **Ghevaert C**. Large-scale production of megakaryocytes from human pluripotent stem cells by chemically defined forward programming. **Nature Communications**. 2016;7:11208.



Fluorescent image of a megakaryocyte produced in the laboratory from stem cells isolated from cord blood. The megakaryocyte is in the process of releasing platelets, the little blood corpuscle that is responsible for blood clotting. The nucleus of the cell is stained in blue with DAPI. In green is one of the main components of the skeleton of the cell (tubulin) that drives the production of platelets.

Credit: Maria Colzani

BERTIE GÖTTGENS

Cellular decision making in normal and leukaemic blood stem cells



We use a combination of experimental and computational approaches to study how transcription factor networks control the function of blood stem cells and how mutations that perturb such networks cause leukaemia.

This integrated approach has resulted in the discovery of new combinatorial interactions between key blood stem cell regulators, as well as experimentally validated computational models for blood stem cells.

Our current research focuses on (i) single cell genomics of early blood development, (ii) modelling the transcriptional landscape of blood stem and progenitor cell differentiation, (iii) transcriptional consequences of leukaemogenic mutations in leukaemia stem/progenitor cells, and (iv) molecular characterisation of human blood stem/progenitor cell populations used in cell and gene therapy protocols.

LAB MEMBERS

Melania Barile
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Myriam Haltalli
Rebecca Hannah
Ivan Imaz-Rosshandler
Alison Kennedy
Sarah Kinston
Iwo Kucinski
Mariana Quiroga Londono
Kat Sturgess
Mai-Linh Ton
Sam Watcham
Xiaonan Wang
Nicola Wilson

KEY PUBLICATIONS

Pijuan-Sala B, Wilson NK, Xia J, Hou X, Hannah RL, Kinston S, Calero-Nieto FJ, Poirion O, Preissl S, Liu F, **Göttgens B**. Single-cell chromatin accessibility maps reveal regulatory programs driving early mouse organogenesis. **Nature Cell Biology**, 22; 487–497.

Pijuan-Sala P, Griffiths JA, Guibentif C, Hiscock TW, Jawaid W, Calero-Nieto FJ, Mulas C, Ibarra-Soria X, Tyser RCV, Ho DLL, Reik W, Srinivas S, Simons BD, Nichols J, Marioni JC, **Göttgens B**. A single-cell molecular map of mouse gastrulation and early organogenesis. **Nature**. 2019 Feb;566(7745):490-495.

Scialdone A, Tanaka Y, Jawaid W, Moignard V, Wilson NK, Macaulay IC, Marioni JC, **Göttgens B**. (2016). Resolving Early Mesoderm Diversification through Single Cell Expression Profiling. **Nature**. 535 (7611): 289-293.



Molecular Map: each dot represents a single cell in the developing embryo. The dots are coloured based on which major cell type they represent.

Credit: Blanca Pijuan-Sala

TONY
GREEN

Haematopoiesis and myeloid malignancies



LAB MEMBERS

Marina Akasaka
Juliet Aungier
Hugo Bastos
Miriam Belmonte
Tina Hamilton
Thorsten Klampfl
Joe Lee
Juan Li
Stephen Loughran
Hyun Jung Park
Dean Pask
Daniel Prins
Rachel Sneade
Michelle Wantoch
Matthew Williams

Our lab focuses on the mechanisms whereby blood stem cells are subverted during the development of haematological malignancies, with particular focus on JAK/STAT signalling, which is dysregulated in many cancers and plays a key role in multiple stem cell systems.

In particular, we explore the molecular and cellular pathogenesis of a group of pre-leukaemic disorders, the myeloproliferative neoplasms (MPNs), in studies which span basic, translational and clinical research. The MPNs harbour mutations that activate the JAK/STAT pathway, are experimentally tractable and provide a paradigm for the earliest stages of tumorigenesis.

In work which transformed MPN diagnosis and catalysed development of therapeutic JAK-family kinase inhibitors, we and others identified phenotypic driver mutations in JAK2 or CALR which activate the JAK/STAT pathway and are present in most MPN patients. We are employing a variety of genomic approaches to explore MPN biology and improve patient management. In parallel, we are investigating the functional consequences of JAK2 and CALR mutations in work which has led to unexpected insights into cancer biology, human haematopoiesis and cytokine signalling.

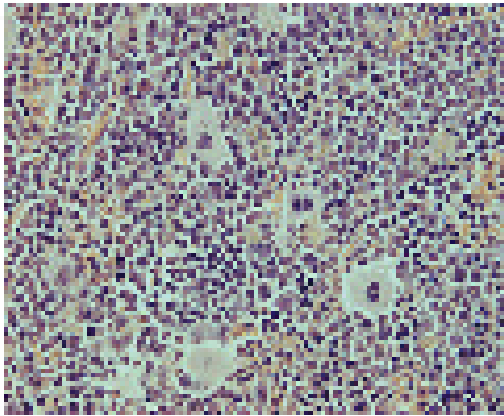
KEY PUBLICATIONS

Grinfeld J, Nanglia J, Baxter EJ, ..., Gerstung M, **Green AR**, Campbell PJ. (2018). Disease heterogeneity and personalized prognosis in myeloproliferative neoplasms. **New England Journal of Medicine**. 2018;379:1416-30.

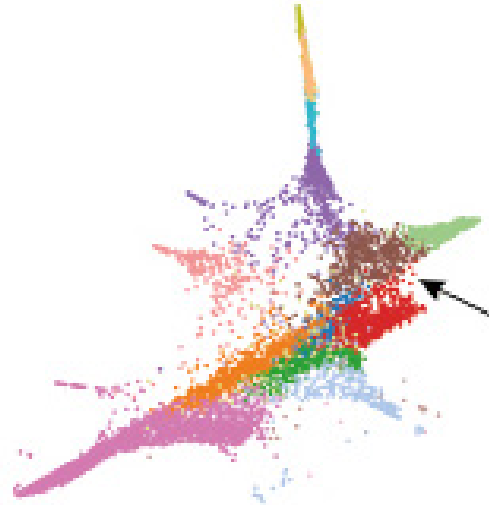
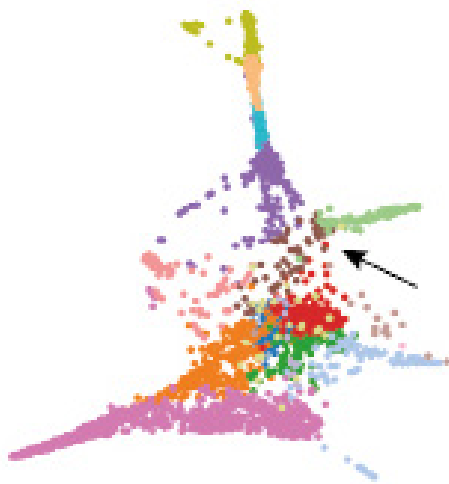
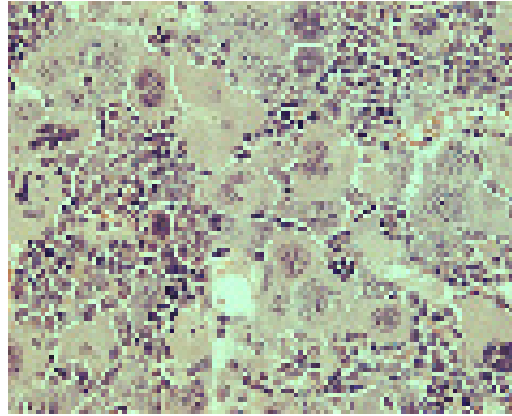
Lee-Six H, Obro NF, Shepherd MS, ..., **Green AR**, Kent DG, Campbell PJ. Population dynamics of normal human blood inferred from spontaneous somatic mutations. **Nature**. 2018 Sep;561(7724):473-478.

Loughran SJ, Comoglio F, Hamey FK, ..., Mead AJ, Hendrich B, **Green AR**. Mbd3/NuRD controls lymphoid cell fate and inhibits tumorigenesis by repressing a B cell transcriptional program. **Journal of Experimental Medicine**. 2017 Oct 2;214(10):3085-3104.

Wildtype



Mutant CALR



Knock-in mice expressing mutant calreticulin develop markedly increased megakaryopoiesis (upper panels). Single cell RNAseq analysis of haematopoietic stem and progenitor cells reveals a novel megakaryocytic progenitor population (lower panels, brown dots indicated by arrows).

Credit: Juan Li, Daniel Prins & Sam Watcham

BRIAN HENDRICH

Transcriptional control of stem cell fate



LAB MEMBERS

Robin Floyd
Dominic Hall
Andria Koulle
Oluwaseun Ogundele
Ramy Ragheb
Nicola Reynolds

Embryonic stem (ES) cells hold enormous promise for personalised medicine and drug discovery since they can be maintained indefinitely and are pluripotent. While pluripotency makes ES cells potentially very useful, it also presents a problem: how do you get them to make the cell type you want, and not one you don't? Differentiation of pluripotent cells is exquisitely organised during normal embryogenesis, but very hard to control in culture.

Since all cells in an organism are genetically identical, the observable differences in their functions and behaviours come down to which genes they express and which genes they repress. Therefore, in order to understand how to direct cellular identity, our lab seeks to understand how subtle differences in gene expression patterns in seemingly identical cells influence any subsequent differentiation decisions.

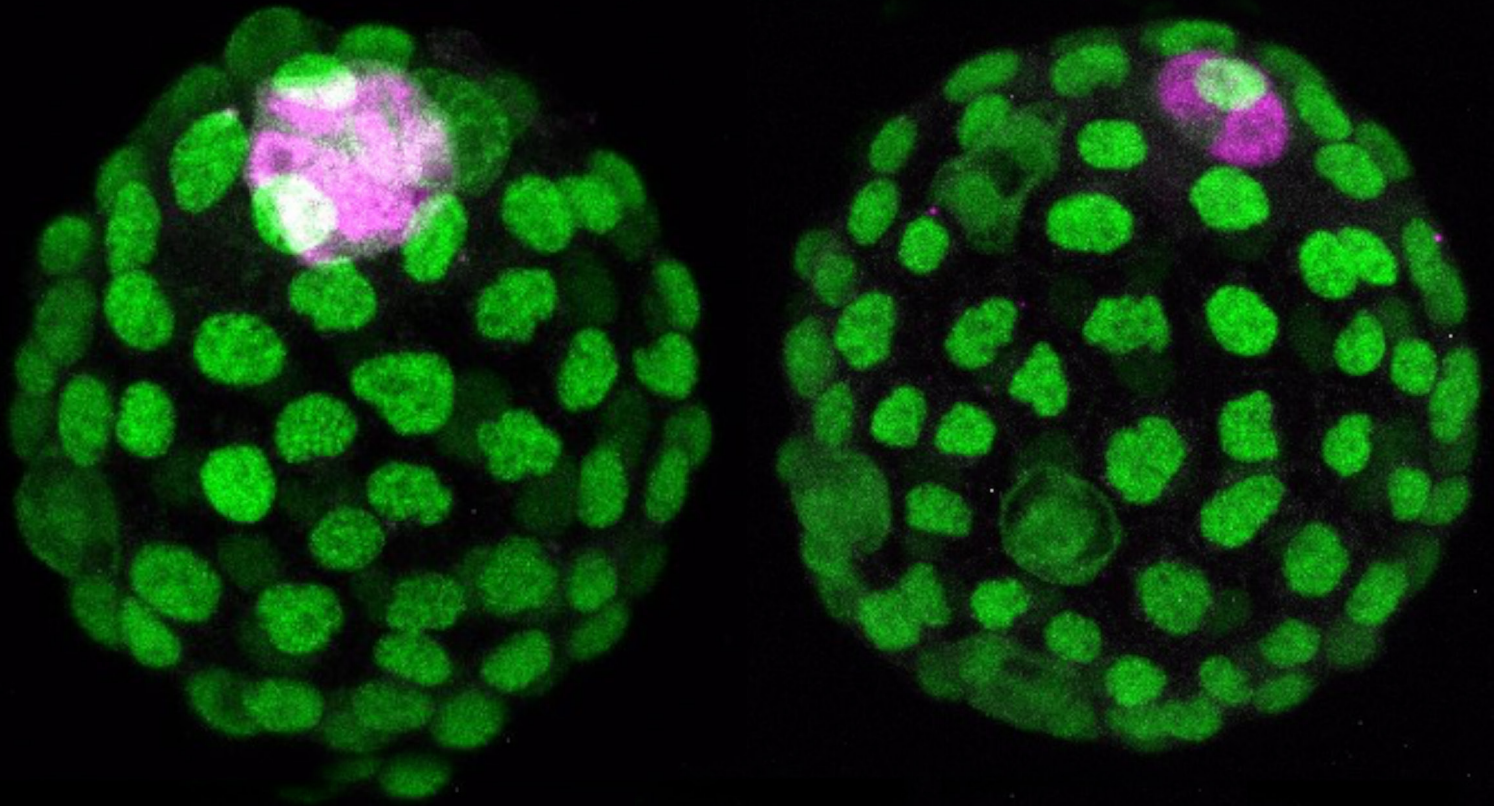
By understanding how ES cells make different developmental decisions, we hope to bring the medical promise of stem cells closer to realisation.

KEY PUBLICATIONS

Ragheb R, Gharbi S, Cramard J, Ogundele O, Kloet S, Burgold T, Vermeulen M, Reynolds N, **Hendrich B**. Differential regulation of lineage commitment in human and mouse primed pluripotent stem cells by NuRD. **bioRxiv** doi: <https://dx.doi.org/10.1101/2020.02.05.935544>.

Burgold T, Barber M, Kloet S, Cramard J, Gharbi S, Floyd R, Kinoshita M, Ralser M, Vermeulen M, Reynolds N, Dietmann S, and **Hendrich B**. The Nucleosome Remodelling and Deacetylation complex suppresses transcriptional noise during lineage commitment. **The EMBO Journal**. 2019 Jun 17;38(12):e100788.

Bornelöv S, Reynolds N, Xenophontos M, Gharbi S, Johnstone E, Floyd R, Ralser M, Signolet J, Loos R, Dietmann S, Bertone P, **Hendrich B**. The Nucleosome Remodeling and Deacetylation Complex Modulates Chromatin Structure at Sites of Active Transcription to Fine-Tune Gene Expression. **Molecular Cell**. 2018 Jul 5;71(1):56-72.e4.



Chimaeric blastocysts were created using wild type mouse embryos and either wild-type embryonic stem (ES) cells (left) or ES cells in which the important chromatin remodelling complex, NuRD, was missing (right). ES cells were labelled with a protein, here shown in white. The embryos are also stained for nuclear markers of trophectoderm (Cdx2 - Green) or inner cell mass (Sox2 - Magenta). Wild type ES cells contribute significantly to the inner cell mass lineage but never to the trophectoderm lineage. In contrast, cells lacking the NuRD complex contribute poorly to chimaeric blastocysts because they are unable to properly respond to the developmental cues and contribute significantly to the inner cell mass, though they also fail to adopt the inappropriate identity of trophectoderm cells. These and other experiments in the lab have shown how chromatin remodelling is important for stem cells to properly execute lineage decisions and form the normal range of differentiated cell types. Burgold et al. 2019 doi: <http://doi.org/10.15252/embj.2018100788>

Photo credit: Nicola Reynolds and Brian Hendrich

DANIEL HODSON

Mutation timing in lymphomagenesis



LAB MEMBERS

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Laura Everton
Rachel Fenner
Jane Gao
Jade Gong
Joanna Krupka
Hendrik Runge

Normal B lymphocytes progress through a series of developmental stages that begin with the haematopoietic stem cell. Progression through each of these stages is tightly controlled at both the transcriptional and post-transcriptional levels. Genetic alterations and mutations, which can occur at any stage from the haematopoietic stem cell to the post-germinal centre B cell, can lead to loss of this normal regulation and subsequently to the development of lymphoid malignancies such as non-Hodgkin Lymphoma (NHL), which is the 6th commonest form of human cancer.

Understanding how these genetic alterations corrupt cell fate choices at each stage of lymphocyte development will be the key to identifying cellular pathways that can be therapeutically targeted. Our group is developing novel cell culture models to study the effects of these genetic alterations in human lymphocytes. In particular, we are interested in studying how these genetic alterations lead to changes at the level of mRNA translation and how these post-transcriptional changes then contribute to lymphomagenesis.

We use a variety of techniques including exome and RNA sequencing, ribosome profiling, iCLIP and xenografts to identify the developmental timing of these genetic alterations, their mechanistic contribution to lymphomagenesis and the implications this has for the treatment and monitoring of patients.

KEY PUBLICATIONS

Gong C, Krupka JA, Gao J, ..., Du MQ, Samarajiwa S & **Hodson DJ**. Sequential inverse dysregulation of the RNA helicases DDX3X and DDX3Y facilitates MYC-driven lymphomagenesis. **Cell Press Sneak Peek**, doi.org/10.2139/ssrn.3520953.

Lacey S, Barrans S, Beer P, ..., Burton C, Crouch S & **Hodson DJ**. Targeted sequencing in DLBCL, molecular subtypes and outcomes: a Haematological Malignancy Research Network report. **Blood**. doi./10.1182/blood.2019003535.

Caeser R, Di Re M, Krupka JA, ..., Mupo A, Bashford-Rogers RJM & **Hodson DJ**. Genetic modification of primary human B cells generates translationally-relevant models of high-grade lymphoma. **Nature Communications**, 10(1):4543.

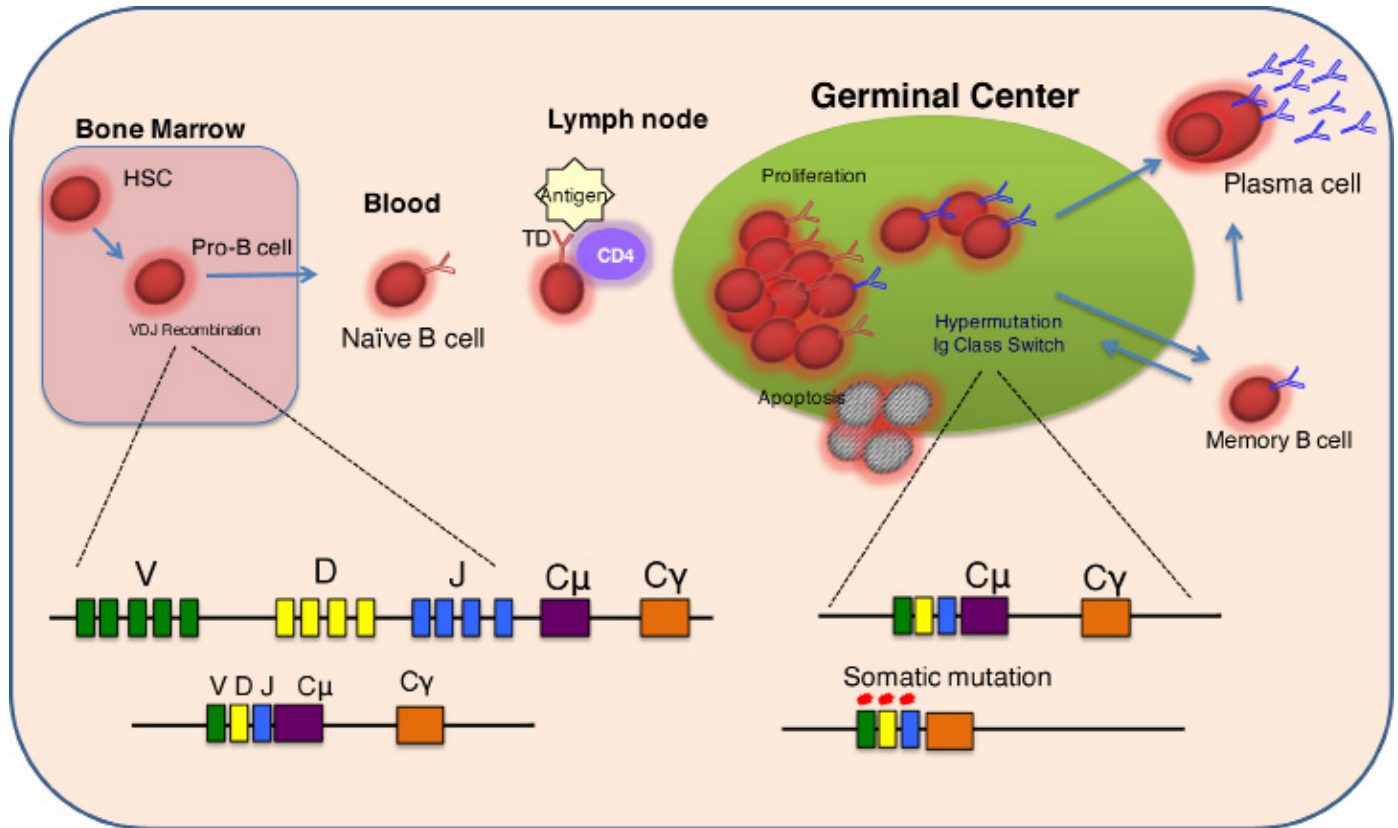


Diagram showing the unique and dangerous life of the B lymphocyte with recurrent episodes of deliberately induced DNA damage contributing to the risk of transformation to B cell lymphoma.

Credit: Hodson Lab

BRIAN HUNTLY

Leukaemia stem cell biology and leukaemogenesis



LAB MEMBERS

Shuchi Agrawal Singh
 Alicia Garcia-Gimenez
 George Giotopoulos
 Sarah Horton
 David Lara
 Pedro Madrigal
 Ludovica Marando
 Eshwar Meduri
 Nisha Narayan
 Simon Richardson
 Nathalie Sakakini

Leukaemias have recently been demonstrated to be wholly dependent upon a small population of so-called cancer stem cells. These cells represent the critical targets for treatment and a greater understanding of their biology and its interface with normal stem cell function is fundamental to improving treatment outcomes.

The focus of our research is on this interface. We use a combination of techniques in cell line and animal models, as well as confirmatory studies in primary human tissue, to dissect stem cell function. Our aim is to understand how normal stem cell function is subverted in cancer and how these processes might be therapeutically targeted to improve the outcome in haematological malignancies. We are examining the role of mutations that occur in, and alter the role of haematopoietic stem and progenitors as early events before leading to the subsequent development of leukaemias and lymphomas (pre-leukaemic stem cells). Many of these mutations alter epigenetic regulation, enhancer function and transcriptional programmes, all of which are ongoing areas of investigation within the lab.

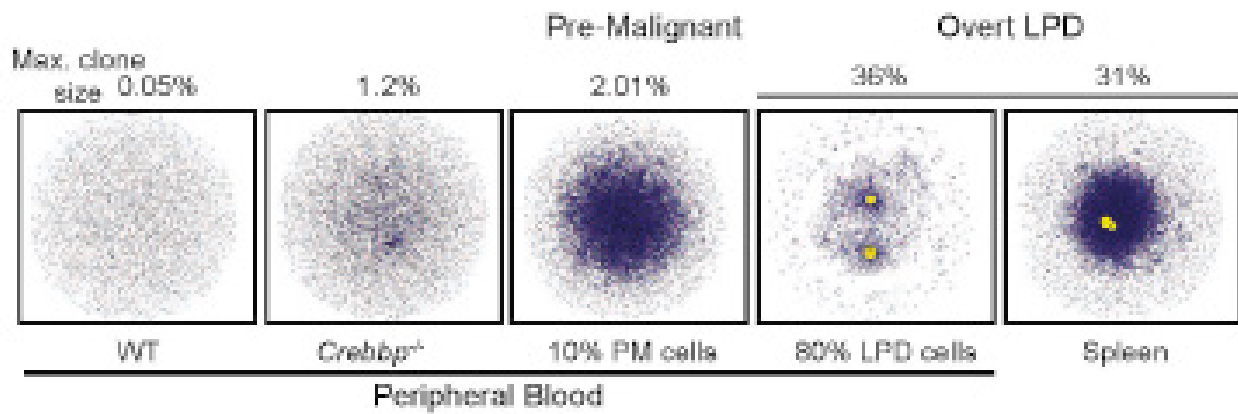
Therapeutically, a recent example of our work is the identification of the Bromodomain and extra terminal (BET) proteins as critical mediators of leukaemia stem cells in Acute Myeloid Leukaemia. An inhibitor of these proteins has already entered early phase clinical trials in relapsed blood cancers.

KEY PUBLICATIONS

Gozdecka M, Meduri E, Mazan M, ..., Adams D, Vassiliou GS, **Huntly BJP**. UTX-mediated enhancer and chromatin remodelling suppresses myeloid leukaemogenesis through non-catalytic inverse regulation of ETS and GATA expression programmes. **Nature Genetics**. 2018 Jun;50(6):883-894.

Horton SJ, Giotopoulos G, Yun H, ..., Du MQ, Adams DJ, **Huntly BJP**. Early loss of Crebbp confers malignant stem cell properties on lymphoid progenitors. **Nature Cell Biology**. 2017 Sep;19(9):1093-1104.

Basheer F, Giotopoulos G, Meduri E, ... Campbell P, Vassiliou G, **Huntly BJP**. Contrasting requirements during disease evolution identify EZH2 as a therapeutic target in AML. **Journal of Experimental Medicine**. 2019 Apr 1;216(4):966-98.



Charting the clonal evolution of lymphoma (from left to right), using specific immunoglobulin rearrangements as specific clonal cellular barcodes (taken from Horton et al, Nature Cell Biology 2017).

Credit: Sarah Horton

RAGNHILDUR THÓRA KÁRADÓTTIR

Neurotransmitter signalling to central nervous system progenitor cells



LAB MEMBERS

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Omar De Faria Junior
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Helena Pivonkova
Sebastian Timmler
Balazs Varga

Our lab investigates how the process of myelination in the central nervous system (CNS) is regulated, and how myelin regeneration could be manipulated to tackle diseases such as cerebral palsy, spinal cord injury and multiple sclerosis.

Unique to the CNS, myelin regeneration can occur spontaneously due to the presence of brain stem cells called oligodendrocyte precursor cells (OPCs) which differentiate into new myelinating oligodendrocytes. However, this process often fails, making OPCs differentiation an important therapeutic target and the focus of investigation in our lab.

We have previously shown that OPCs express neurotransmitter receptors and receive synaptic inputs from neuronal axons in the white matter, hence are capable of sensing changes in neuronal activity. The lab aims to understand how signals from neurons induce OPCs to differentiate and myelinate axons during development and with normal ageing; this also could be an underlying mechanism for white matter plasticity.

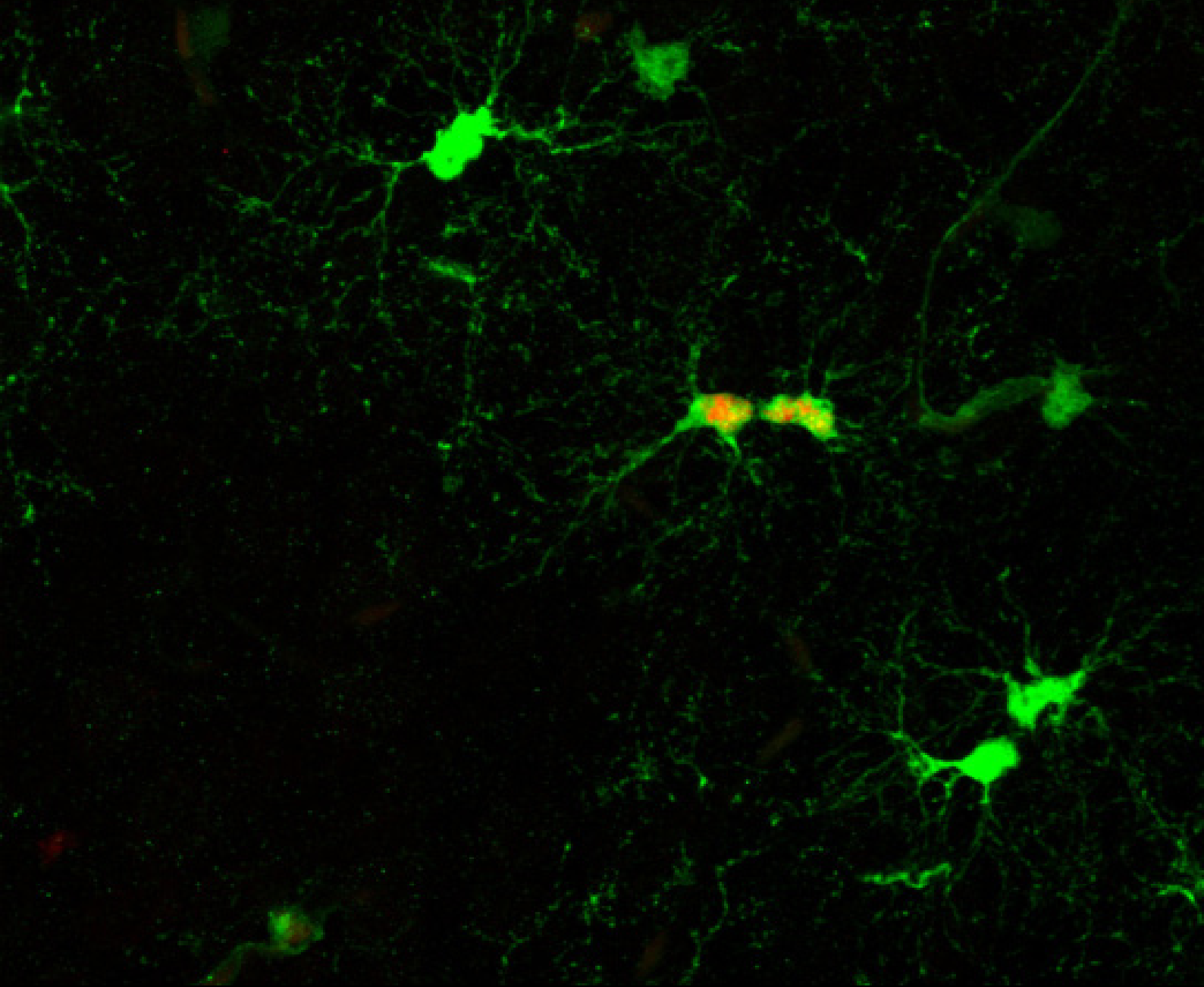
We are actively investigating how OPCs respond to myelin injury and whether neuronal activity and neurotransmitter signalling may regulate the myelin repair process. Our ultimate aim is to find new treatments for white matter disease.

KEY PUBLICATIONS

Spitzer S, Sitnikov S, Kamen Y, Evans KA, Kronenberg-Versteeg D, Dietmann S, de Faria O, Agathou S & **Káradóttir R**. Oligodendrocyte progenitor cells become regionally diverse and heterogeneous with age. **Neuron**. 2019 Feb 6;101(3):459-471.

Gautier HO, Evans K, Lundgaard I, James F, Lao-Peregrin C, Franklin RJM, **Káradóttir R**. Neuronal activity regulates remyelination via glutamate signalling to oligodendrocyte progenitors. **Nature Communications**. 2015 Oct 6;6:8518.

Lundgaard I, Luzhynskaya A, Stockley JH, ..., ffrench-Constant C, Attwell D, **Káradóttir R**. Neuregulin and BDNF induce a switch to NMDA receptor dependent myelination by oligodendrocytes. **PLoS Biology**. 2013 Dec;11(12):e1001743.



Proliferating oligodendrocyte progenitor cells in a mouse brain slice.

Credit: Kimberley Evans

ELISA
LAURENTI

Human haematopoietic stem cells biology in health and disease



LAB MEMBERS

Emily Calderbank
Anna Clay
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Daniel Hayler
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Nicole Mende
Kendig Sham
Aditi Vedi

Haematopoietic stem cells (HSC) are responsible for life-long blood production. They are the best-studied stem cell type owing to decades of research with animal models.

Despite the high incidence of blood-related diseases, and accumulating evidence that certain aspects of HSC biology are species-specific, very little is known on human HSC. Our laboratory develops integrated approaches combining *in vitro* and *in vivo* single cell assays, transcriptomics and bioinformatics to study human HSC and progenitor cells.

We are currently investigating how cell cycle regulation, inflammation and ageing, processes intimately linked to disease initiation, affect human HSC unique molecular and functional properties.

We also study HSC clonal dynamics in humans, in particular how these are affected by specific preleukaemic mutations. Understanding how the cellular and molecular composition of the HSC/progenitor compartment changes in stress conditions and throughout a human lifetime has important implications for regenerative medicine and treatment of blood cancers.

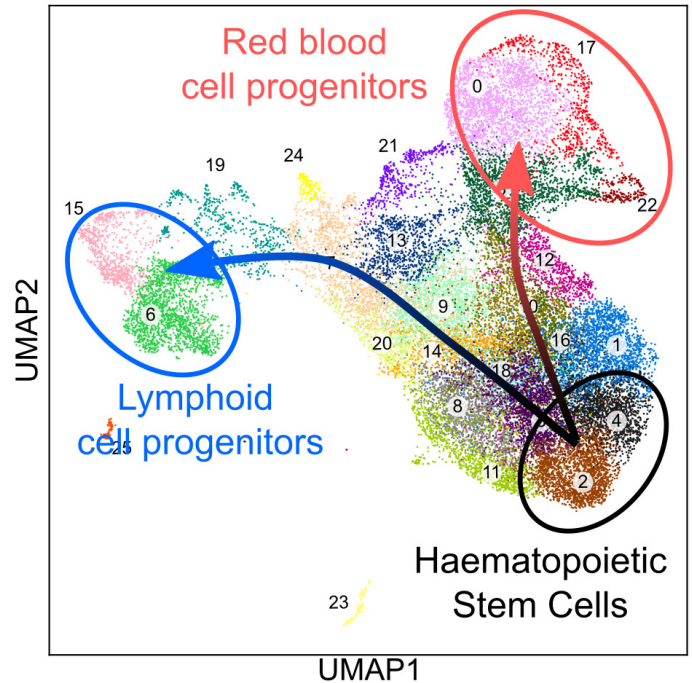
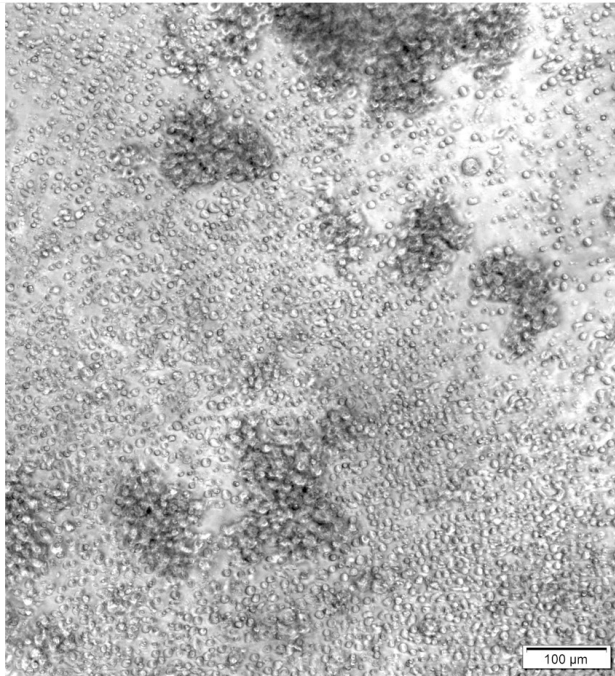
KEY PUBLICATIONS

Popescu D, Botting RA, Stephenson E, Green K, Webb S, Jardine L, Calderbank EF, ... Regev AD, Chedotal A, Roberts I, Gottgens B, Behjati S*, **Laurenti E***, Teichmann SD*, Haniffa M*. Decoding human fetal liver haematopoiesis. **Nature**, 574, 2019, 365-371. *equal contribution

Belluschi S, Calderbank EF, Ciaurro V, Pijuan-Sala B, Santoro A, Mende N, Diamanti E, Sham KYC, Wang X, Lau WWY, Jawaid W, Göttgens B, **Laurenti E**. Myelo-lymphoid lineage restriction occurs in the human haematopoietic stem cell compartment before lymphoid-primed multipotent progenitors. **Nature Communications**. 2018 Oct 5;9(1):4100.

Laurenti E & Göttgens B. From haematopoietic stem cells to complex differentiation landscapes. **Nature**. 2018 Jan 24;553(7689):418-426.

Connecting human haematopoietic stem cell function to blood formation over a human lifetime



Left: image of different blood cell types originated from a single human haematopoietic stem cell in vitro (credit: Aditi VEDI);

Right: the transcriptional landscape of the human haematopoietic hierarchy, derived from single cell transcriptomics (credit: Hugo Bastos, Nicole Mende).

JOO-HYEON LEE

Stem cells and niches



LAB MEMBERS

Jinwook Choi
Catherine Dabrowska
Frances England
Kelly Evans
Rhys Fox
Antranik Mavousian
Vishal Menon
Sagar Varankar
June Young Park

The lung is a very slow cycling organ that is composed of diverse epithelial and stromal cell types, but has capacity to rapidly regenerate new cells after injury. Our group is trying to understand how stem cells respond to different signals from their local environment and orchestrate the changes in chromatin, transcription, translation, and cellular dynamics in homeostasis and injury repair.

We investigate the regulatory networks that need to be turned on and off at the right time and place for stem cells to become activated and generate specialised cell types during regeneration. We are also interested in defining cellular heterogeneity and plasticity during this process. Elucidating the normal process of lung dynamics will provide us a foundation to understand lung diseases and cancer.

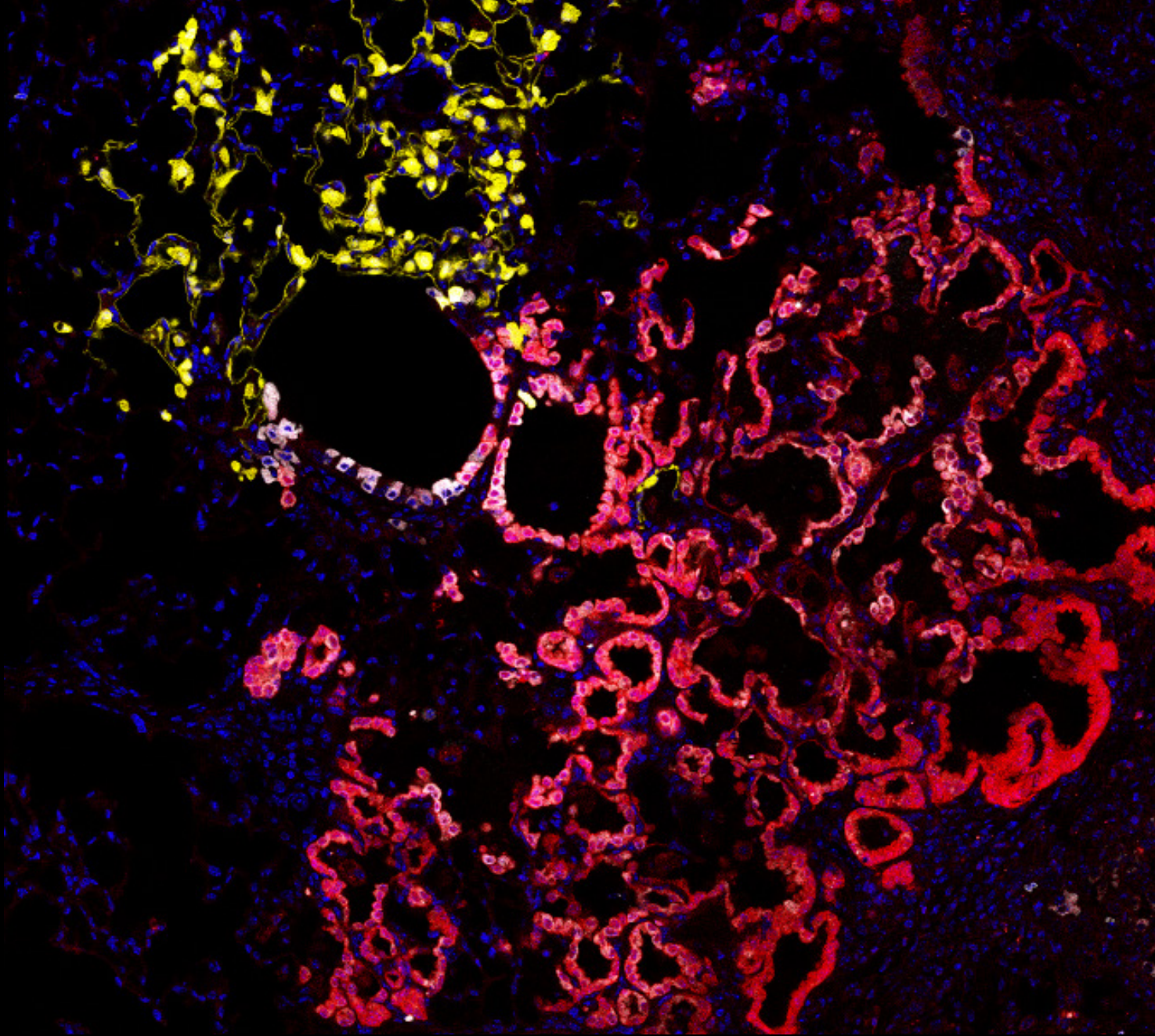
We couple *ex vivo* 3D organoid cultures of human and mouse lungs with genetic tools, *in vivo* transgenic mouse models with lineage tracing techniques, quantitative mathematic modelling of clonal dynamics, and bioinformatics at the single cell level.

KEY PUBLICATIONS

Ombrato L, Nolan E, Kuerlac I, Mavousian A, Bridgeman V, Heinze I, Chakravarty P, Horswell S, Gonzales-Gualda E, Maticcchione G, Weston A, Kirkpatrick J, Husain E, Speirs V, Collinson L, Ori A, **Lee JH**[†] and Malanchi I[†]. Metastatic niche labelling reveals tissue parenchyma stem cell features. **Nature**, 572 (7771), 603-608.

Lee JH, Tammela T, Hofree M, Choi J, Marjanovic ND, Han S, Canner DA, Wu K, Paschini M, Bhang DH, Jacks T, Regev A, Kim CF. Anatomically and functionally distinct lung mesenchymal populations marked by Lgr5 and Lgr6. **Cell**, 170(6):1149-1163.e12.

Lee JH, Bhang DH, Beede A, Huang TL, Stripp B, Bloch KD, Wagers AJ, Tseng YH, Ryeom S, Kim CF. Lung stem cell differentiation in mice directed by endothelial cells via a BMP4-NFATc1-Thrombospondin-1 axis. **Cell**, 156(3):440-55.



Fluorescent image of a mouse lung undergoing repair after injury. Airway secretory club cells labelled with yellow are differentiating into alveolar lineage cells, whereas mutant secretory club cells labelled with red show perturbed differentiation capacity. Nuclei of individual cells are stained with DAPI (blue).

Credit: Catherine Dabrowska

ANDREW MCCASKIE



LAB MEMBERS

Mohammad Alkhrayef
 Francesca Beaton
 Mark Birch
 Roger Brooks
 Stephen Gadomski
 Frances Henson
 William Hotham
 Andrew Hotchen
 Wasim Khan
 Sarah Lindsay
 Stephen McDonnell
 Karin Newell
 Matthew Seah

Regenerative therapies for bone and cartilage repair

The research aim in our lab is to develop innovative therapies for musculoskeletal disease, particularly in Osteoarthritis (OA) which affects around 8 million people in the UK alone. We are currently developing translational pathways for regenerative therapy in this area, linking laboratory research with clinical treatment, including clinical trials.

Our research programmes focus on the opportunity to use adult stem/stromal populations, along with other relevant cell types (haematopoietic and chondrocyte) either alone or with tissue engineering approaches to target early disease. Research also considers the mechanisms of joint destruction relevant to repair.

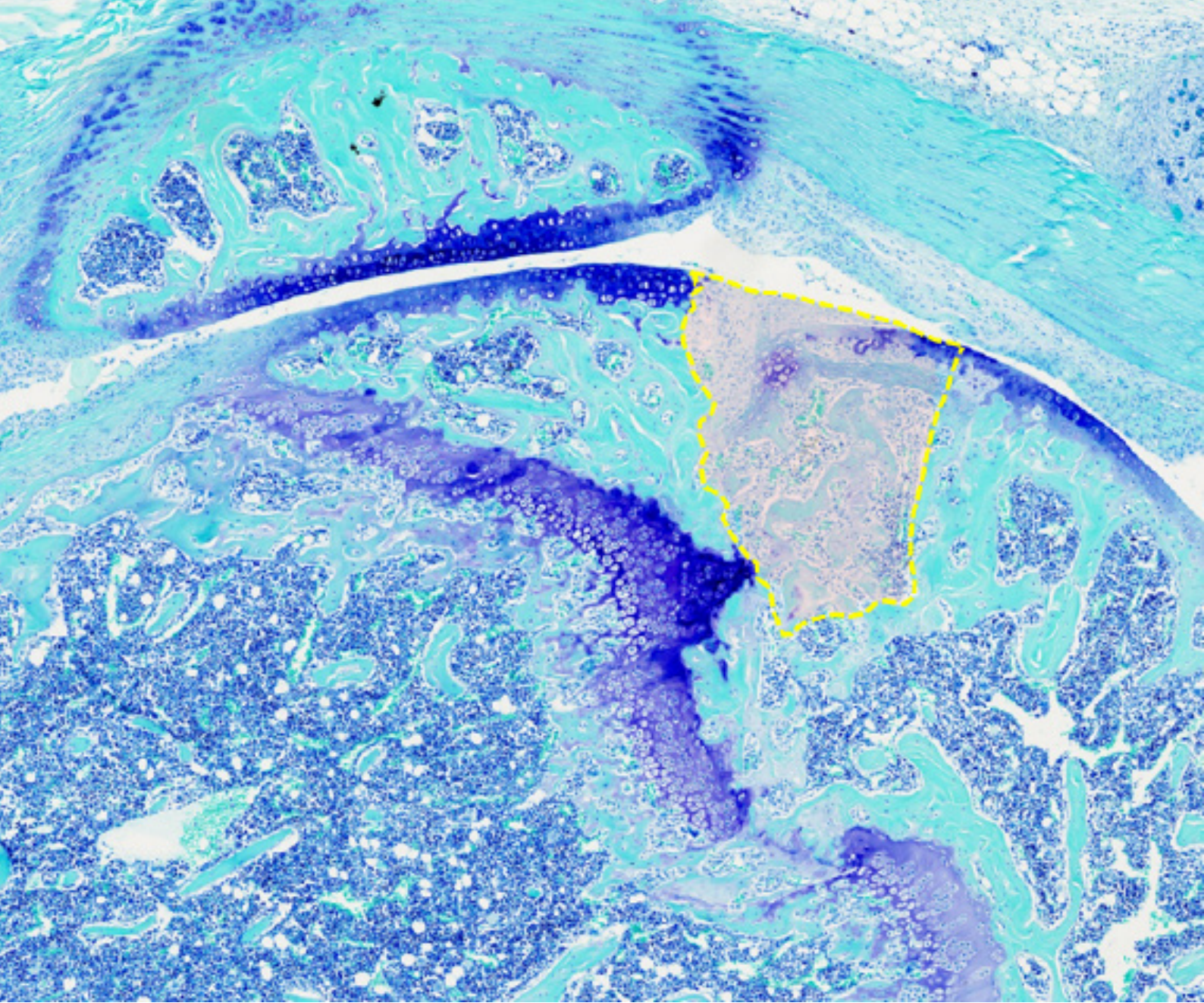
The translational and clinical programmes seek to use stratified and experimental medicine approaches, particularly focused on imaging and tissue analysis during cartilage repair surgery. The latter will include cell characterisation by phenotype and single cell analysis to understand the role played by cell therapies in the repair of joint tissues.

KEY PUBLICATIONS

Palmer AJR, Ayyar Gupta V, Fernquest S, ... **McCaskie AW**, ... Carr AJ, Beard DJ, Glyn-Jones S, FAIT Study Group. Arthroscopic hip surgery compared with physiotherapy and activity modification for the treatment of symptomatic femoroacetabular impingement: multicentre randomised controlled trial. *BMJ* 364, I185.

Steinberg J, Brooks R, Southam L, ... **McCaskie AW**, Zeggini E. Widespread Epigenomic, Transcriptomic and Proteomic Differences Between Hip Osteophytic and Articular Chondrocytes in Osteoarthritis. **Rheumatology** (Oxford). 2018 Aug 1;57(8):1481-1489.

Evangelou E, Kerkhof HJ. ... **McCaskie A**,... Valdes AM. A meta-analysis of genome-wide association studies identifies novel variants associated with osteoarthritis of the hip. **Annals of the Rheumatic Diseases**. 2014 Dec;73(12):2130-6.



Osteochondral repair of a synovial joint injury.

Credit: Francesca Beaton

SIMÓN MÉNDEZ-FERRER

Blood stem cell niches



LAB MEMBERS

Claire Fielding
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Antony (Ya-Hsuan) Ho
Thomas McKerrell
Francesca Panvini
Antonio Rodriguez Romera
Jun Zhang

Our research focuses on the regulation of the haematopoietic stem-cell niche in health and disease. Blood stem cells reside in specialised niches which allows them to self-renew, proliferate, differentiate and migrate according to the organism's requirements.

The group studies multisystem regulatory mechanisms by which the haematopoietic stem cell niche fulfils these complex functions and how the deregulation of these mechanisms contributes to haematological disorders. The group has demonstrated that the brain regulates a peripheral stem cell niche in the bone marrow partly through sympathetic innervation of nestin+ niche cells. Protection of this regulatory network, whose constituents might share a related ancestry, can block the manifestation of myeloproliferative neoplasms.

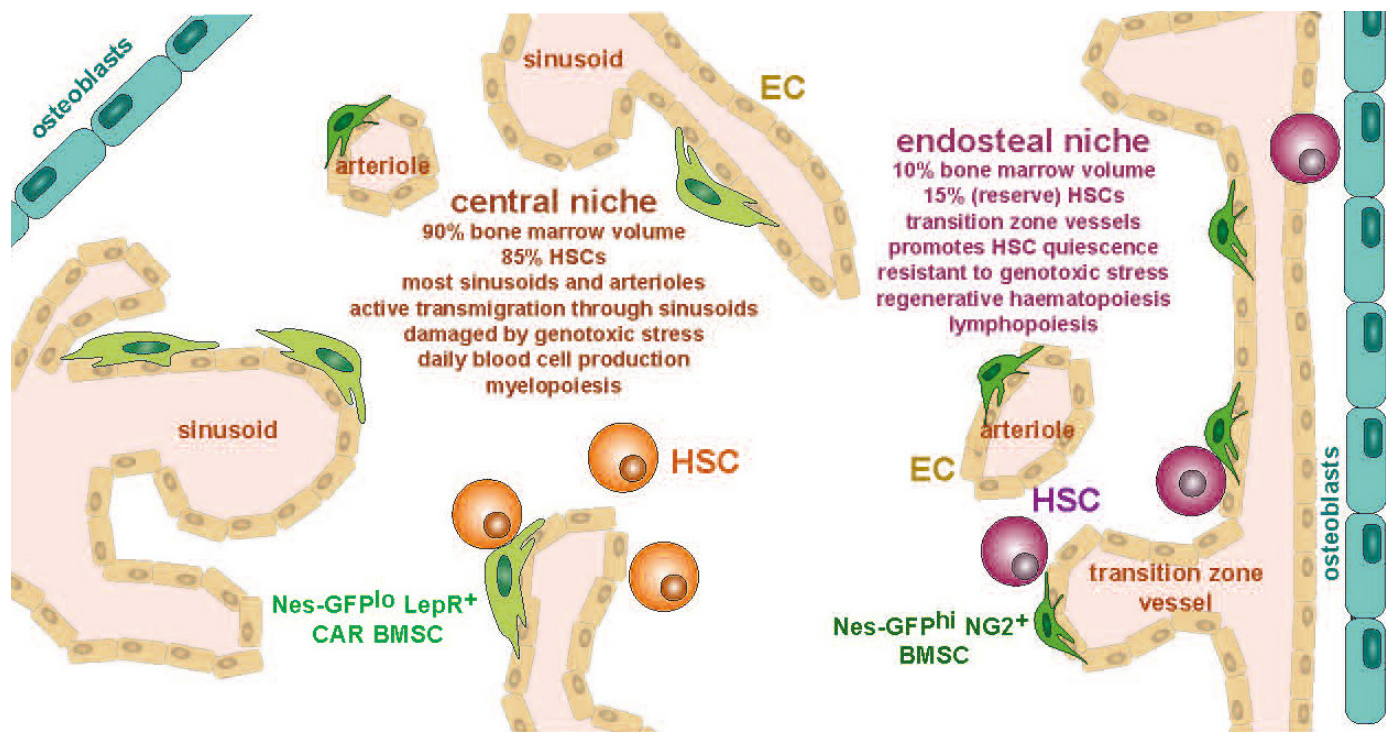
Our research indicates that neuroendocrine regulation of bone marrow stem cells by adrenergic signals or by sex hormones could potentially offer novel therapeutic approaches. We study the interaction of mesenchymal and haematopoietic stem cells and its implications for bone marrow transplantation procedures and the development of myeloproliferative neoplasias.

KEY PUBLICATIONS

Ho YH, Del Toro R, Rivera-Torres J, ..., Louache F, Andrés V, **Méndez-Ferrer S**. Remodelling of bone marrow hematopoietic stem cell niches promotes myeloid cell expansion during premature or physiological aging. **Cell Stem Cell**, 25(3):407-418.e6.

García-García A, Korn C, García-Fernández M, Domingues O, Villadiego J, Martín-Perez D, Isern J, Bejarano-García JA, Zimmer J, Pérez-Simón JA, Toledo-Aral JJ, Michel T, Airaksinen MS, **Méndez-Ferrer S**. Dual cholinergic signals regulate daily migration of hematopoietic stem cells and leukocytes. **Blood**, 133(3):224-236.

Del Toro R, Chèvre R, Rodríguez C, Ordóñez A, Martínez-González J, Andrés V, **Méndez-Ferrer S**. Nestin(+) cells direct inflammatory cell migration in atherosclerosis. **Nature Communications**. 2016 Sep 2;7:12706.



Schematic summarizing the key cell types and functional features of the central and endosteal bone marrow (BM) niches of haematopoietic stem cells (HSCs).

Credit: Méndez-Ferrer, S., Bonnet, D., Steensma, D.P. et al. Bone marrow niches in haematological malignancies. *Nat Rev Cancer* (2020). <https://doi.org/10.1038/s41568-020-0245-2>

JENNIFER NICHOLS



LAB MEMBERS

Takuya Azami
Elena Corujo-Simon
Kenneth Jones
Tim Lohoff
Sophie Morgani
Arthur Radley
Connor Ross

Embryonic pluripotency

The Nichols lab is interested in how the mammalian embryo is formed and regulated. We scrutinise the roles of transcription factors and signalling pathways involved in establishing and maintaining the pluripotent epiblast (founder of the foetus) during embryogenesis. For this, we employ immunocytochemistry and quantitative confocal analysis as well as single cell transcriptional profiling to complement more traditional experimental tools.

We have developed robust protocols for derivation of embryonic stem cells (ESCs) from mouse embryos, which we utilise as an in vitro proxy to investigate the processes of lineage specification using directed differentiation and various types of organoids and micropattern devices. Much of our current work involves the generation of chimaeras using injection of wild type or genetically modified ESCs into host mouse embryos. This allows us to determine the roles of genes in cell potency, morphology and behaviour in the context of a normal embryonic environment as well as the regulative response of the host embryo as it adapts to the influx of supernumerary cells.

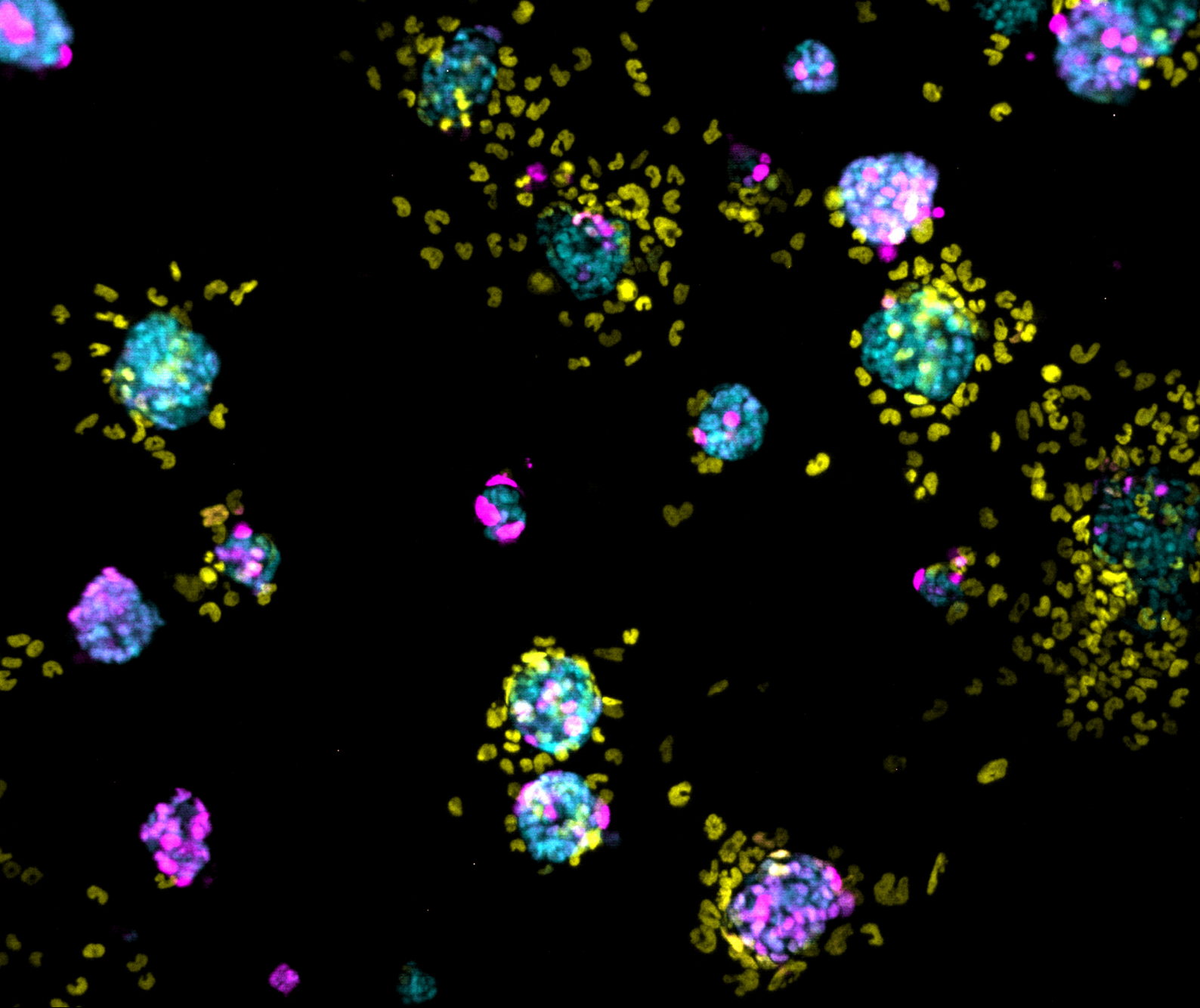
We benefit from the kind donation of spare human embryos from assisted conception programmes to our project for derivation of human ESCs. We are expanding and characterising multiple clonal cell lines to further our understanding of early human development.

KEY PUBLICATIONS

Mulas C, Chia G, Jones KA, Hodgson AC, Stirparo GG, **Nichols J**. Oct4 regulates the embryonic axis and coordinates exit from pluripotency and germ layer specification in the mouse embryo. **Development**. 2018 Jun 18;145(12).

Guo G, von Meyenn F, Santos F, Chen Y, Reik W, Bertone P, Smith A, **Nichols J**. Naïve pluripotent stem cells derived directly from isolated cells of the human inner cell mass. **Stem Cell Reports**. 2016 Apr 12;6(4):437-446.

Alexandrova S, Kalkan T, Humphreys P, Riddell A, Scognamiglio R, Trumpp A, **Nichols J**. Selection and dynamics of embryonic stem cell integration into early mouse embryos. **Development**. 2016 Jan 1;143(1):24-34.



Confocal image of early differentiating human naive embryonic stem cell colonies showing markers for whole epiblast (cyan), naive epiblast (yellow), hypoblast (magenta).

Credit: Nichols lab

ANNA PHILPOTT

Proneural transcription factors



LAB MEMBERS

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Frances Connor
Imogen Duncan
Fani Memi
Lidiya Mykhailechko
Lydia Parkinson
Toshiaki Shigeoka
Laura Woods

We aim to understand how cells adopt a specific fate and to uncover mechanisms that co-ordinate cell cycling with stem cell maintenance and differentiation during development, homeostasis and disease. In particular, we have uncovered a conserved regulatory mechanism where cdk-dependent phosphorylation of multiple proneural proteins promotes maintenance of progenitor/stem status, while dephosphorylation drives differentiation in tissues as diverse as nerve, muscle, pancreas and gut.

Our future aims are three-fold, we will:

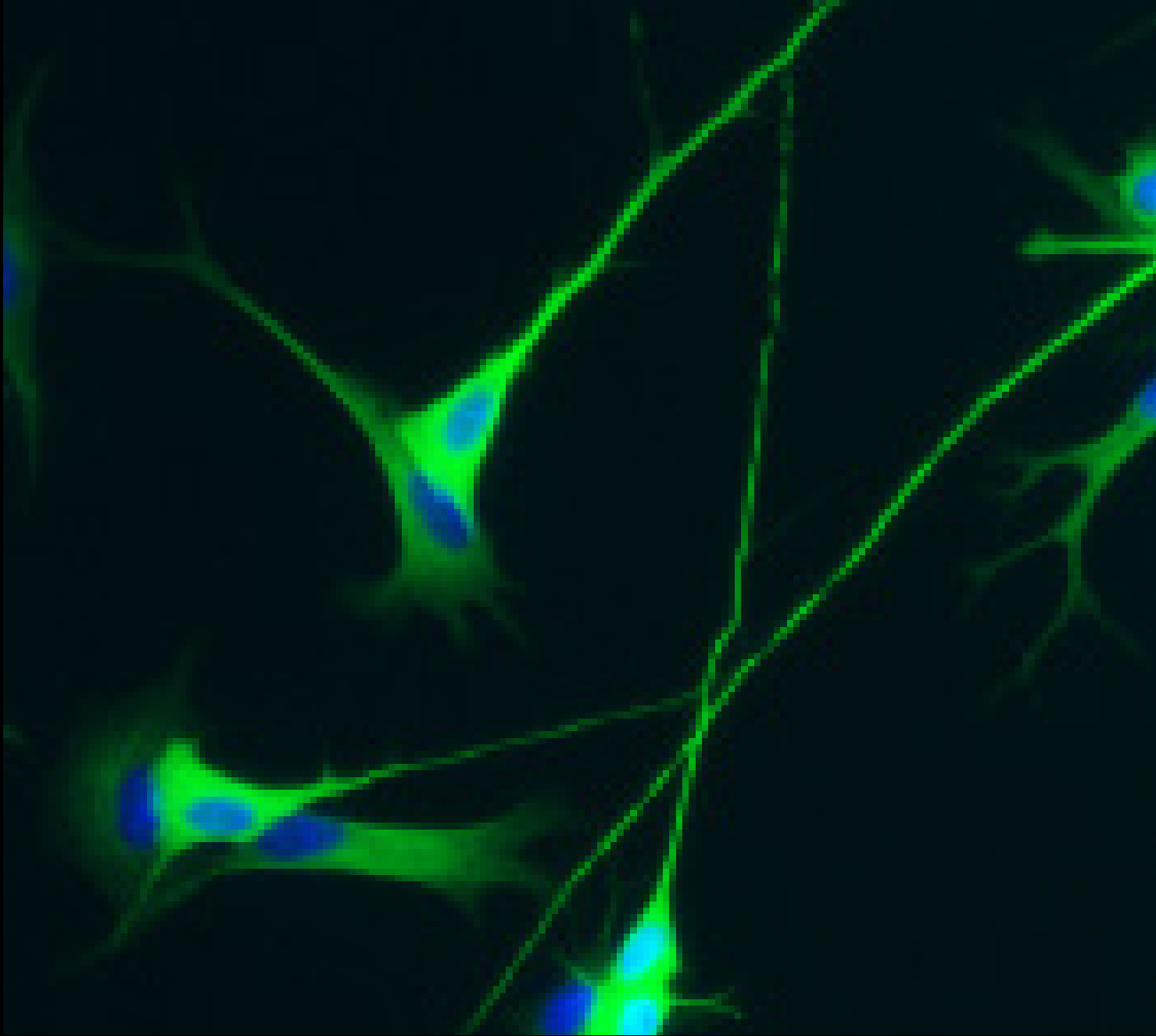
1. Further characterise the molecular mechanisms that link cell cycling and differentiation
2. Investigate perturbation of the balance between stem-ness/progenitor maintenance and differentiation that is a frequent hallmark of multiple cancers, focussing on molecular regulation of proliferation and differentiation in neuroblastoma, glioblastoma and insulinoma, with the aim of developing new therapeutic strategies
3. Probe fundamental mechanisms that determine the fate trajectory and differentiation of different cell types during development focussing on the epigenome and co-factors that control this process at the level of individual cells.

KEY PUBLICATIONS

Tomic G, Morrissey E, Kozar S, Ben-Moshe S, Hoyle A, Azzarelli R, Kemp R, Chilamakuri CSR, Itzkovitz S, **Philpott A**, Winton DJ. Phospho-regulation of ATOH1 Is Required for Plasticity of Secretory Progenitors and Tissue Regeneration. **Cell Stem Cell**. 2018 Sep 6;23(3):436-443.

Azzarelli R, Simons B and **Philpott A**. The developmental ontogeny of neurological cancers: a cellular and molecular framework. **Development**. 2018 May 14;145(10).

Sznurkowska MK, Hannezo E, Rulands S, Nestorowa S, Hindley CJ, Azzarelli R, Nichols J, Göttgens B, Huch M, **Philpott A***, Simons B*. Defining lineage potential and fate behavior of precursors during pancreas development. **Developmental Cell**. Aug 6;46(3):360-375



Neuroblastoma cancer cells stop dividing and differentiate into neurons on treatment with drugs targeting the key transcriptional regulator ASCL1.

Credit: Daniel Marcos-Corchado



LAB MEMBERS

Jingyu Chen
Maurizio Mangolini
Andrew Moore
Eugene Park
Antonella Santoro

Our research group studies the molecular processes underlying the cross-communication between normal and malignant haematopoiesis and their microenvironment. It has recently been shown that CD34+ CD38- HSPCs can contribute to oncogenic transformation into mature B cell malignancies, such as Chronic Lymphocytic Leukemia. Similar to normal cells, tumour cells require signals from their surrounding environment to survive and proliferate. Our work has identified vulnerabilities of cancer cells in this cross-talk, which can be exploited therapeutically. We have identified kinases activated in bone marrow stroma cells, which are essential for tumour-associated inflammation and tumour progression. We are now (1) focussed on defining how tumour-host interactions are modulated along tumour-evolution, from the Cell-of-Origin (COO) to lymphoma-stem cells to therapy-refractory disease. These studies are important to (2) fully understand lymphomagenesis and how the tumour microenvironment contributes to drug resistance and minimal-residual disease (MRD) following systemic anti-cancer therapies.

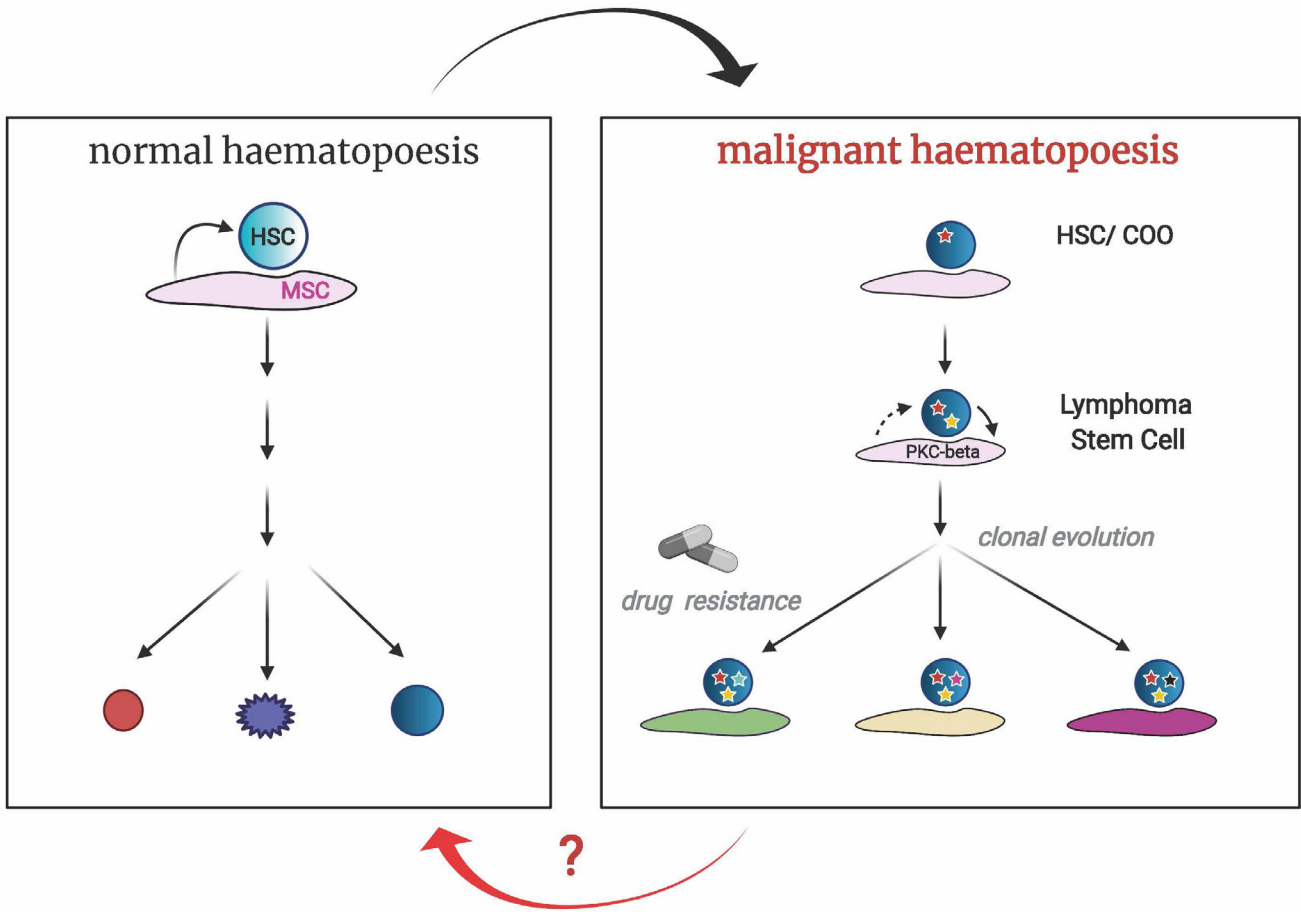
In parallel (3), we study how the tumour-remodelled bone marrow microenvironment affects normal HSC function and contributes to secondary bone marrow failure. Therapeutic interventions with these cross-talks could be exploited to maintain HSC function and thereby eliminate tumour cells, while maintaining and improving immune functions in patients with B cell malignancies.

KEY PUBLICATIONS

Park E, Chen J, Moore A, Mangolini M, Santoro A, Boyd JR, Schjerven, Ecker V, Buchner M, Williamson JC, Lehner PJ, Gasparoli L, Williams O, Bloehdorn J, Stilgenbauer S, Leitges M, Egle A, Schmidt-Supprian M, Fietze S, **Ringshausen I**. Stromal cell protein kinase C-β inhibition enhances chemosensitivity in B cell malignancies and overcomes drug resistance. **Science Translational Medicine**, 2020 Jan 15;12 (526), eaax9340.

Mangolini M, Götte F, Moore A, ..., Hodson DJ, Schmidt-Supprian M, **Ringshausen I**. Notch2 controls non-autonomous Wnt-signalling in chronic lymphocytic leukaemia. **Nature Communications**. 2018 Sep 21;9(1):3839.

Lutzny G, Kocher T, Rudelius M, ..., Peschel C, Egle A, **Ringshausen I**. Proteinkinase C-β dependent activation of NF-κB in stromal cells is indispensable for the survival of chronic lymphocytic leukemia B-cells in vivo. **Cancer Cell**. 2013 Jan 14;23(1):77-92.



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DAVID ROWITCH



LAB MEMBERS

Theresa Bartels
 Harry Bullstrode
 Mark Charlton-Perkins
 Courtney French
 Gemma Girdler
 Staffan Holmqvist
 Ajay Kumar
 Guy Lam
 Katherine Ridley
 John Stockley
 Adrien Vaquie
 Zhaoyang Xu

Glial cells and response to injury

My lab investigates genetic factors that determine development and diversity of glial cells of the brain and the response to injury. Our research to establish functional diversity of astrocytes is funded by Wellcome Trust, and new ERC-funded studies will investigate precise synthetic mechanisms of oligodendrocytes during myelination. We have applied these principles to better understand white matter injury in premature infants, brain cancer, multiple sclerosis and leukodystrophy.

Building on this fundamental research, I led the first human clinical trial of direct neural stem cell transplantation focused on the rare and fatal leukodystrophy, Pelizaeus-Merzbacher Disease (PMD); we are using stem cell biology to decipher the basis for failed myelination in these patients.

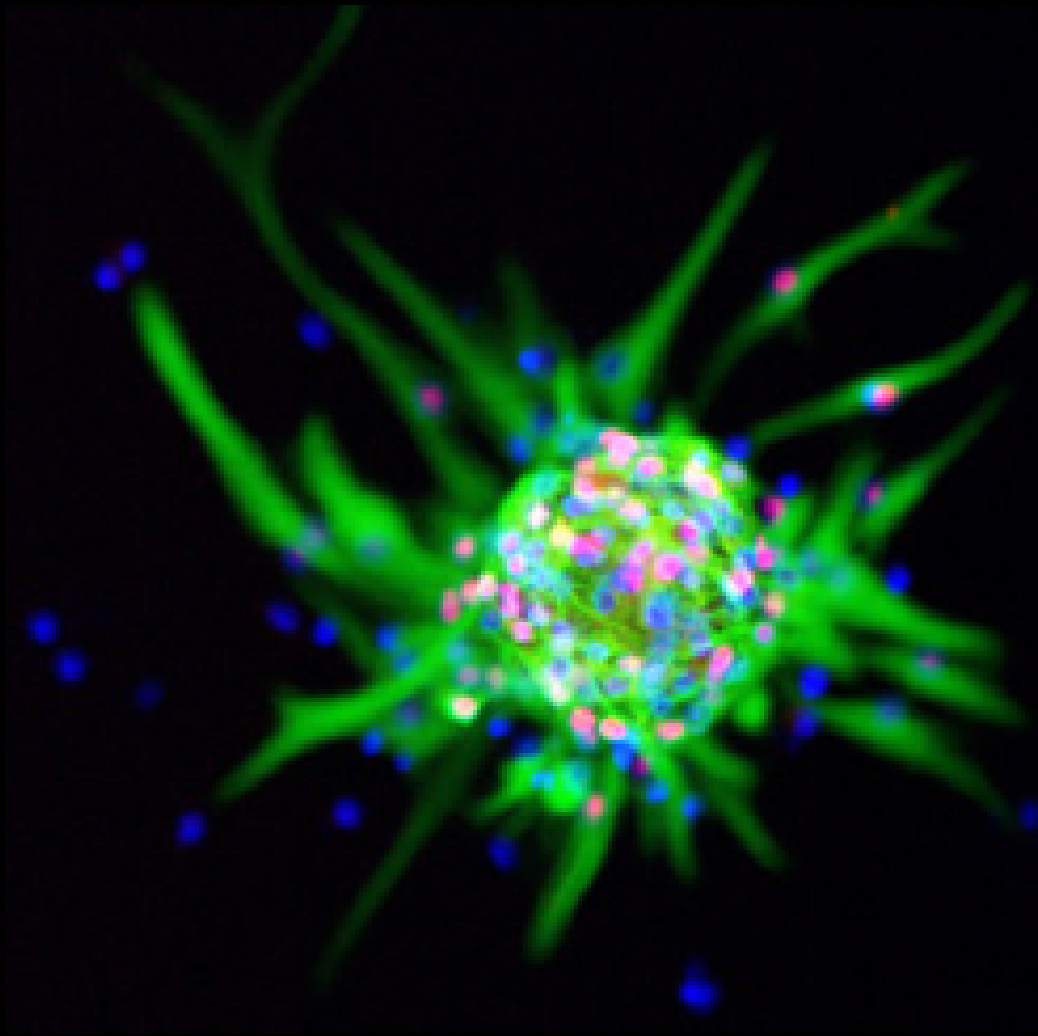
Our interest in precision medicine focuses on applications of genomic technologies to diagnose and better understand the biological basis and rational treatment of rare neurological disorders.

KEY PUBLICATIONS

Bayraktar OA, Bartels T, Holmqvist S, ..., Holt MG, Geschwind DH, **Rowitch DH**. Astrocyte layers in the mammalian cerebral cortex revealed by a single-cell in situ transcriptomic map. **Nature Neuroscience**, 8(7641):427–10.

Schirmer L, Velmeshev D, Holmqvist S, ..., Shioh LR, Kriegstein AR, **Rowitch DH**. Neuronal vulnerability and multilineage diversity in multiple sclerosis. **Nature**. 573 (7772), 75-82.

Nobuta H, Yang N, Ng YH, Marro SG, Sabeur K, Chavali M, Stockley JH, Killilea DW, Walter PB, Zhao C, Huie P, Goldman SA, Kriegstein AR, Franklin RJM, **Rowitch DH**, Wernig M. Oligodendrocyte Death in Pelizaeus-Merzbacher Disease Is Rescued by Iron Chelation. **Cell Stem Cell** 25(4):531-541.e6



Collection neural stem cells stained with nestin (green), and Olig2 (red), a marker of glial progenitors. Such cell collections can “self-organize” to generate specialized cell progeny without external instructions.

Credit: Vivi Heine



LAB MEMBERS

Lawrence Bates
Yael Costa
Imogen Stockwell
Daniel Yamamoto

The research in our lab is focussed on understanding the biology of reprogramming a differentiated cell identity back into a naïve pluripotent stem cell identity, a process known as induced pluripotency. We build on this acquired knowledge to also study the principles governing cell identity change, cell potency, epigenetic regulation and the mechanisms regulating developmental processes taking place in naïve pluripotent stem cells. Our central lines of research are:

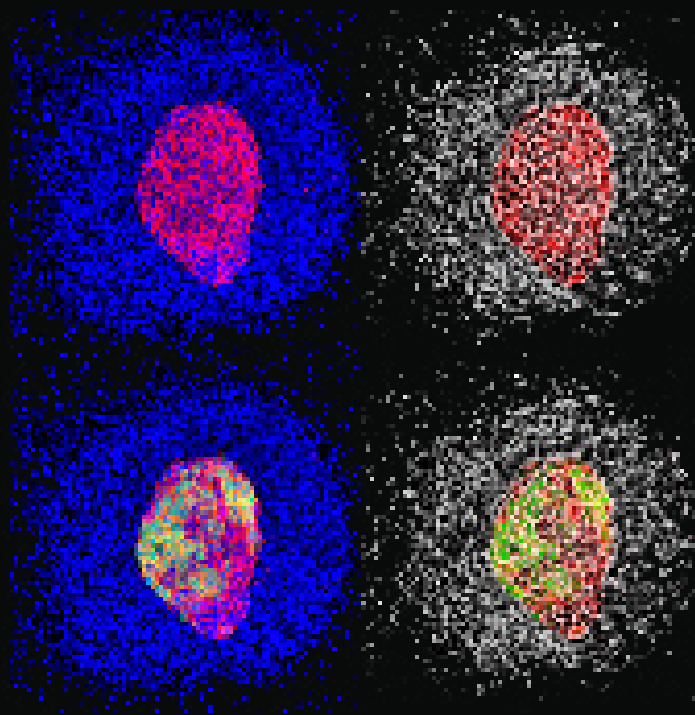
1. Understanding the fundamental biology of nuclear reprogramming. Nuclear reprogramming is a fundamental process in biology and also a great model system to study cell identity change.
2. Determining the potential of programming pluripotent stem cells to defined stem cell types of interest. The creation of bonafide pluripotent stem cells from somatic cells by the use of defined factors has opened up the possibility for the generation of any cell type in the petri dish.
3. Studying the relationship between drivers of reprogramming and epigenetic processes taking place in naïve pluripotent stem cells. Normal development is somewhat a mirror of reprogramming and we are now asking if the drivers of reprogramming also regulate other cellular processes such as the initiation of X-chromosome inactivation.

KEY PUBLICATIONS

Stuart HT, Stirparo GG, Lohoff T, Bates LE, Kinoshita M, Lim CY, Sousa EJ, Maskalenka K, Radzisheuskaya A, Malcolm AA, Alves MRP, Lloyd RL, Nestorowa S, Humphreys P, Mansfield W, Reik W, Bertone P, Nichols J, Göttgens B, **Silva JCR**. Distinct molecular trajectories converge to induce naïve pluripotency. **Cell Stem Cell**, 25(3):388-406.e8

Sousa EJ, Stuart HT, Bates LE, Ghorbani M, Nichols J, Dietmann S, **Silva JCR**. Exit from Naïve Pluripotency Induces a Transient X Chromosome Inactivation-like State in Males. **Cell Stem Cell**. 2018 Jun 1;22(6):919-928.e6.

Santos R, Tosti L, Radzisheuskaya A, Caballero I, Kaji K, Hendrich B, **Silva JCR**. Mbd3/NuRD facilitates induced pluripotency in a context dependent manner. **Cell Stem Cell**. 2014 Jul 3;15(1):102-10.



Panel depicting the appearance of induced pluripotent stem cells (green) from an identified pre-determined to reprogram cell population (red). Non-red cells (grey and blue only) failed to acquire reprogramming competence.

Credit: Chibeza Agley

BEN
SIMONS

Tracing stem cell fate in development, maintenance and disease



LAB MEMBERS

Ignacio Bordeu
Lemonia Chatzeli
Adrien Hallou
Seungmin Han
Tom Hiscock
Daniel Kunz
Min-Kyu Yum

Research in our group combines experimental lineage tracing strategies and single-cell methods with concepts from non-equilibrium statistical physics and mathematics to address the fate behaviour of stem and progenitor cells in the development, maintenance and regeneration of tissues, and factors leading to their dysregulation in diseased states. In particular, we have resolved strategies of stem cell self-renewal in the maintenance of epithelial tissues, including mammalian brain, epidermis, intestine, lung and testis.

Our studies have emphasized the role of stochastic renewal programmes in the regulation of stem cell fate, questioning the nature of stem cell identity and function. We have extended these approaches to study the development and patterning of adult tissues, including the eye, heart, mammary epithelium, pancreas and skin epidermis. At the same, we are collaborating with partner labs to address the cellular basis of tumour initiation.

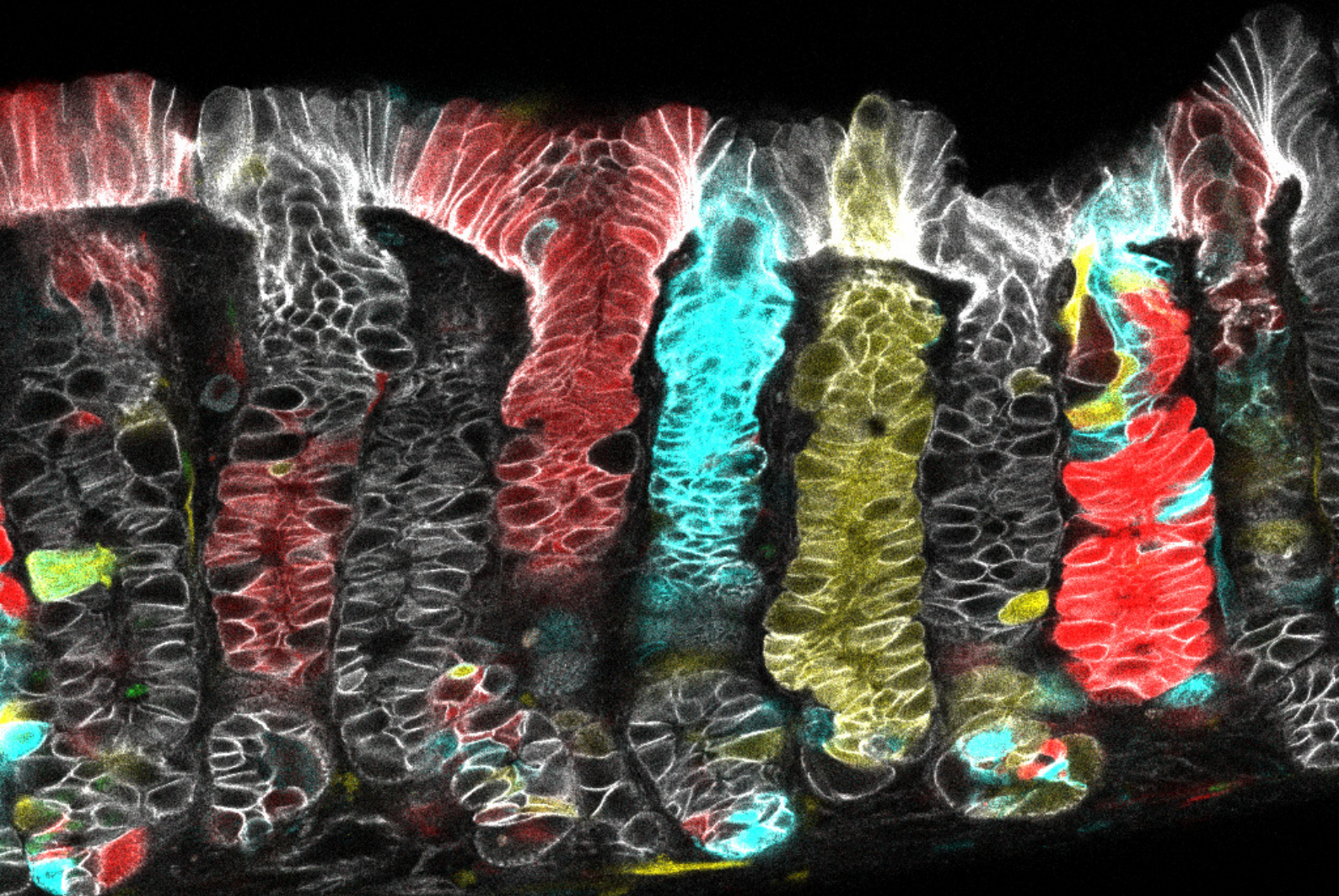
We also work with colleagues at the Babraham Institute to develop statistical approaches to address single cell transcriptional and epigenetic profiling data, with the aim of resolving the factors controlling symmetry breaking and cell fate specification in the developing mouse embryo.

KEY PUBLICATIONS

Han S, Fink J, Jörg DJ, ..., Kim JK, **Simons BD**, Koo BK. Defining the Identity and Dynamics of Adult Gastric Isthmus Stem Cells. **Cell Stem Cell**, 25(3):342-356.e7.

Kitadate Y, Jörg DJ, Tokue M, ..., Takahashi S, **Simons BD**, Yoshida S. Competition for Mitogens Regulates Spermatogenic Stem Cell Homeostasis in an Open Niche. **Cell Stem Cell**, 24(1):79-92.e6.

Hannezo E, Scheele CLGJ, Moad M, Drogo N, Heer R, Sampogna RV, van Rheenen J, **Simons BD**. A Unifying Theory of Branching Morphogenesis. **Cell**, 171(1):242-255.e27.



Mouse stomach corpus glands labelled with a multicolour confetti reporter system.

Credit: Simons Lab

SANJAY SINHA

Cardiovascular disease



Our lab's overall aim is to develop new treatments for cardiovascular diseases, in particular those involving vascular smooth muscle cells and epicardium, using a stem cell based approach.

We have pioneered the generation of embryonic lineage-specific vascular smooth muscle cells, through the lateral mesoderm, paraxial mesoderm, neural crest and epicardium, from human embryonic stem cells (hESC) and induced pluripotent stem cells, using chemically defined conditions. We have utilised this system to model genetically triggered aortopathies, such as Marfan and Loeys-Dietz syndromes. These "disease-in-a-dish" models are being used to understand the pathobiology of these conditions and to screen for new treatments.

Additionally we are testing the regenerative potential of hESC-derived epicardium and other cardiovascular cell types for heart repair after myocardial infarction, either through direct injection or in the form of an *in vitro* generated myocardial "patch".

LAB MEMBERS

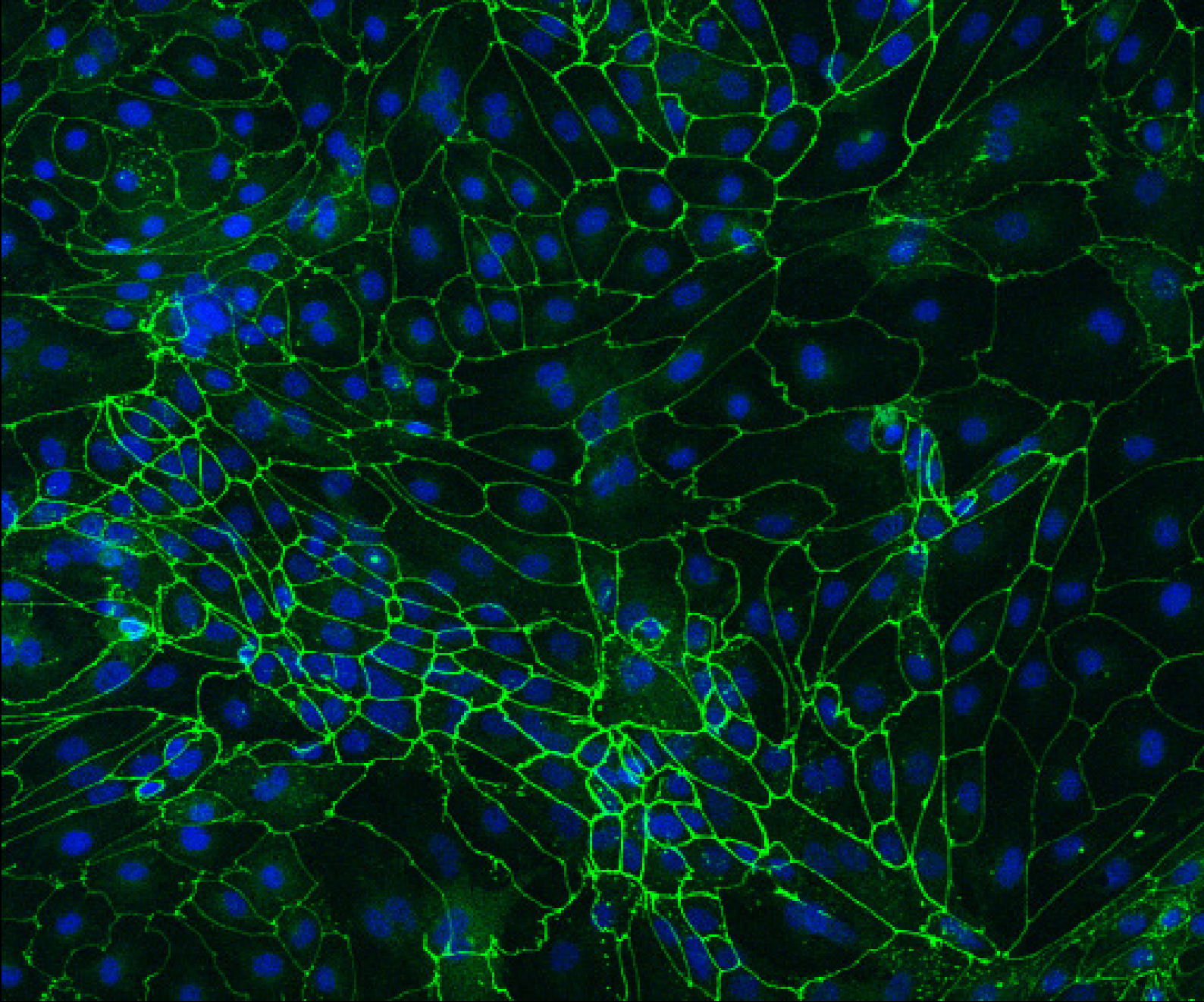
Semih Bayraktar
 Maria Colzani
 Hongorzul Davaapil
 Laure Gambardella
 Peter Holt
 Aishwarya Jacob
 Vincent Knight-Schrijver
 Robyn Macrae
 Ping Ong
 Deborah Passey
 Alex Petchey
 Deeti Shetty

KEY PUBLICATIONS

Gambardella L, McManus S, Moignard V, Sebukhan D, Delaune A, Andrews S, Bernard WG, Morrison M, Riley PR, Göttgens B, Le Novère N, **Sinha S**. BNC1 regulates epicardial heterogeneity and function. **Development** 2019;146 (24) dev174441.

Bargehr J, Ong LP, Colzani M, Davaapil H, Hofsteen P, Bhandari S, Gambardella L, Le Novère N, Iyer D, Sampaziotis F, Weinberger F, Bertero A, Leonard A, Bernard WG, Martinson A, Figg N, Regnier M, Bennett M, Murry CE, and **Sinha S**. Human Embryonic Stem Cell-Derived Epicardial Cells Augment Cardiomyocyte-Driven Heart Regeneration. **Nature Biotechnology** 2019;37:895-906.

Granata A, Serrano F, Bernard WG, McNamara M, Low L, Sastry P, and **Sinha S**. An iPSC-derived vascular model of Marfan syndrome identifies key mediators of smooth muscle cell death. **Nature Genetics**. 2017;49:97-109.



Epicardial cells generated from human embryonic stem cells form a sheet, also known as mesothelium. The tight junction protein1 (ZO1) is seen at cell peripheries (green) and contributes to the cell-cell junctions.

Credit: Laure Gambardella

AUSTIN SMITH

Pluripotent stem cell biology



We study pluripotent stem cells derived from early embryos or generated by somatic cell reprogramming. These cell lines harbour the potential to generate all somatic cell types.

Our goal is to understand how pluripotent stem cells maintain broad developmental potency and how they prepare for and make cell fate decisions. We compare pluripotent cells from different mammals to elucidate common principles and species-specific adaptations.

Our research focusses on the developmental origins and plasticity of the pluripotency gene regulatory network. We seek to expose the molecular logic governing early development, pluripotency transitions, stem cell self-renewal, and lineage potential.

LAB MEMBERS

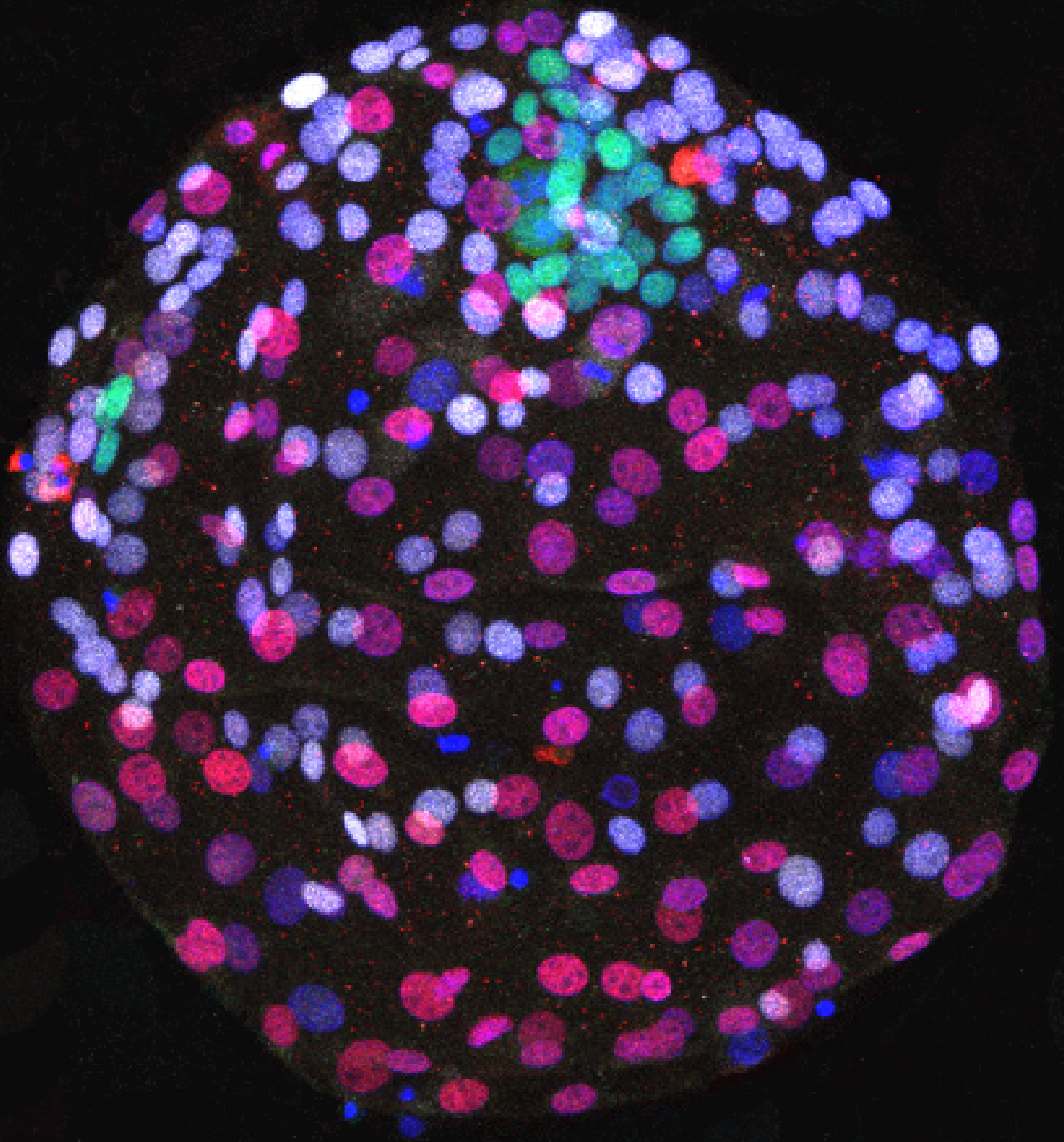
James Clarke
Anish Dattani
Rosalind Drummond
Ge Guo
Tao Huang
Masaki Kinoshita
Arthur Radley
Giuliano Stirparo
Ayaka Yanagida

KEY PUBLICATIONS

Guo G, Stirparo GG, Strawbridge S, Yang J, Clarke J, Li MA, Myers S, Özel BN, Nichols J, **Smith A**. Trophectoderm Potency is Retained Exclusively in Human Naïve Cells. **bioRxiv**, 2020.2002.2004.933812.

Kalkan T, Bornelöv S, Mulas C, Diamanti E, Lohoff T, Ralser M, Middelkamp S, Lombard P, Nichols J, **Smith A**. Complementary Activity of ETV5, RBPJ and TCF3 Drives Formative Transition from Naïve Pluripotency. **Cell Stem Cell**, 24(5):785-801.e7

Smith A. Formative pluripotency: the executive phase in a developmental continuum. **Development**. 2017; 144: 365-373.



Sheep blastocyst regenerated from an isolated inner cell mass. Immunostaining shows lineage-specific transcription factors: SOX2 (epiblast, green); GATA3 (trophoblast, red); SOX17 (hypoblast, white). Nuclei are visualised with Dapi (blue).

Credit: Ge Guo

LUDOVIC VALLIER



LAB MEMBERS

Teresa Brevini
Stephanie Brown
Dominika Dziedzicka
Vasileios Galanakis
Chris Gribben
Fatima Lugtu
Carola Morell
Shota Nakanoh
Anna Osnato
Marion Perrin
Laetitia Pinte
Alexander Ross
Fotios Sampaziotis
Rute Tomaz
Marta Vila-Gonzalez
Brandon Wesley
Katherina Zacharis

Mechanisms controlling differentiation of pluripotent stem cells into definitive endoderm

Understanding the mechanisms controlling early cell fate decisions in human development has major implications for regenerative medicine. Indeed the generation of fully functional cell types from stem cells is only achievable by recapitulating a natural succession of cell fate choice. The first event of differentiation of the embryo proper occurs at the stage of gastrulation with the specification of the three primary germ layers ectoderm, mesoderm and endoderm, from which all the cells of adult tissues are derived.

The main objective of our group is to define the molecular mechanisms controlling the transition between pluripotency and the endoderm lineage. For that, we use human pluripotent stem cells (hESCs and hiPSCs) as *in vitro* model of development to study the interplay between transcriptional networks, epigenetic modifications and cell cycle which ultimately orchestrate the earliest step of differentiation. The resulting knowledge allows the development of new culture systems to drive differentiation of pluripotent stem cells into pancreatic, hepatic, lung and gut cells. These cells are then used to model disease *in vitro*, especially metabolic disorders affecting the liver.

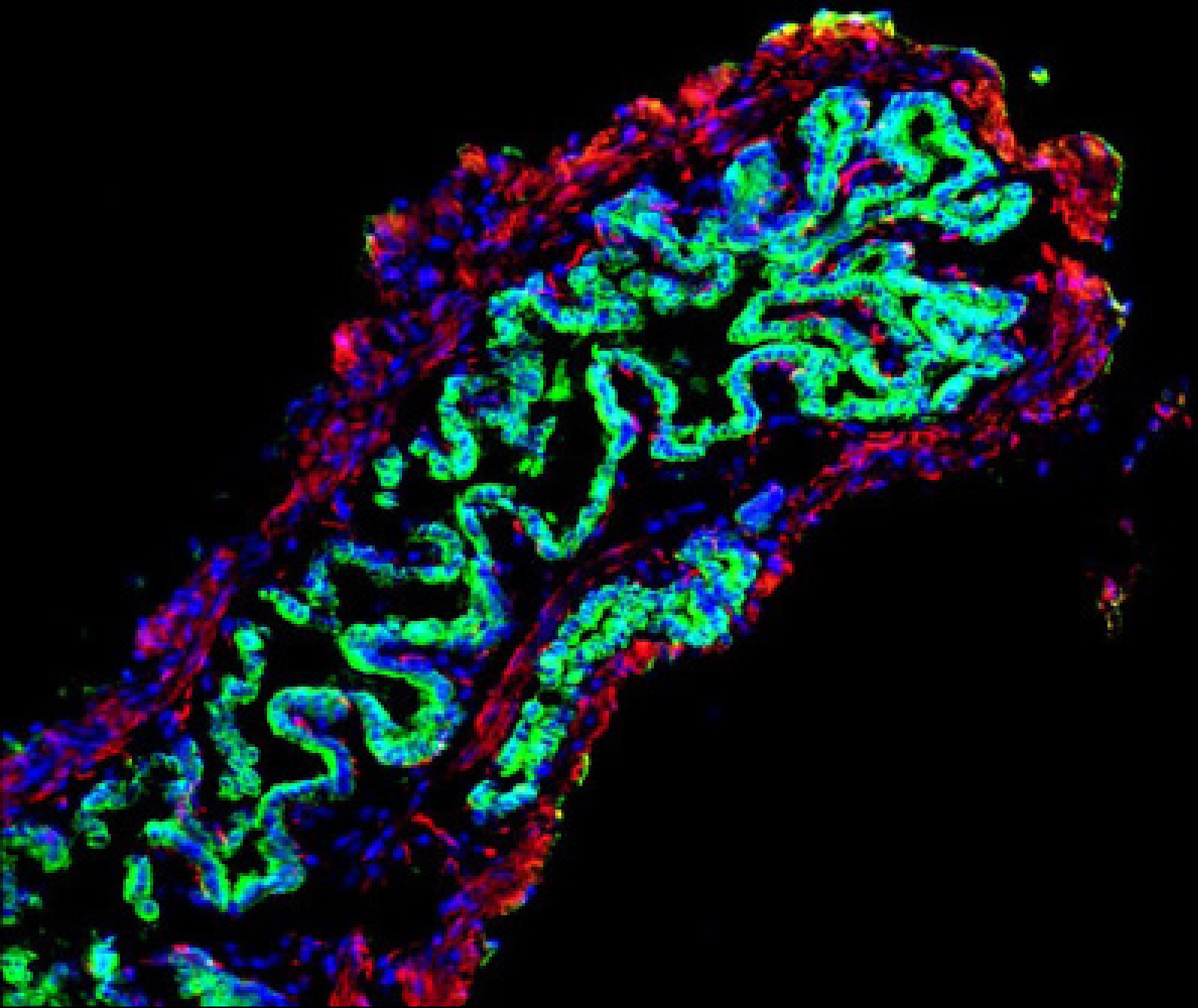
Furthermore, we are currently investigating how similar mechanisms could regulate adult stem cells self-renewal /differentiation during organ regeneration. Overall, our objective is to understand basic mechanisms of differentiation to generate cells for a diversity of clinical applications including disease modelling and cell based therapy.

KEY PUBLICATIONS

Yiangou L, Grandy RA, Morell CM, ..., Simonic I, Sinha S, **Vallier L**. Method to Synchronize Cell Cycle of Human Pluripotent Stem Cells without Affecting Their Fundamental Characteristics. **Stem Cell Reports**, 12(1):165-179.

Segeritz CP, Rashid ST, Cardoso de Brito M, ..., Gooptu B, Lomas DA, **Vallier L**. hiPSC hepatocyte model demonstrates the role of unfolded protein response and inflammatory networks in α 1-antitrypsin deficiency. **Journal of Hepatology**, 69 (4), 851-860.

Bertero A, Brown S, Madrigal P, ..., Stunnenberg HG, Mendjan S, **Vallier L**. The SMAD2/3 interactome reveals that TGF β controls m6A mRNA methylation in pluripotency. **Nature**, 555(7695):256-259.



Mouse gallbladder following repair with human cholangiocytes organoids (green).

Credit: Fotis Sampaziotis

GEORGE VASSILIOU



LAB MEMBERS

Clea Barcena
Jonathan Cooper
Aristi Damaskou
Filipa Domingues
Monika Dudek
Margarete Fabre
Dimitris Garyfallos
Gonia Gozdecka
Muxin Gu
Suruchi Pacharne
Pedro Quiros
Justyna Rak
Thomas Sheahan

Leukaemic haemopoietic stem cells

Our group seeks to understand the cell-autonomous and cell-non-autonomous processes involved in transformation of normal to leukaemic haemopoietic stem cells and to identify genetic vulnerabilities of myeloid malignancies that can be exploited as targets of novel anti-leukaemic therapies.

To achieve these aims the group uses three main approaches:

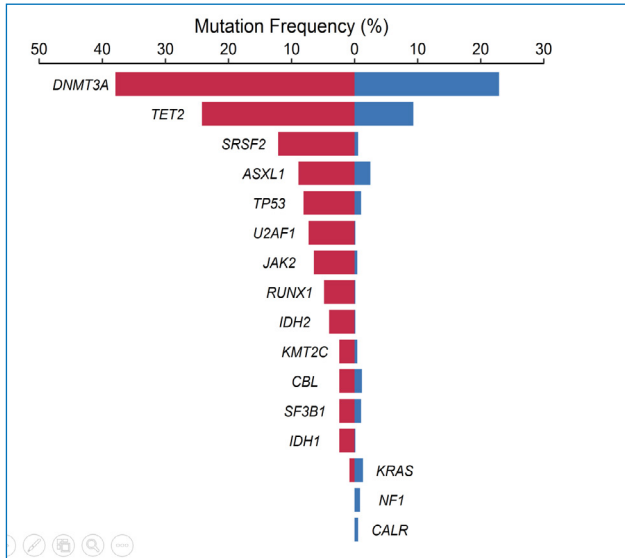
1. Application of genetic screens to identify and investigate genetic vulnerabilities of acute myeloid leukaemia and related cancers in order to develop new therapeutic approaches;
2. Generation and study of bespoke mouse models of somatic mutations driving human myeloid malignancies, in order to define their molecular, genomic and phenotypic effects on haemopoietic stem and progenitor cells and to develop leukaemic models for study in translational studies;
3. Detection and tracking of the evolution of clonal haematopoiesis in healthy individuals, in order to understand the factors involved in leukaemic progression and develop new approaches for early detection.

KEY PUBLICATIONS

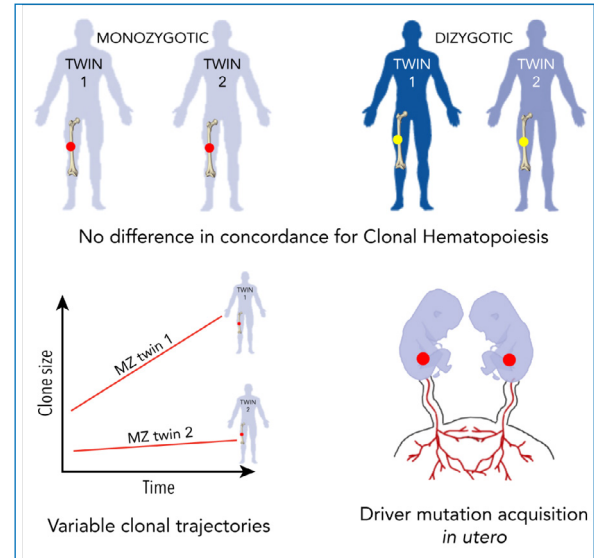
Abelson S, Collord G, Ng SWK, ..., Gerstung M, Dick JE, Brennan P, **Vassiliou GS**, Shlush LI. Prediction of acute myeloid leukaemia risk in healthy individuals. **Nature**. 2018 Jul;559(7714):400-404.

Gozdecka M, Meduri E, Mazan M, ..., Adams D, **Vassiliou GS**, Huntly BJP. UTX-mediated enhancer and chromatin remodeling suppresses myeloid leukemogenesis through noncatalytic inverse regulation of ETS and GATA programs. **Nature Genetics**. 2018 Jun;50(6):883-894.

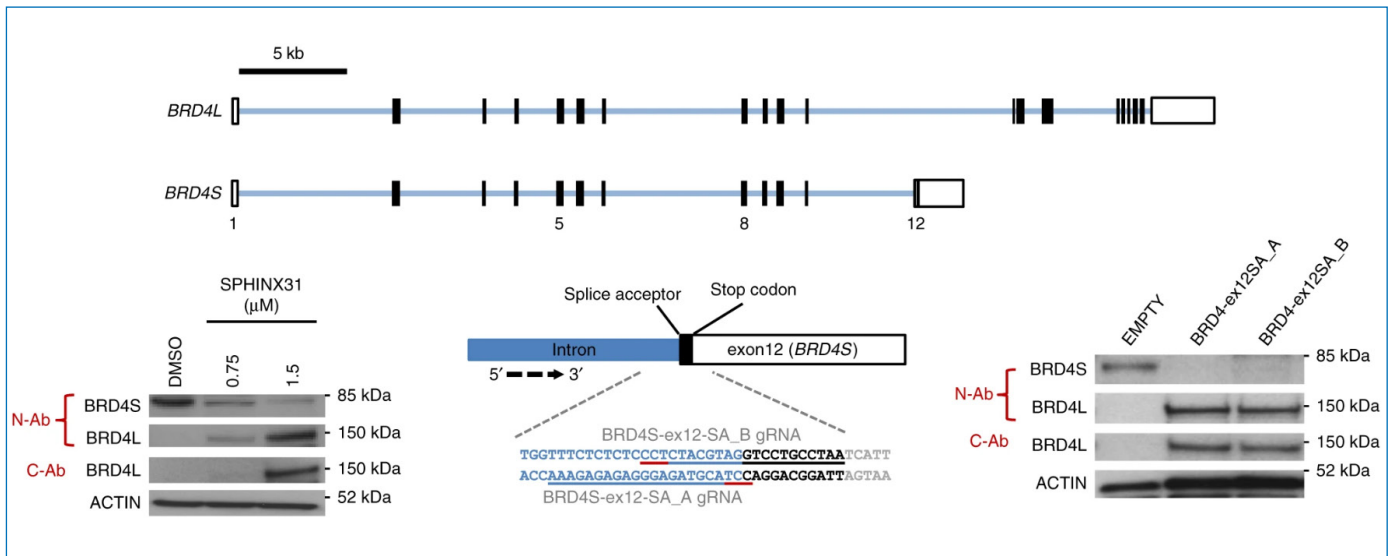
Barbieri I, Tzelepis K, Pandolfini L, Shi J, Millan-Zambrano G, Robson SC, Aspris D, Migliori V, Bannister AJ, Han N, De Braekeleer E, Ponstingl H, Hendrick A, Vakoc CR, **Vassiliou GS** & Kouzarides T. Promoter-bound METTL3 maintains myeloid leukaemia by m(6)A-dependent translation control. **Nature**. 2017 Dec 7;552(7683):126-131.



Prediction of AML development on healthy individuals
(Abelson, Collord et al, Nature 2018)



Characteristics of clonal haematopoiesis in identical twins
(Fabre et al, Blood 2020)



New therapeutic targets in AML: SRPK1 inhibition alters splicing of BRD4
(Tzelepis et al, Nature Comms 2018)

Alumni

DAVID
KENT

Single cell fate choice in normal and malignant stem cells



In 2019, David Kent moved his lab to the University of York and the York Biomedical Research Institute to take up a position as Reader in Haematology and continue his research in the field.

The Kent lab focuses on how cell fate decisions are made on a single cell level in an effort to understand how to expand stem cell populations outside the body (for cell replacement or as a cell source for gene therapy) and how subversion of this process leads to cancer.

Key research themes under investigation in the lab are:

1. The molecular drivers of stem cell heterogeneity (self-renewal durability, lineage commitment)
2. The physical and quantitative biology of stem cells (mechanical signalling, mathematical modelling)
3. The early stages of cancer evolution from single cells (myeloproliferative neoplasms and myelodysplastic syndromes)
4. The role of the immune cell microenvironment in disease evolution

Areas of particular interest include normal stem cell fate choice, clonal evolution of myeloid malignancies, physical biology of stem cells, and tools/approaches for expanding blood stem cells outside the body.

Research Publications

Institute researchers produced 166 publications in 2019 across a range of topics and research themes.

Conejós-Sánchez I, Gallon E, Niño-Pariente A, Smith JA, De la Fuente AG, Di Canio L, Pluchino S, **Franklin RJM**, Vicent MJ. Polyornithine-based polyplexes to boost effective gene silencing in CNS disorders. *Nanoscale*. 2020 Mar 21;12(11):6285-6299. Epub 2019 Dec 16.

Vuono R, Kouli A, Legault EM, Chagnon L, Allinson KS, La Spada A; REGISTRY Investigators of the European Huntington's Disease Network, Biunno I, **Barker RA**, Drouin-Ouellet J. Association Between Toll-Like Receptor 4 (TLR4) and Triggering Receptor Expressed on Myeloid Cells 2 (TREM2) Genetic Variants and Clinical Progression of Huntington's Disease. *Mov Disord*. 2020 Mar;35(3):401-408. Epub 2019 Nov 14.

Coll AP, Chen M, Taskar P, Rimmington D, Patel S, Tadross JA, Cimino I, Yang M, Welsh P, Virtue S, Goldspink DA, Miedzybrodzka EL, Konopka AR, Esponda RR, Huang JT, Tung YCL, Rodriguez-Cuenca S, Tomaz RA, Harding HP, Melvin A, Yeo GSH, Preiss D, Vidal-Puig A, **Vallier L**, Nair KS, Wareham NJ, Ron D, Gribble FM, Reimann F, Sattar N, Savage DB, Allan BB, O'Rahilly S. GDF15 mediates the effects of metformin on body weight and energy balance. *Nature*. 2020 Feb;578(7795):444-448. Epub 2019 Dec 25.

Solti K, Kuan WL, F6rizs B, Kustos G, Mih6ly J, Varga Z, Herberth B, Moravcsik 6, Kiss R, K6rpati M, Mikes A, Zhao Y, Imre T, Rochet JC, Aigbirhio F, Williams-Gray CH, **Barker RA**, T6th G. DJ-1 can form β -sheet structured aggregates that co-localize with pathological amyloid deposits. *Neurobiol Dis*. 2020 Feb;134:104629. Epub 2019 Nov 11.

Fabre MA, McKerrell T, Zwiebel M, Vijayabaskar MS, Park N, Wells PM, Rad R, Deloukas P, Small K, Steves CJ, **Vassiliou GS**. Concordance for clonal hematopoiesis is limited in elderly twins. *Blood*. 2020 Jan 23;135(4):269-273.

Mayer D, Stadler MB, Rittirsch M, Hess D, Lukonin I, Winzi M, **Smith A**, Buchholz F, Betschinger J. Zfp281 orchestrates interconversion of pluripotent states by engaging Ehmt1 and Zic2. *EMBO J*. 2020 Jan 15;39(2):e102591. Epub 2019 Nov 29.

Ho YH, **M6ndez-Ferrer S**. Microenvironmental contributions to hematopoietic stem cell aging. *Haematologica*. 2020 Jan;105(1):38-46. Epub 2019 Dec 5.

Hardison RC, Zhang Y, Keller CA, Xiang G, Heuston EF, An L, Lichtenberg J, Giardine BM, Bodine D, Mahony S, Li Q, Yue F, Weiss MJ, Blobel GA, Taylor J, Hughes J, Higgs DR, **G6ttgens B**. Systematic integration of GATA transcription factors and epigenomes via IDEAS paints the regulatory landscape of hematopoietic cells. *IUBMB Life*. 2020 Jan;72(1):27-38. Epub 2019 Nov 25.

Fitch SR, Kapeni C, Tsitsopoulou A, Wilson NK, **G6ttgens B**, de Bruijn MF, Ottersbach K. Gata3 targets Runx1 in the embryonic haematopoietic stem cell niche. *IUBMB Life*. 2020 Jan;72(1):45-52. Epub 2019 Oct 21.

MacPherson L, Anokye J, Yeung MM, Lam EYN, Chan YC, Weng CF, Yeh P, Knezevic K, Butler MS, Hoegl A, Chan KL, Burr ML, Gearing LJ, Willson T, Liu J, Choi J, Yang Y, Bilardi RA, Falk H, Nguyen N, Stuppel PA, Peat TS, Zhang M, de Silva M, Carrasco-Pozo C, Avery VM, Khoo PS, Dolezal O, Dennis ML, Nuttall S, Surjadi R, Newman J, Ren B, Leaver DJ, Sun Y, Baell JB, Dovey O, **Vassiliou GS**, Grebien F, Dawson SJ, Street IP, Monahan BJ, Burns CJ, Choudhary C, Blewitt ME, Voss AK, Thomas T, Dawson MA. HBO1 is required for the maintenance of leukaemia stem cells. *Nature*. 2020 Jan;577(7789):266-270. Epub 2019 Dec 11.

Fourrier A, Delbos F, Menoret S, Collet C, Thi Thuy LT, Myara A, Petit F, Tolosa L, Laplanche S, G6mez-Lech6n MJ, Labrune P, Aneg6n I, **Vallier L**, Garnier D, Nguyen TH. Regenerative cell therapy for the treatment of hyperbilirubinemic Gunn rats with fresh and frozen human induced pluripotent stem cells-derived hepatic stem cells. *Xenotransplantation*. 2020 Jan;27(1):e12544. Epub 2019 Jul 25.

Vassiliou GS. The curious incident of TdT-mediated mutations in AML. *Blood*. 2019 Dec 19;134(25):2229-2231.

Fukushima T, Tanaka Y, Hamey FK, Chang CH, Oki T, Asada S, Hayashi Y, Fujino T, Yonezawa T, Takeda R, Kawabata KC, Fukuyama T, Umemoto T, Takubo K, Takizawa H, Goyama S, Ishihama Y, Honda H, **Göttgens B**, Kitamura T. Discrimination of Dormant and Active Hematopoietic Stem Cells by G(0) Marker Reveals Dormancy Regulation by Cytoplasmic Calcium. *Cell Rep*. 2019 Dec 7;29(12):4144-4158.e7.

Gambardella L, McManus SA, Moignard V, Sebukhan D, Delaune A, Andrews S, Bernard WG, Morrison MA, Riley PR, **Göttgens B**, Gambardella Le Novère N, **Sinha S**. BNC1 regulates cell heterogeneity in human pluripotent stem cell-derived epicardium. *Development*. 2019 Dec 13;146(24).

Sasca D, Yun H, Giotopoulos G, Szybinski J, Evan T, Wilson NK, Gerstung M, Gallipoli P, **Green AR**, Hills R, Russell N, Osborne CS, Papaemmanuil E, **Göttgens B**, Campbell P, **Huntly BJP**. Cohesin-dependent regulation of gene expression during differentiation is lost in cohesin-mutated myeloid malignancies. *Blood*. 2019 Dec 12;134(24):2195-2208.

McMurrin CE, Guzman de la Fuente A, Penalva R, Ben Menachem-Zidon O, Dombrowski Y, Falconer J, Gonzalez GA, Zhao C, Krause FN, Young AMH, Griffin JL, Jones CA, Hollins C, Heimesaat MM, Fitzgerald DC, **Franklin RJM**. The microbiota regulates murine inflammatory responses to toxin-induced CNS demyelination but has minimal impact on remyelination. *Proc Natl Acad Sci U S A*. 2019 Dec 10;116(50):25311-25321.

Bredenkamp N, Yang J, Clarke J, Stirparo GG, von Meyenn F, Dietmann S, Baker D, Drummond R, Ren Y, Li D, Wu C, Rostovskaya M, Eminli-Meissner S, **Smith A**, Guo G. Wnt Inhibition Facilitates RNA-Mediated Reprogramming of Human Somatic Cells to Naive Pluripotency. *Stem Cell Reports*. 2019 Dec 10;13(6):1083-1098.

Padella A, Simonetti G, Paciello G, Giotopoulos G, Baldazzi C, Righi S, Ghetti M, Stengel A, Guadagnuolo V, De Tommaso R, Papayannidis C, Robustelli V, Franchini E, Rorà AGLD, Ferrari A, Fontana MC, Bruno S, Ottaviani E, Soverini S, Storlazzi CT, Haferlach C, Sabatini E, Testoni N, Iacobucci I, **Huntly BJP**, Ficarra E, Martinelli G. Novel and Rare Fusion Transcripts Involving Transcription Factors and Tumor Suppressor Genes in Acute Myeloid Leukemia. *Cancers (Basel)*. 2019 Dec 5;11(12).

Barral A, Rollan I, Sanchez-Iranzo H, Jawaid W, Badia-Careaga C, Menchero S, Gomez MJ, Torroja C, Sanchez-Cabo F, **Göttgens B**, Manzanares M, Sainz de Aja J. Nanog regulates Pou3f1 expression at the exit from pluripotency during gastrulation. *Biol Open*. 2019 Dec 2;8(11).

Van Steenwinckel J, Schang AL, Krishnan ML, Degos V, Delahaye-Duriez A, Bokobza C, Csaba Z, Verdonk F, Montané A, Sigaut S, Hennebert O, Lebon S, Schwendimann L, Le Charpentier T, Hassan-Abdi R, Ball G, Aljabar P, Saxena A, Holloway RK, Birchmeier W, Baud O, **Rowitch D**, Miron V, Chretien F, Leconte C, Besson VC, Petretto EG, Edwards AD, Hagberg H, Soussi-Yanicostas N, Fleiss B, Gressens P. Decreased microglial Wnt/ β -catenin signalling drives microglial pro-inflammatory activation in the developing brain. *Brain*. 2019 Dec 1;142(12):3806-3833.

Lopez CK, Noguera E, Stavropoulou V, Robert E, Aid Z, Ballerini P, Bilhou-Nabera C, Lapillonne H, Boudia F, Thirant C, Fagnan A, Arcangeli ML, Kinston SJ, Diop M, Job B, Lecluse Y, Brunet E, Babin L, Villeval JL, Delabesse E, Peters AHFM, Vainchenker W, Gaudry M, Masetti R, Locatelli F, Malinge S, Nerlov C, Droin N, Lobry C, Godin I, Bernard OA, **Göttgens B**, Petit A, Pflumio F, Schwaller J, Mercher T. Ontogenic Changes in Hematopoietic Hierarchy Determine Pediatric Specificity and Disease Phenotype in Fusion Oncogene-Driven Myeloid Leukemia. *Cancer Discov*. 2019 Dec;9(12):1736-1753.

Gordon GSD, Joseph J, **Alcolea MP**, Sawyer T, Williams C, Fitzpatrick CRM, Jones PH, di Pietro M, Fitzgerald RC, Wilkinson TD, Bohndiek SE. Quantitative phase and polarization imaging through an optical fiber applied to detection of early esophageal tumorigenesis. *J Biomed Opt*. 2019 Dec;24(12):1-13.

Ellis JK, Sorrells SF, Mikhailova S, Chavali M, Chang S, Sabour K, Mcquillen P, **Rowitch DH**. Ferret brain possesses young interneuron collections equivalent to human postnatal migratory streams. *J Comp Neurol*. 2019 Dec 1;527(17):2843-2859.

Neumann B, Segel M, **Chalut KJ**, **Franklin RJ**. Remyelination and ageing: Reversing the ravages of time. *Mult Scler*. 2019 Dec;25(14):1835-1841.

Argelaguet R, Clark SJ, Mohammed H, Stapel LC, Krueger C, Kapourani CA, Imaz-Rosshandler I, Lohoff T, Xiang Y, Hanna CW, Smallwood S, Ibarra-Soria X, Buettner F, Sanguinetti G, Xie W, Krueger F, **Göttgens B**, Rugg-Gunn PJ, Kelsey G, Dean W, **Nichols J**, Stegle O, Marioni JC, Reik W. Multi-omics profiling of mouse gastrulation at single-cell resolution. *Nature*. 2019 Dec;576(7787):487-491.

Papaioannou D, Petri A, Dovey OM, Terreri S, Wang E, Collins FA, Woodward LA, Walker AE, Nicolet D, Pepe F, Kumchala P, Bill M, Walker CJ, Karunasiri M, Mrózek K, Gardner ML, Camilotto V, Zitzer N, Cooper JL, Cai X, Rong-Mullins X, Kohlschmidt J, Archer KJ, Freitas MA, Zheng Y, Lee RJ, Aifantis I, **Vassiliou G**, Singh G, Kauppinen S, Bloomfield CD, Dorrance AM, Garzon R. The long non-coding RNA HOXB-AS3 regulates ribosomal RNA transcription in NPM1-mutated acute myeloid leukemia. *Nat Commun*. 2019 Nov 25;10(1):5351.

Yiangou L, Grandy RA, Osnato A, Ortmann D, **Sinha S**, **Vallier L**. Cell cycle regulators control mesoderm specification in human pluripotent stem cells. *J Biol Chem*. 2019 Nov 22;294(47):17903-17914.

Kuan WL, Stott K, He X, Wood TC, Yang S, Kwok JCF, Hall K, Zhao Y, Tietz O, Aigbirhio FI, Vernon AC, **Barker RA**. Systemic α -synuclein injection triggers selective neuronal pathology as seen in patients with Parkinson's disease. *Mol Psychiatry*. 2019 Nov 22. doi: 10.1038/s41380-019-0608-9.

Huefner A, Kuan WL, Mason SL, Mahajan S, **Barker RA**. Serum Raman spectroscopy as a diagnostic tool in patients with Huntington's disease. *Chem Sci*. 2019 Nov 14;11(2):525-533.

Xie SZ, Garcia-Prat L, Voisin V, Ferrari R, Gan OI, Wagenblast E, Kaufmann KB, Zeng AGX, Takayanagi SI, Patel I, Lee EK, Jargstorf J, Holmes G, Romm G, Pan K, Shoong M, VEDI A, Luberto C, Minden MD, Bader GD, **Laurenti E**, Dick JE. Sphingolipid Modulation Activates Proteostasis Programs to Govern Human Hematopoietic Stem Cell Self-Renewal. *Cell Stem Cell*. 2019 Nov 7;25(5):639-653. e7.

Mowforth OD, Davies BM, **Kotter MR**. Quality of Life Among Informal Caregivers of Patients With Degenerative Cervical Myelopathy: Cross-Sectional Questionnaire Study. *Interact J Med Res*. 2019 Nov 7;8(4):e12381.

Krieger TG, Moran CM, Frangini A, Visser WE, Schoenmakers E, Muntoni F, Clark CA, Gadian D, Chong WK, Kuczynski A, Dattani M, Lyons G, Efthymiadou A, Varga-Khadem F, **Simons BD**, Chatterjee K, Livesey FJ. Mutations in thyroid hormone receptor $\alpha 1$ cause premature neurogenesis and progenitor cell depletion in human cortical development. *Proc Natl Acad Sci U S A*. 2019 Nov 5;116(45):22754-22763.

Vijayabaskar MS, Goode DK, Obier N, Lichtinger M, Emmett AML, Abidin FNZ, Shar N, Hannah R, Assi SA, Lie-A-Ling M, **Göttgens B**, Lacaud G, Kouskoff V, Bonifer C, Westhead DR. Identification of gene specific cis-regulatory elements during differentiation of mouse embryonic stem cells: An integrative approach using high-throughput datasets. *PLoS Comput Biol*. 2019 Nov 4;15(11):e1007337.

McPherson S, Greenfield G, Andersen C, Grinfeld J, Hasselbalch HC, Nangalia J, Mills KI, McMullin MF. Methylation age as a correlate for allele burden, disease status, and clinical response in myeloproliferative neoplasm patients treated with vorinostat. *Exp Hematol*. 2019 Nov;79:26-34.

Baker AM, Gabbutt C, Williams MJ, Cereser B, Jawad N, Rodriguez-Justo M, Jansen M, Barnes CP, **Simons BD**, McDonald SA, Graham TA, Wright NA. Crypt fusion as a homeostatic mechanism in the human colon. *Gut*. 2019 Nov;68(11):1986-1993.

Saçma M, Pospiech J, Bogeska R, de Back W, Mallm JP, Sakk V, Soller K, Marka G, Vollmer A, Karns R, Cabezas-Wallscheid N, Trumpp A, **Méndez-Ferrer S**, Milsom MD, Mulaw MA, Geiger H, Florian MC. Haematopoietic stem cells in perisinusoidal niches are protected from ageing. *Nat Cell Biol*. 2019 Nov;21(11):1309-1320.

Aloia L, McKie MA, Vernaz G, Cordero-Espinoza L, Aleksieva N, van den Aamele J, Antonica F, Font-Cunill B, Raven A, Aiese Cigliano R, Belenguer G, Mort RL, Brand AH, Zernicka-Goetz M, Forbes SJ, Miska EA, Huch M. Epigenetic remodelling licences adult cholangiocytes for organoid formation and liver regeneration. *Nat Cell Biol*. 2019 Nov;21(11):1321-1333.

Ostermann L, Ladewig J, Müller FJ, Kesavan J, Tailor J, **Smith A**, Brüstle O, Koch P. In Vitro Recapitulation of Developmental Transitions in Human Neural Stem Cells. *Stem Cells*. 2019 Nov;37(11):1429-1440.

Gu M, Zwiebel M, Ong SH, Boughton N, Nomdedeu J, Basheer F, Nannya Y, Quiros PM, Ogawa S, Cazzola M, Rad R, Butler AP, Vijayabaskar MS, **Vassiliou G**. RNAmut: robust identification of somatic mutations in acute myeloid leukemia using RNA-seq. *Haematologica*. 2019 Oct 24. pii: haematol.2019.230821.

Méndez-Ferrer S. Molecular interactome between HSCs and their niches. *Blood*. 2019 Oct 10;134(15):1197-1198.

Caesar R, Di Re M, Krupka JA, Gao J, Lara-Chica M, Dias JML, Cooke SL, Fenner R, Usheva Z, Runge HFP, Beer PA, Eldaly H, Pak HK, Park CS, **Vassiliou GS**, **Huntly BJP**, Mupo A, Bashford-Rogers RJM, **Hodson DJ**. Genetic modification of primary human B cells to model high-grade lymphoma. *Nat Commun*. 2019 Oct 4;10(1):4543.

Nobuta H, Yang N, Ng YH, Marro SG, Sabeur K, Chavali M, Stockley JH, Killilea DW, Walter PB, Zhao C, Huie P Jr, Goldman SA, Kriegstein AR, **Franklin RJM**, **Rowitch DH**, Wernig M. Oligodendrocyte Death in Pelizaeus-Merzbacher Disease Is Rescued by Iron Chelation. *Cell Stem Cell*. 2019 Oct 3;25(4):531-541.e6.

Neumann B, Baror R, Zhao C, Segel M, Dietmann S, Rawji KS, Foerster S, McClain CR, **Chalut K**, van Wijngaarden P, **Franklin RJM**. Metformin Restores CNS Remyelination Capacity by Rejuvenating Aged Stem Cells. *Cell Stem Cell*. 2019 Oct 3;25(4):473-485.e8.

Harris KL, Armstrong M, Swain R, Erzinclioglu S, Das T, Burgess N, **Barker RA**, Mason SL. Huntington's disease patients display progressive deficits in hippocampal-dependent cognition during a task of spatial memory. *Cortex*. 2019 Oct;119:417-427.

Hannezo E, **Simons BD**. Multiscale dynamics of branching morphogenesis. *Curr Opin Cell Biol*. 2019 Oct;60:99-105.

Hamey FK, **Göttgens B**. Machine learning predicts putative hematopoietic stem cells within large single-cell transcriptomics data sets. *Exp Hematol*. 2019 Oct;78:11-20.

Foerster S, Hill MFE, **Franklin RJM**. Diversity in the oligodendrocyte lineage: Plasticity or heterogeneity? *Glia*. 2019 Oct;67(10):1797-1805.

Forte D, Krause DS, Andreeff M, Bonnet D, **Méndez-Ferrer S**. Updates on the hematologic tumor microenvironment and its therapeutic targeting. *Haematologica*. 2019 Oct;104(10):1928-1934.

Stott SR, Randle SJ, Al Rawi S, Rowicka PA, Harris R, Mason B, Xia J, Dalley JW, **Barker RA**, Laman H. Loss of FBXO7 results in a Parkinson's-like dopaminergic degeneration via an RPL23-MDM2-TP53 pathway. *J Pathol*. 2019 Oct;249(2):241-254.

Popescu DM, Botting RA, Stephenson E, Green K, Webb S, Jardine L, Calderbank EF, Polanski K, Goh I, Efremova M, Acres M, Maunder D, Vegh P, Gitton Y, Park JE, Vento-Tormo R, Miao Z, Dixon D, Rowell R, McDonald D, Fletcher J, Poyner E, Reynolds G, Mather M, Moldovan C, Mamanova L, Greig F, Young MD, Meyer KB, Lisgo S, Bacardit J, Fuller A, Millar B, Innes B, Lindsay S, Stubbington MJT, Kowalczyk MS, Li B, Ashenberg O, Tabaka M, Dionne D, Tickle TL, Slyper M, Rozenblatt-Rosen O, Filby A, Carey P, Villani AC, Roy A, Regev A, Chédotal A, Roberts I, **Göttgens B**, Behjati S, **Laurenti E**, Teichmann SA, Haniiffa M. Decoding human fetal liver haematopoiesis. *Nature*. 2019 Oct;574(7778):365-371.

Waqar M, Davies BM, Zakaria R, Barone DG, Koliass AG, Yu KK, Ellenbogen JR, Brennan PM, **Kotter MR**, Gray WP, King AT, Turner C, Kitchen N, May P, Jenkinson MD, Hutchinson PJ. Academic neurosurgery in the UK: present and future directions. *Postgrad Med J*. 2019 Oct;95(1128):524-530.

Tanase CE, Qutachi O, White LJ, Shakesheff KM, **McCaskie AW**, Best SM, Cameron RE. Targeted protein delivery: carbodiimide crosslinking influences protein release from microparticles incorporated within collagen scaffolds. *Regen Biomater*. 2019 Oct;6(5):279-287.

Castaño J, Aranda S, Bueno C, Calero-Nieto FJ, Mejia-Ramirez E, Mosquera JL, Blanco E, Wang X, Prieto C, Zabaleta L, Mereu E, Rovira M, Jiménez-Delgado S, Matson DR, Heyn H, Bresnick EH, **Göttgens B**, Di Croce L, Menendez P, Raya A, Giorgetti A. GATA2 Promotes Hematopoietic Development and Represses Cardiac Differentiation of Human Mesoderm. *Stem Cell Reports*. 2019 Sep 10;13(3):515-529.

Han S, Fink J, Jörg DJ, Lee E, Yum MK, Chatzeli L, Merker SR, Jossierand M, Trendafilova T, Andersson-Rolf A, Dabrowska C, Kim H, Naumann R, Lee JH, Sasaki N, Mort RL, Basak O, Clevers H, Stange DE, **Philpott A**, Kim JK, **Simons BD**, Koo BK. Defining the Identity and Dynamics of Adult Gastric Isthmus Stem Cells. *Cell Stem Cell*. 2019 Sep 5;25(3):342-356.e7.

Stuart HT, Stirparo GG, Lohoff T, Bates LE, Kinoshita M, Lim CY, Sousa EJ, Maskalenka K, Radziskeuskaya A, Malcolm AA, Alves MRP, Lloyd RL, Nestorowa S, Humphreys P, Mansfield W, Reik W, Bertone P, **Nichols J, Göttgens B, Silva JCR**. Distinct Molecular Trajectories Converge to Induce Naive Pluripotency. *Cell Stem Cell*. 2019 Sep 5;25(3):388-406.e8.

Ho YH, Del Toro R, Rivera-Torres J, Rak J, Korn C, García-García A, Macías D, González-Gómez C, Del Monte A, Wittner M, Waller AK, Foster HR, López-Otín C, Johnson RS, Nerlov C, **Ghevaert C**, Vainchenker W, Louache F, Andrés V, **Méndez-Ferrer S**. Remodeling of Bone Marrow Hematopoietic Stem Cell Niches Promotes Myeloid Cell Expansion during Premature or Physiological Aging. *Cell Stem Cell*. 2019 Sep 5;25(3):407-418.e6.

Huang M, Taylor J, Zhen Q, Gillmor AH, Miller ML, Weishaupt H, Chen J, Zheng T, Nash EK, McHenry LK, An Z, Ye F, Takashima Y, Clarke J, Ayetey H, Cavalli FMG, Luu B, Moriarity BS, Ilkhanizadeh S, Chavez L, Yu C, Kurian KM, Magnaldo T, Sevenet N, Koch P, Pollard SM, Dirks P, Snyder MP, Largaespada DA, Cho YJ, Phillips JJ, Swartling FJ, Morrissy AS, Kool M, Pfister SM, Taylor MD, **Smith A**, Weiss WA. Engineering Genetic Predisposition in Human Neuroepithelial Stem Cells Recapitulates Medulloblastoma Tumorigenesis. *Cell Stem Cell*. 2019 Sep 5;25(3):433-446.e7.

in X, Wanga S, Fellows AL, Barallobre-Barreiro J, Lu R, Davaapil H, Franken R, Fava M, Baig F, Skroblin P, Xing Q, Koolbergen DR, Groenink M, Zwinderman AH, Balm R, de Vries CJM, Mulder BJM, Viner R, Jahangiri M, Reinhardt DP, **Sinha S**, de Waard V, Mayr M. Glycoproteomic Analysis of the Aortic Extracellular Matrix in Marfan Patients. *Arterioscler Thromb Vasc Biol*. 2019 Sep;39(9):1859-1873.

Segel M, Neumann B, Hill MFE, Weber IP, Viscomi C, Zhao C, Young A, Agle CC, Thompson AJ, Gonzalez GA, Sharma A, Holmqvist S, **Rowitch DH**, Franze K, **Franklin RJM**, **Chalut KJ**. Niche stiffness underlies the ageing of central nervous system progenitor cells. *Nature*. 2019 Sep;573(7772):130-134.

Schirmer L, Velmshch D, Holmqvist S, Kaufmann M, Werneburg S, Jung D, Vistnes S, Stockley JH, Young A, Steindel M, Tung B, Goyal N, Bhaduri A, Mayer S, Engler JB, Bayraktar OA, **Franklin RJM**, Haeussler M, Reynolds R, Schafer DP, Friese MA, Shiow LR, Kriegstein AR, **Rowitch DH**. Neuronal vulnerability and multilineage diversity in multiple sclerosis. *Nature*. 2019 Sep;573(7772):75-82.

Woo AJ, Patry CA, Ghamari A, Pregernig G, Yuan D, Zheng K, Piers T, Hibbs M, Li J, Fidalgo M, Wang JY, **Lee JH**, Leedman PJ, Wang J, Fraenkel E, Cantor AB. Zfp281 (ZBP-99) plays a functionally redundant role with Zfp148 (ZBP-89) during erythroid development. *Blood Adv*. 2019 Aug 27;3(16):2499-2511.

Nakanoh S, Agata K. Evolutionary view of pluripotency seen from early development of non-mammalian amniotes. *Dev Biol*. 2019 Aug 15;452(2):95-103.

Gupta N, Henry RG, Kang SM, Strober J, Lim DA, Ryan T, Perry R, Farrell J, Ulman M, Rajalingam R, Gage A, Huhn SL, Barkovich AJ, **Rowitch DH**. Long-Term Safety, Immunologic Response, and Imaging Outcomes following Neural Stem Cell Transplantation for Pelizaeus-Merzbacher Disease. *Stem Cell Reports*. 2019 Aug 13;13(2):254-261.

Sugarman J, **Barker RA**, Charo RA. A Professional Standard for Informed Consent for Stem Cell Therapies. *JAMA*. 2019 Aug 12. doi: 10.1001/jama.2019.11290.

Li Z, Solomonidis EG, Meloni M, Taylor RS, Duffin R, Dobie R, Magalhaes MS, Henderson BEP, Louwe PA, D'Amico G, Hodivala-Dilke KM, Shah AM, Mills NL, **Simons BD**, Gray GA, Henderson NC, Baker AH, Brittan M. Single-cell transcriptome analyses reveal novel targets modulating cardiac neovascularization by resident endothelial cells following myocardial infarction. *Eur Heart J*. 2019 Aug 7;40(30):2507-2520.

Wilkinson AC, Ryan DJ, Kucinski I, Wang W, Yang J, Nestorowa S, Diamanti E, Tsang JC, Wang J, Campos LS, Yang F, Fu B, Wilson N, Liu P, **Gottgens B**. Expanded potential stem cell media as a tool to study human developmental hematopoiesis in vitro. *Exp Hematol*. 2019 Aug;76:1-12.e5.

Rivera FJ, de la Fuente AG, Zhao C, Silva ME, Gonzalez GA, Wodnar R, Feichtner M, Lange S, Errea O, Priglinger E, O'Sullivan A, Romanelli P, Jadasz JJ, Brachtl G, Greil R, Tempfer H, Traweger A, Bätz LF, Küry P, Couillard-Despres S, **Franklin RJM**, Aigner L. Aging restricts the ability of mesenchymal stem cells to promote the generation of oligodendrocytes during remyelination. *Glia*. 2019 Aug;67(8):1510-1525.

Bargehr J, Ong LP, Colzani M, Davaapil H, Hofsteen P, Bhandari S, Gambardella L, Le Novère N, Iyer D, Sampaziotis F, Weinberger F, Bertero A, Leonard A, Bernard WG, Martinson A, Figg N, Regnier M, Bennett MR, Murry CE, **Sinha S**. Epicardial cells derived from human embryonic stem cells augment cardiomyocyte-driven heart regeneration. *Nat Biotechnol*. 2019 Aug;37(8):895-906.

Andersen MS, Hannezo E, Ulyanchenko S, Estrach S, Antoku Y, Pisano S, Boonekamp KE, Sendrup S, Maimets M, Pedersen MT, Johansen JV, Clement DL, Feral CC, **Simons BD**, Jensen KB. Tracing the cellular dynamics of sebaceous gland development in normal and perturbed states. *Nat Cell Biol*. 2019 Aug;21(8):924-932.

Aldape K, Brindle KM, Chesler L, Chopra R, Gajjar A, Gilbert MR, Gottardo N, Gutmann DH, Hargrave D, Holland EC, Jones DTW, Joyce JA, Kearns P, Kieran MW, Mellinghoff IK, Merchant M, Pfister SM, Pollard SM, Ramaswamy V, Rich JN, Robinson GW, **Rowitch DH**, Sampson JH, Taylor MD, Workman P, Gilbertson RJ. Challenges to curing primary brain tumours. *Nat Rev Clin Oncol*. 2019 Aug;16(8):509-520.

Ombrato L, Nolan E, Kurelac I, Mavousian A, Bridgeman VL, Heinze I, Chakravarty P, Horswell S, Gonzalez-Gualda E, Matacchione G, Weston A, Kirkpatrick J, Husain E, Speirs V, Collinson L, Ori A, **Lee JH**, Malanchi I. Metastatic-niche labelling reveals parenchymal cells with stem features. *Nature*. 2019 Aug;572(7771):603-608.

Rudorf A, Müller TA, Klingeberg C, Kreutmair S, Poggio T, Gorantla SP, Rückert T, Schmitt-Graeff A, Gengenbacher A, Paschka P, Baldus C, Zeiser R, **Vassiliou GS**, Bradley A, Duyster J, Illert AL. NPM1c alters FLT3-D835Y localization and signaling in acute myeloid leukemia. *Blood*. 2019 Jul 25;134(4):383-388.

Tempest-Mitchell J, Hilton B, Davies BM, Nouri A, Hutchinson PJ, Scoffings DJ, Mannion RJ, Trivedi R, Timofeev I, Crawford JR, Hay D, Laing RJ, **Kotter MRN**. A comparison of radiological descriptions of spinal cord compression with quantitative measures, and their role in non-specialist clinical management. *PLoS One*. 2019 Jul 22;14(7):e0219380.

Vassiliou GS. Triple-mutant AML: too clever by HLF? *Blood*. 2019 Jul 18;134(3):222-224.

Nakagaki-Silva EE, Gooding C, Llorian M, Jacob AG, Richards F, Buckroyd A, **Sinha S**, Smith CW. Identification of RBPMS as a mammalian smooth muscle master splicing regulator via proximity of its gene with super-enhancers. *Elife*. 2019 Jul 8;8. pii: e46327.

Buczacki S. Fate plasticity in the intestine: The devil is in the detail. *World J Gastroenterol*. 2019 Jul 7;25(25):3116-3122.

Ranzoni AM, Strzelecka PM, **Cvejic A**. Application of single-cell RNA sequencing methodologies in understanding haematopoiesis and immunology. *Essays Biochem*. 2019 Jul 3;63(2):217-225.

Hardwick LJA, **Philpott A**. Analysis of Chromatin Binding of Ectopically Expressed Proteins in Early *Xenopus* Embryos. *Cold Spring Harb Protoc*. 2019 Jul 1;2019(7).

Hardwick LJA, **Philpott A**. Analysis of Phosphorylation Status of Ectopically Expressed Proteins in Early *Xenopus* Embryos. *Cold Spring Harb Protoc*. 2019 Jul 1;2019(7).

Shepherd MS, **Kent DG**. Emerging single-cell tools are primed to reveal functional and molecular heterogeneity in malignant hematopoietic stem cells. *Curr Opin Hematol*. 2019 Jul;26(4):214-221.

Baror R, Neumann B, Segel M, **Chalut KJ**, Fancy SPJ, Schafer DP, **Franklin RJM**. Transforming growth factor-beta renders ageing microglia inhibitory to oligodendrocyte generation by CNS progenitors. *Glia*. 2019 Jul;67(7):1374-1384.

Barker RA; TRANSEURO consortium. Designing stem-cell-based dopamine cell replacement trials for Parkinson's disease. *Nat Med*. 2019 Jul;25(7):1045-1053.

Vieira Braga FA, Kar G, Berg M, Carpaij OA, Polanski K, Simon LM, Brouwer S, Gomes T, Hesse L, Jiang J, Fasouli ES, Efremova M, Vento-Tormo R, Talavera-López C, Jonker MR, Affleck K, Palit S, Strzelecka PM, Firth HV, Mahbubani KT, **Cvejic A**, Meyer KB, Saeb-Parsy K, Luinge M, Brandsma CA, Timens W, Angelidis I, Strunz M, Koppelman GH, van Oosterhout AJ, Schiller HB, Theis FJ, van den Berge M, Nawijn MC, Teichmann SA. A cellular census of human lungs identifies novel cell states in health and in asthma. *Nat Med*. 2019 Jul;25(7):1153-1163.

Ng NHJ, Jasmen JB, Lim CS, Lau HH, Krishnan VG, Kadiwala J, Kulkarni RN, Ræder H, **Vallier L**, Hoon S, Teo AKK. HNF4A Haploinsufficiency in MODY1 Abrogates Liver and Pancreas Differentiation from Patient-Derived Induced Pluripotent Stem Cells. *iScience*. 2019 Jun 28;16:192-205.

- Edri S, Hayward P, Jawaid W, Martinez Arias A. Neuro-mesodermal progenitors (NMPs): a comparative study between pluripotent stem cells and embryo-derived populations. *Development*. 2019 Jun 24;146(12). pii: dev180190.
- Nguyen CH, Glüxam T, Schlerka A, Bauer K, Grandits AM, Hackl H, Dovey O, Zöchbauer-Müller S, Cooper JL, **Vassiliou GS**, Stoiber D, Wieser R, Heller G. SOCS2 is part of a highly prognostic 4-gene signature in AML and promotes disease aggressiveness. *Sci Rep*. 2019 Jun 24;9(1):9139.
- Burgold T, Barber M, Kloet S, Cramard J, Gharbi S, Floyd R, Kinoshita M, Ralsler M, Vermeulen M, Reynolds N, Dietmann S, **Hendrich B**. The Nucleosome Remodelling and Deacetylation complex suppresses transcriptional noise during lineage commitment. *EMBO J*. 2019 Jun 17;38(12).
- Gkatza NA, Castro C, Harvey RF, Heiß M, Popis MC, Blanco S, Bornelöv S, Sajini AA, Gleeson JG, Griffin JL, West JA, Kellner S, Willis AE, Dietmann S, **Frye M**. Cytosine-5 RNA methylation links protein synthesis to cell metabolism. *PLoS Biol*. 2019 Jun 14;17(6):e3000297.
- Prior N, Hindley CJ, Rost F, Meléndez E, Lau WWY, **Göttgens B**, Rulands S, **Simons BD**, Huch M. Lgr5(+) stem and progenitor cells reside at the apex of a heterogeneous embryonic hepatoblast pool. *Development*. 2019 Jun 12;146(12).
- Sajini AA, Choudhury NR, Wagner RE, Bornelöv S, Selmi T, Spanos C, Dietmann S, Rappsilber J, Michlewski G, **Frye M**. Loss of 5-methylcytosine alters the biogenesis of vault-derived small RNAs to coordinate epidermal differentiation. *Nat Commun*. 2019 Jun 11;10(1):2550.
- Jia W, Kamen Y, Pivonkova H, **Káradóttir RT**. Neuronal activity-dependent myelin repair after stroke. *Neurosci Lett*. 2019 Jun 11;703:139-144.
- Lu Y, Stuart JH, Talbot-Cooper C, Agrawal-Singh S, **Huntly B**, Smid AI, Snowden JS, Dupont L, Smith GL. Histone deacetylase 4 promotes type I interferon signaling, restricts DNA viruses, and is degraded via vaccinia virus protein C6. *Proc Natl Acad Sci U S A*. 2019 Jun 11;116(24):11997-12006.
- Bredenkamp N, Stirparo GG, **Nichols J, Smith A**, Guo G. The Cell-Surface Marker Sushi Containing Domain 2 Facilitates Establishment of Human Naive Pluripotent Stem Cells. *Stem Cell Reports*. 2019 Jun 11;12(6):1212-1222.
- Bornelöv S, Selmi T, Flad S, Dietmann S, **Frye M**. Codon usage optimization in pluripotent embryonic stem cells. *Genome Biol*. 2019 Jun 7;20(1):119.
- Clevers H, Conder RK, Li VSW, Lutolf MP, **Vallier L**, Chan S, Grikscheit TC, Jensen KB, De Coppi P. Tissue-Engineering the Intestine: The Trials before the Trials. *Cell Stem Cell*. 2019 Jun 6;24(6):855-859.
- Lawrence M, Theunissen TW, Lombard P, Adams DJ, **Silva JCR**. ZMYM2 inhibits NANOG-mediated reprogramming. *Wellcome Open Res*. 2019 Jun 6;4:88.
- Horton S, **Huntly BJP**. Kinase Networks Regulate Metabolism: I'D(H1) Never Have Guessed! *Cancer Discov*. 2019 Jun;9(6):699-701.
- Günther C, Brevini T, Sampaziotis F, Neurath MF. What gastroenterologists and hepatologists should know about organoids in 2019. *Dig Liver Dis*. 2019 Jun;51(6):753-760.
- Bueno C, Calero-Nieto FJ, Wang X, Valdés-Mas R, Gutiérrez-Agüera F, Roca-Ho H, Ayllon V, Real PJ, Arambilet D, Espinosa L, Torres-Ruiz R, Agraz-Doblas A, Varela I, de Boer J, Bigas A, **Gottgens B**, Marschalek R, Menendez P. Enhanced hemato-endothelial specification during human embryonic differentiation through developmental cooperation between AF4-MLL and MLL-AF4 fusions. *Haematologica*. 2019 Jun;104(6):1189-1201.
- Tysoe OC, Justin AW, Brevini T, Chen SE, Mahbubani KT, Frank AK, Zedira H, Melum E, Saeb-Parsy K, Markaki AE, **Vallier L**, Sampaziotis F. Isolation and propagation of primary human cholangiocyte organoids for the generation of bioengineered biliary tissue. *Nat Protoc*. 2019 Jun;14(6):1884-1925.
- Guiu J, Hannezo E, Yui S, Demharter S, Ulyanchenko S, Maimets M, Jørgensen A, Perlman S, Lundvall L, Mamsen LS, Larsen A, Olesen RH, Andersen CY, Thuesen LL, Hare KJ, Pers TH, Khodosevich K, **Simons BD**, Jensen KB. Tracing the origin of adult intestinal stem cells. *Nature*. 2019 Jun;570(7759):107-111.
- Hodson DJ**, Screen M, Turner M. RNA-binding proteins in hematopoiesis and hematological malignancy. *Blood*. 2019 May 30;133(22):2365-2373.

De Mattos-Arruda L, Sammut SJ, Ross EM, Bashford-Rogers R, Greenstein E, Markus H, Morganella S, Teng Y, Maruvka Y, Pereira B, Rueda OM, Chin SF, Contente-Cuomo T, Mayor R, Arias A, Ali HR, Cope W, Tiezzi D, Dariush A, Dias Amarante T, Reshef D, Ciriaco N, Martinez-Saez E, Peg V, Ramon Y Cajal S, Cortes J, **Vassiliou G**, Getz G, Nik-Zainal S, Murtaza M, Friedman N, Markowitz F, Seoane J, Caldas C. The Genomic and Immune Landscapes of Lethal Metastatic Breast Cancer. *Cell Rep*. 2019 May 28;27(9):2690-2708. e10.

Velmeshev D, Schirmer L, Jung D, Haeussler M, Perez Y, Mayer S, Bhaduri A, Goyal N, **Rowitch DH**, Kriegstein AR. Single-cell genomics identifies cell type-specific molecular changes in autism. *Science*. 2019 May 17;364(6441):685-689.

Aztekin C, Hiscock TW, Marioni JC, Gurdon JB, **Simons BD**, Jullien J. Identification of a regeneration-organizing cell in the *Xenopus* tail. *Science*. 2019 May 17;364(6441):653-658.

Filipponi D, Emelyanov A, Muller J, Molina C, **Nichols J**, Bulavin DV. DNA Damage Signaling-Induced Cancer Cell Reprogramming as a Driver of Tumor Relapse. *Mol Cell*. 2019 May 16;74(4):651-663.e8.

Kalkan T, Bornelöv S, Mulas C, Diamanti E, Lohoff T, Ralsler M, Middelkamp S, Lombard P, **Nichols J**, **Smith A**. Complementary Activity of ETV5, RBPJ, and TCF3 Drives Formative Transition from Naïve Pluripotency. *Cell Stem Cell*. 2019 May 2;24(5):785-801.e7.

Hoffman M, Gillmor AH, Kunz DJ, Johnston MJ, Nikolic A, Narta K, Zarrei M, King J, Ellestad K, Dang NH, Cavalli FMG, Kushida MM, Coutinho FJ, Zhu Y, Luu B, Ma Y, Mungall AJ, Moore R, Marra MA, Taylor MD, Pugh TJ, Dirks PB, Strother D, Lafay-Cousin L, Resnick AC, Scherer S, Senger DL, **Simons BD**, Chan JA, Morrissy AS, Gallo M. Intratumoral Genetic and Functional Heterogeneity in Pediatric Glioblastoma. *Cancer Res*. 2019 May 1;79(9):2111-2123.

Davies BM, Khan DZ, Mowforth OD, McNair AGK, Gronlund T, Koliaf AG, Tetreault L, Starkey ML, Sadler I, Sarewitz E, Houlton D, Carter J, Kalsi-Ryan S, Aarabi B, Kwon BK, Kurpad SN, Harrop J, Wilson JR, Grossman R, Curt A, Fehlings MG, **Kotter MRN**. RE-CODE DCM (REsearch Objectives and Common Data Elements for Degenerative Cervical Myelopathy): A Consensus Process to Improve Research Efficiency in DCM, Through Establishment of a Standardized Dataset for Clinical Research and the Definition of the Research Priorities. *Global Spine J*. 2019 May;9(1 Suppl):655-76S.

French CE, Delon I, Dolling H, Sanchis-Juan A, Shamardina O, Mégy K, Abbs S, Austin T, Bowdin S, Branco RG, Firth H; NIH BioResource—Rare Disease; Next Generation Children Project, **Rowitch DH**, Raymond FL. Whole genome sequencing reveals that genetic conditions are frequent in intensively ill children. *Intensive Care Med*. 2019 May;45(5):627-636.

Waller AK, Lantos L, Sammut A, Salgin B, McKinney H, Foster HR, Kriek N, Gibbins JM, Stanworth SJ, Garner SF, Venkatesh V, Curley A, Belteki G, **Ghevaert C**. Flow cytometry for near-patient testing in premature neonates reveals variation in platelet function: a novel approach to guide platelet transfusion. *Pediatr Res*. 2019 May;85(6):874-884.

Kalna V, Yang Y, Peghaire CR, Frudd K, Hannah R, Shah AV, Osuna Almagro L, Boyle JJ, **Göttgens B**, Ferrer J, Randi AM, Birdsey GM. The Transcription Factor ERG Regulates Super-Enhancers Associated With an Endothelial-Specific Gene Expression Program. *Circ Res*. 2019 Apr 26;124(9):1337-1349.

Teo YV, Rattanavirotkul N, Olova N, Salzano A, Quintanilla A, Tarrats N, Kiourtis C, Müller M, **Green AR**, Adams PD, Acosta JC, Bird TG, Kirschner K, Neretti N, Chandra T. Notch Signaling Mediates Secondary Senescence. *Cell Rep*. 2019 Apr 23;27(4):997-1007.e5.

Leeman KT, Pessina P, **Lee JH**, Kim CF. Mesenchymal Stem Cells Increase Alveolar Differentiation in Lung Progenitor Organoid Cultures. *Sci Rep*. 2019 Apr 23;9(1):6479.

Mowforth OD, Davies BM, **Kotter MR**. "I am not delusional!" Sensory dysaesthesia secondary to degenerative cervical myelopathy. *BMJ Case Rep*. 2019 Apr 11;12(4). pii: e229033.

Peñalosa-Ruiz G, Bousgouni V, Gerlach JP, Waarlo S, van de Ven JV, Veenstra TE, **Silva JCR**, van Heeringen SJ, Bakal C, Mulder KW, Veenstra GJC. WDR5, BRCA1, and BARD1 Co-regulate the DNA Damage Response and Modulate the Mesenchymal-to-Epithelial Transition during Early Reprogramming. *Stem Cell Reports*. 2019 Apr 9;12(4):743-756.

Rostovskaya M, Stirparo GG, **Smith A**. Capacitation of human naïve pluripotent stem cells for multi-lineage differentiation. *Development*. 2019 Apr 3;146(7). pii: dev172916.

Wiese M, Bannister AJ, **Basu S**, Boucher W, Wohlfahrt K, Christophorou MA, Nielsen ML, Klenerman D, Laue ED, Kouzarides T. Citrullination of HP1 γ chromodomain affects association with chromatin. *Epigenetics Chromatin*. 2019 Apr 2;12(1):21.

Kaiser K, Gyllborg D, Procházka J, Salašová A, Kompaníková P, Molina FL, Laguna-Goya R, Radaszkiewicz T, Harnoš J, Procházková M, Potěšil D, **Barker RA**, Casado ÁG, Zdráhal Z, Sedláček R, Arenas E, Villaescusa JC, Bryja V. WNT5A is transported via lipoprotein particles in the cerebrospinal fluid to regulate hindbrain morphogenesis. *Nat Commun.* 2019 Apr 2;10(1):1498.

Glass HC, Ferriero DM, **Rowitch DH**, Shimotake TK. The neurointensive nursery: concept, development, and insights gained. *Curr Opin Pediatr.* 2019 Apr;31(2):202-209.

Bernstock JD, Peruzzotti-Jametti L, Leonardi T, Vicario N, Ye D, Lee YJ, Maric D, Johnson KR, Mou Y, Van Den Bosch A, Winterbone M, Friedman GK, **Franklin RJM**, Hallenbeck JM, Pluchino S. SUMOylation promotes survival and integration of neural stem cell grafts in ischemic stroke. *EBioMedicine.* 2019 Apr;42:214-224.

Sainz de Aja J, Menchero S, Rollan I, Barral A, Tiana M, Jawaid W, Cossio I, Alvarez A, Carreño-Tarragona G, Badia-Careaga C, **Nichols J, Göttgens B**, Isern J, Manzanares M. The pluripotency factor NANOG controls primitive hematopoiesis and directly regulates Tal1. *EMBO J.* 2019 Apr 1;38(7). pii: e99122.

Rasmussen KD, Berest I, Keßler S, Nishimura K, Simón-Carrasco L, **Vassiliou GS**, Pedersen MT, Christensen J, Zaugg JB, Helin K. TET2 binding to enhancers facilitates transcription factor recruitment in hematopoietic cells. *Genome Res.* 2019 Apr;29(4):564-575.

Basheer F, Giotopoulos G, Meduri E, Yun H, Mazan M, Sasca D, Gallipoli P, Marando L, Gozdecka M, Asby R, Sheppard O, Dudek M, Bullinger L, Döhner H, Dillon R, Freeman S, Ottmann O, Burnett A, Russell N, Papaemmanuil E, Hills R, Campbell P, **Vassiliou GS, Huntly BJP**. Contrasting requirements during disease evolution identify EZH2 as a therapeutic target in AML. *J Exp Med.* 2019 Apr 1;216(4):966-981.

Weber J, de la Rosa J, Grove CS, Schick M, Rad L, Baranov O, Strong A, Pfau A, Friedrich MJ, Engleitner T, Lersch R, Öllinger R, Grau M, Menendez IG, Martella M, Kohlhofer U, Banerjee R, Turchaninova MA, Scherger A, Hoffman GJ, Hess J, Kuhn LB, Ammon T, Kim J, Schneider G, Unger K, Zimmer-Strobl U, Heikenwälder M, Schmidt-Supprian M, Yang F, Saur D, Liu P, Steiger K, Chudakov DM, Lenz G, Quintanilla-Martinez L, Keller U, **Vassiliou GS**, Cadiñanos J, Bradley A, Rad R. PiggyBac transposon tools for recessive screening identify B-cell lymphoma drivers in mice. *Nat Commun.* 2019 Mar 29;10(1):1415.

Watcham S, Kucinski I, **Gottgens B**. New insights into hematopoietic differentiation landscapes from single-cell RNA sequencing. *Blood.* 2019 Mar 28;133(13):1415-1426.

Nangalia J, Mitchell E, **Green AR**. Clonal approaches to understanding the impact of mutations on hematologic disease development. *Blood.* 2019 Mar 28;133(13):1436-1445.

Silva ME, Lange S, Hinrichsen B, Philp AR, Reyes CR, Halabi D, Mansilla JB, Rotheneichner P, Guzman de la Fuente A, Couillard-Despres S, Bätz LF, **Franklin RJM**, Aigner L, Rivera FJ. Pericytes Favor Oligodendrocyte Fate Choice in Adult Neural Stem Cells. *Front Cell Neurosci.* 2019 Mar 27;13:85.

Hardwick LJA, Davies JD, **Philpott A**. Multi-site phosphorylation controls the neurogenic and myogenic activity of E47. *Biochem Biophys Res Commun.* 2019 Mar 26;511(1):111-116.

Mulas C, Kalkan T, von Meyenn F, Leitch HG, **Nichols J, Smith A**. Defined conditions for propagation and manipulation of mouse embryonic stem cells. *Development.* 2019 Mar 26;146(6). pii: dev173146.

Wolf FA, Hamey FK, Plass M, Solana J, Dahlin JS, **Göttgens B**, Rajewsky N, Simon L, Theis FJ. PAGA: graph abstraction reconciles clustering with trajectory inference through a topology preserving map of single cells. *Genome Biol.* 2019 Mar 19;20(1):59.

Recho P, Hallou A, Hannezo E. Theory of mechanochemical patterning in biphasic biological tissues. *Proc Natl Acad Sci U S A.* 2019 Mar 19;116(12):5344-5349.

Gallipoli P, **Huntly BJP**. Histone modifiers are oxygen sensors. *Science.* 2019 Mar 15;363(6432):1148-1149. doi: 10.1126/science.aaw8373. PubMed PMID: 30872506.

Hurrell T, Segeritz CP, **Vallier L**, Lilley KS, Cromarty AD. A proteomic time course through the differentiation of human induced pluripotent stem cells into hepatocyte-like cells. *Sci Rep.* 2019 Mar 1;9(1):3270.

Jörg DJ, Caygill EE, Hakes AE, Contreras EG, Brand AH, **Simons BD**. The proneural wave in the *Drosophila* optic lobe is driven by an excitable reaction-diffusion mechanism. *Elife.* 2019 Feb 22;8. pii: e40919.

Fragniere AMC, Stott SRW, Fazal SV, Andreasen M, Scott K, **Barker RA**. Hyperosmotic stress induces cell-dependent aggregation of α -synuclein. *Sci Rep*. 2019 Feb 19;9(1):2288.

Linker SM, Urban L, Clark SJ, Chhatiwala M, Amatya S, McCarthy DJ, Ebersberger I, **Vallier L**, Reik W, Stegle O, Bonder MJ. Combined single-cell profiling of expression and DNA methylation reveals splicing regulation and heterogeneity. *Genome Biol*. 2019 Feb 11;20(1):30.

Palmer AJR, Ayyar Gupta V, Fernquest S, Rombach I, Dutton SJ, Mansour R, Wood S, Khanduja V, Pollard TCB, **McCaskie AW**, Barker KL, Andrade TJMD, Carr AJ, Beard DJ, Glyn-Jones S; FAIT Study Group. Arthroscopic hip surgery compared with physiotherapy and activity modification for the treatment of symptomatic femoroacetabular impingement: multicentre randomised controlled trial. *BMJ*. 2019 Feb 7;364:l185.

Spitzer SO, Sitnikov S, Kamen Y, Evans KA, Kronenberg-Versteeg D, Dietmann S, de Faria O Jr, Agathou S, **Káradóttir RT**. Oligodendrocyte Progenitor Cells Become Regionally Diverse and Heterogeneous with Age. *Neuron*. 2019 Feb 6;101(3):459-471.e5.

Basheer FT, **Vassiliou GS**. Genome-scale drop-out screens to identify cancer cell vulnerabilities in AML. *Curr Opin Genet Dev*. 2019 Feb;54:83-87.

Ramos D, Catita J, López-Luppo M, Valença A, Bonet A, Carretero A, Navarro M, Nacher V, **Mendez-Ferrer S**, Meseguer A, Casellas A, Mendes-Jorge L, Ruberte J. Vascular Interstitial Cells in Retinal Arteriolar Annuli Are Altered During Hypertension. *Invest Ophthalmol Vis Sci*. 2019 Feb 1;60(2):473-487.

Pijuan-Sala B, Griffiths JA, Guibentif C, Hiscock TW, Jawaid W, Calero-Nieto FJ, Mulas C, Ibarra-Soria X, Tyser RCV, Ho DLL, Reik W, Srinivas S, **Simons BD**, **Nichols J**, Marioni JC, **Göttgens B**. A single-cell molecular map of mouse gastrulation and early organogenesis. *Nature*. 2019 Feb;566(7745):490-495.

Verstreken CM, Labouesse C, Agle CC, **Chalut KJ**. Embryonic stem cells become mechanoresponsive upon exit from ground state of pluripotency. *Open Biol*. 2019 Jan 31;9(1):180203.

Revell C, Blumenfeld R, **Chalut KJ**. Force-based three-dimensional model predicts mechanical drivers of cell sorting. *Proc Biol Sci*. 2019 Jan 30;286(1895):20182495.

Davies BM, Munro CF, **Kotter MR**. A Novel Insight Into the Challenges of Diagnosing Degenerative Cervical Myelopathy Using Web-Based Symptom Checkers. *J Med Internet Res*. 2019 Jan 11;21(1):e10868.

Chia CY, Madrigal P, Denil SLIJ, Martinez I, Garcia-Bernardo J, El-Khairi R, Chhatiwala M, Shepherd MH, Hattersley AT, Dunn NR, **Vallier L**. GATA6 Cooperates with EOMES/SMAD2/3 to Deploy the Gene Regulatory Network Governing Human Definitive Endoderm and Pancreas Formation. *Stem Cell Reports*. 2019 Jan 8;12(1):57-70.

Zhang W, Chronis C, Chen X, Zhang H, Spalinskas R, Pardo M, Chen L, Wu G, Zhu Z, Yu Y, Yu L, Choudhary J, **Nichols J**, Parast MM, Greber B, Sahlén P, Plath K. The BAF and PRC2 Complex Subunits Dpf2 and Eed Antagonistically Converge on Tbx3 to Control ESC Differentiation. *Cell Stem Cell*. 2019 Jan 3;24(1):138-152.e8.

Sybirna A, Wong FCK, Surani MA. Genetic basis for primordial germ cells specification in mouse and human: Conserved and divergent roles of PRDM and SOX transcription factors. *Curr Top Dev Biol*. 2019;135:35-89.

Hamey FK, **Göttgens B**. Reconstructing Gene Regulatory Networks That Control Hematopoietic Commitment. *Methods Mol Biol*. 2019;1975:239-249.

Sampaziotis F, Tysoe O, Brevini T, **Vallier L**. Use of Biliary Organoids in Cholestasis Research. *Methods Mol Biol*. 2019;1981:373-382. doi: 10.1007/978-1-4939-9420-5_25. PubMed PMID: 31016668.

Snijders KE, Cooper JD, **Vallier L**, Bertero A. Conditional Gene Knockout in Human Cells with Inducible CRISPR/Cas9. *Methods Mol Biol*. 2019;1961:185-209.

McMurrin CE, Zhao C, **Franklin RJM**. Toxin-Based Models to Investigate Demyelination and Remyelination. *Methods Mol Biol*. 2019;1936:377-396.

Agathou S, **Káradóttir RT**. Whole-Cell Patch Clamp Recordings from Oligodendrocyte Lineage Cells in Brain Slices. *Methods Mol Biol*. 2019;1936:141-168.

Penfold CA, Gherman I, Sybirna A, Wild DL. Inferring Gene Regulatory Networks from Multiple Datasets. *Methods Mol Biol*. 2019;1883:251-282.



